

Adsorption of Purified Cellulases on Cotton Fibers

ANAND PURUSHOTTAM KANCHAGAR

(Under the direction of Dr. J. Nolan Eppers)

Commercial cellulases are a mixture of endoglucanases (EGs), exoglucanases (CBHs) and glucosidases. EGs and CBHs differ in their action on cellulose as a virtue of their structure but are adsorbed on cellulose to conduct hydrolysis. The nature of cellulase adsorption on cellulose has been explained by adsorption models such as Langmuir model in the past.

In the present investigation, Linear, Langmuir, Freundlich and combined Langmuir-Freundlich adsorption models were used to explain the adsorption of CBH I, II and EG II. CBH I, II and EG II were purified and adsorption of the purified enzymes on cotton fibers was collected. Statistical analysis was performed to determine the best fitting model.

It was found that more than one adsorption model can be used to explain the adsorption of CBH I, II and EG II on cotton fibers in the given range of enzyme concentration indicating that the substrate is not saturated by the enzyme. The temperature of adsorption was found to affect

adsorption of CBH I, II and EG II on cotton fibers to a small extent. Adsorption of CBH I and II was found to be higher at 55 °C as compared to 25, 35 and 45 °C. Effect of temperature on adsorption of EG II on cotton fibers was more pronounced at higher concentrations of the enzyme.

INDEX WORDS: Cellulases, Endoglucanase, Exoglucanase,
Cotton, Adsorption, Purification, Adsorption
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ADSORPTION OF PURIFIED CELLULASES ON COTTON FIBERS

by

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CHAPTER 1

INTRODUCTION

The use of enzymes in industrial textile processing dates back to 1857, when malt extract was used to remove starch-based size from printed textiles. Since then, commercial application of enzymes in textile processing has been slow [1]. Not until the 1960's did detergents make their appearance in the consumer market. Proteases were added to detergents to remove organic protein-based stains. Later, in the 1970's cellulases were added to detergent formulations to remove fibrils during fabric washing [2]. In the last decade, the use of enzymes in textile processing increased rapidly. Now, enzymes are being used in textile processes such as scouring, bleaching and finishing of textile substrates.

Enzymes are protein molecules that function as biological catalysts. After being adsorbed to a specific substrate, enzymes accelerate reactions at the surface of the substrate under environmentally benign conditions of temperature and pH. As a result, enzymatic processing leads to savings of energy and water. Being protein molecules, enzymes are biodegradable and, as catalysts, are required in small quantities [3, 4, 5]. Thus,

the chemical oxygen demand (COD) and biological oxygen demand (BOD) of effluents from enzymatic treatments are low compared to chemical treatments [6, 7].

Among the different enzymes used in textile processing such as amylases, pectinases, cellulases, proteases and lipases, cellulases have been widely used in processing of cotton with great commercial success. Cellulases find applications such as additives in laundry detergents, defibrillation, bio-finishing of cotton fabrics and stone-washing of denim garments [8, 9, 10]. Commercially available cellulases are a mixture of three enzymes,

1. Endoglucanases (EG), also called "CMCase"
2. Exoglucanases (CBH), also called "Avicelase" or "Cellobiohydrolases"
3. Cellobiase [8,10]

Endoglucanases attack cellulose chains randomly along their lengths, acting on the amorphous regions of cellulose.

Exoglucanases are thought to act on the crystalline regions of cellulose, attacking the polymer chain ends, generating cellobiose. Cellobiose is further hydrolyzed to glucose by cellobiase. For complete hydrolysis of cellulose to glucose, all three enzymes are required [8, 10].

Cellulases are produced by fungi and bacteria. *Trichoderma*, *Aspergillus*, *Penicillium* and *Fusarium* are common sources of

fungal cellulases with *Trichoderma reesei* being the source of most commercially available cellulases [8]. Fungi may produce an array of endoglucanases and exoglucanases that may differ slightly in their action and topology. For example, cellulases from *Trichoderma reesei* contain two types of exoglucanases, CBH I and II, and six types of endoglucanases, EG I - VI. These different types of endoglucanases and exoglucanases differ in their structure and action on cellulose. It is difficult to break a large polymeric structure such as cellulose that is heterogeneous in its organization. Hence, multiple forms of endoglucanases and exoglucanases are required in order to bring about extensive hydrolysis of cellulose [7].

Recent studies on cellulase treatment of cotton fabrics are aimed at increasing its efficiency and effectiveness on cotton. Understanding adsorption characteristics of isolated endoglucanases and exoglucanases can throw light on the mechanism of their action on cotton fibers. Recycling and reuse of cellulase treatment baths can also be perfected with knowledge of adsorption characteristics of purified cellulases. Formulation of cellulase mixtures that can impart desired properties to cotton fabrics such as softness and removal of fuzz without substantial loss in weight and weakening of fabric can be possible with better understanding of the adsorption characteristics of purified cellulases.

Significance of Study

Cellulose is insoluble in water and hence the cellulose-cellulase system is heterogeneous system. Hydrolysis of cellulose by cellulase involves the following steps [11]:

1. Approach of enzyme to the substrate
2. Adsorption of enzyme on the substrate
3. Formation of an enzyme-substrate complex
4. Bond cleavage caused by cellulase and time course hydrolysis.

From the above steps listed involved in hydrolysis of cellulose by cellulase, it can be said that adsorption is a pre-requisite for cellulases to bring about cellulose hydrolysis. Adsorption of cellulases on the cellulose substrate is affected by a number of factors such as accessibility of the substrate, crystallinity and degree of polymerization [12, 13, 14, 15]. Cellulose substrates such as microcrystalline cellulose, pre-treated pulp and bagasse have been investigated for the action of complete cellulase systems and purified cellulases [16, 17, 18, 19, 20, 21, 22]. Microcrystalline cellulose is a commonly used substrate for such adsorption studies. Mathematical and empirical models have been developed to explain the kinetics of hydrolysis and understand the mechanism of hydrolysis. Since endoglucanases and exoglucanases differ in their action on microcrystalline cellulose, the composition of the enzyme

mixture plays an important role in determining its action on cellulose [17].

Cotton differs in its crystallinity and accessibility from other cellulose substrates such as paper pulp, bagasse and pretreated wood chips. As a result, adsorption of purified cellulases on cotton is expected to be different from the other substrates. But studies involving the adsorption of endoglucanase and exoglucanase on cotton are very few [23, 24, 25]. Adsorption studies on cellulosic substrates including cotton have been conducted at low temperatures such as 3 °C and 25 °C in order to avoid the influence of hydrolysis due to cellulase action. In practice, enzymatic processing of cotton is conducted in the temperature range of 45 - 55 °C [26]. Hence, understanding adsorption characteristics of cellulase and its components on cotton at such practical conditions of treatment is important in order to make enzymatic processing of cotton fabrics more effective and efficient.

Adsorption models such as the Langmuir isotherm and Freundlich isotherm have been successfully used to describe the adsorption of dyes to the surface of textile fibers, the dyeing mechanism, as well as the kinetics of dyeing. Since the adsorption of cellulases on cotton is a surface phenomenon, adsorption isotherms may have the potential to explain the adsorption of cellulases on cotton fibers. Studying the

adsorption characteristics of single enzymes may be important in optimizing the formulation and utilization of cellulase mixtures for treatment of cotton fabrics.

Purpose and Objectives

The purpose of this study was to determine the nature of adsorption of purified endoglucanases and exoglucanases on cotton fibers. Adsorption isotherms such as the Langmuir isotherm can provide a better understanding of the adsorption characteristics of endoglucanases and exoglucanases on cotton. Since adsorption isotherms describe the interaction of the enzyme with the substrate, isotherms can be useful in formulating mixtures of cellulases to impart specific properties to cotton fabrics. In order to conduct adsorption studies, endoglucanases and exoglucanases were purified from a commercial mixture of cellulases. Adsorption of increasing concentrations of purified endoglucanase and exoglucanases was assessed at different temperatures. Adsorption data was statistically analyzed performed to determine whether the data could be explained by different adsorption isotherms. Linear Regression analysis was conducted on the data using SAS[®] software.

The objective of this study was to determine whether the adsorption of endoglucanases and exoglucanases on cotton fibers

can be explained using linear, Langmuir, Freundlich or Combined Langmuir-Freundlich adsorption isotherms.

Limitations of Study

1. Substrate hydrolysis during the course of adsorption of enzymes is not taken into account
2. Structural features of the substrate such as surface area and crystallinity are not studied. Any changes in the structural features due to hydrolysis are not taken into account when analyzing the results
3. Enzyme inhibition by substrate and by products is assumed to be minimal
4. Results and conclusions are valid over the range of temperatures and concentrations studied in this project which reflect the practical range used in enzymatic processing of cotton.
5. The isolated enzymes used in the present study may not be 100 percent pure.

CHAPTER 2

LITERATURE REVIEW

The processing of cellulose fibers can be categorized into dry processing and wet processing. In dry processing, bales of cotton fibers are opened and conditioned, cleaned by carding and combing, drawn and then spun into yarns. Yarns are then woven or knitted into fabrics. The aesthetic value of fabrics is increased by dyeing, finishing or printing before or after the fabric is converted into the end product [27]. In order to impart color and other desired properties such as softness, wrinkle-resistance and flame-retardance, the fabric should be absorbent. The presence of non-absorbent impurities such as natural waxes, pectin and added impurities such as lubricants and sizing agents makes it necessary to pre-treat the fabric. Pre-treatment processes include desizing, scouring, bleaching and mercerization. Pre-treatment processes require large amounts of water and energy. For example, a typical open width bleach range consists of desize, caustic and peroxide bleach stages. Each stage has a washing train that requires about 50-75 gallons of water per minute. The temperature of the water needed in such a range is at least 181 °F. The range consumes

total of 16,450 pounds of steam per hour of which 10,350 pounds of steam is used by washing systems. The cost of processing textiles increases due to the increase in water consumption. For example, the amounts of water and energy used in a typical bleach range are listed in table 1 [28].

It has been estimated that careful optimization of bleach range configuration for types of fabrics processed, constant control of water and maintenance to avoid waste can lead to a savings of \$20,000 per year in water costs (at the rate of \$1.00/1000 gallons of water) and about \$110,000 per year in energy costs (at the rate of \$7.00/1000 pounds of oil-fired steam) [28]. Thus, reduction in water and energy consumption can account for a great reduction in the cost of textile wet processing.

Until the 1970s, production at low costs was the only consideration for a successful textile manufacturing facility. Pollution due to uncontrolled effluent release and inefficient use of resources were considered part of manufacturing cost. In recent years, as a result of increased level of environmental awareness of people and the government, companies unable to restrict levels of pollutants released into the environment are penalized [29]. As a result, the focus of textile manufacturers is to avoid environmental problems rather than find a solution after causing environmental problems.

Table 1. Water and Energy use in a Three-Stage Open-Width Bleach Range [28]

	Water (gal.hour) Incl. steam	Energy (lb. steam/hour)		
		Useful	Losses	Total
Saturators	550	300	400	700
Steamer and J-boxes	150	1150	150	1300
Washers				
Desize	3700)			
Caustic	3100)	8100	2250	10,350
Bleach	3150)			
Dry Cans	450	3600	500	4100
TOTAL	11,100	13,150	3300	16,450

Management strategies aimed at reducing water and energy costs include energy and water audits that help to identify water and energy costs for every process in the manufacturing unit. These costs can then be reduced by modifying the process. Textile mills are making efforts to reduce the level of pollutants in textile effluents by implementing the following strategies:

1. Treatment of effluents before release into the environment
2. Reuse of effluents after treatment
3. Recovery of chemicals and processing agents added
4. Substitution of chemicals that contribute to high COD and BOD of effluent

Enzymatic processing of textiles seems to be an attractive cleaner technology route to more energy efficient, less polluting and cheaper production processes.

Enzymes for Textile Applications

Enzymes have found applications in the following textile processes:

Desizing

Before weaving, warp yarns are strengthened in order to resist wear and tear during weaving by application of a size such as starch. After weaving, the size is removed to facilitate subsequent processing of the fabric. Alpha-amylases

are used to remove starch-based size from yarns. Starch molecules are broken down to soluble compounds by α - amylase that are easily removed during washing [30]. Lipases are also added to desizing treatment baths in order to remove lubricants added to size formulations [31].

Scouring

Natural waxes embedded in the matrix of the primary cell wall of a cotton fiber protect it from the elements in nature. During scouring, hydrophobic impurities are removed along with some natural color to make cotton absorbent and white. Cellulases and pectinases are used in bioscouring of cotton fibers [32, 33, 34]. An alkaline pectinase is now commercially available for large scale enzymatic scouring. Bioscouring has been reported to reduce total water consumption by approximately 25%, retain fabric strength and lower weight loss [35]. Efforts are now being made to reuse cellulases and make bioscouring a continuous process rather than a batch process [36, 37, 38].

Bleaching

Traditionally, bleaching of cotton fabrics is done by hydrogen peroxide treatment at high temperature and pH. Hydrogen peroxide itself decomposes to water and oxygen and hence is environmentally safe, but the conditions required for peroxide bleaching are harsh to the environment. Hence a cheap and environmentally safe bleaching process is being

investigated. Laccases, glucose-oxidases and xylanases (used in bleaching paper pulp) have been explored as bio-bleaching agents. Glucose-oxidases have been found to be the best alternative because of the following reasons:

- i) Glucose-oxidases produce the highest improvement in whiteness of cotton fibers and
- ii) Desizing effluents rich in glucose can be used in the bleaching process since glucose is required for the oxidation-reduction reaction [30, 39].

Bleach Clean-up

Residual peroxide from the bleaching process must be removed to ensure a good dyeing. In conventional processes, hydrogen peroxide is removed by two or more hot water rinses which make it a very energy and water intensive process.

Catalases decompose hydrogen peroxide to oxygen and water and their use in residual bleach clean-up needs only one hot water rinse at 80-95 °C. Not only is the residual hydrogen peroxide completely removed by catalase, but it also results in an overall process that is 6 to 8% cheaper than the conventional bleaching process [40, 41].

Denim Stone-washing

Denim stonewashing is a technique used to accelerate the fading and softening of jeans by washing denim garments along with pumice stones, patented by Francois Girbaud in 1978 [42].

The faded effect created by abrasion against pumice stones cannot be controlled and causes excessive damage to garments during washing. Also, a high content of suspended particles in the wash water leads to environmental problems. Since 1987, denim stonewashing is done using cellulases instead of pumice stones [43].

Bio-Finishing

The removal of surface fuzz (short fibrils and microfibrils) from cellulosic fibers with cellulase is termed as biofinishing or biopolishing. Enzymatic removal of surface fibrils results in smooth, clean-looking fabrics that retain their original color value. Other advantages of biofinishing are improved drapability, fashionable wash-down effects and improved moisture absorbency [43].

Effluent Treatment

Effluents from textile processes have to be treated in order to remove excessive pollutants, but mainly any colored matter present in the effluent. Living or dead organisms and selected microbes or enzymes may be used to treat effluents [44]. White rot fungi are found to degrade coloring matter very effectively in nature and hence enzymes from these fungi are being investigated to degrade colored impurities in textile effluents [45].

Current research focused on enzymatic processing of textiles is directed towards:

1. Making current enzymatic treatment methods efficient and effective [46]
2. Identifying new enzymes that can improve the properties of textiles [47].

For example, treatment with serine proteases on cross-linked cotton fabrics is being investigated. Serine proteases hydrolyze the amide bonds of cross-linked fabrics resulting in recovery of strength loss due to crosslinking. A short treatment with a low concentration of serine proteases at neutral pH and low temperature can result in efficient recovery of strength loss in amide crosslinked cotton fabrics [48].

Phosphorylation of cotton cellulose using hexokinase has been investigated as an alternative to chemical phosphorylation, which is a complicated process to introduce phosphorus groups in cellulose [49].

Advances in genetic engineering and enzyme technology are making it possible to develop enzymes with improved properties for textile applications. Biotechnology in textile industry will continue to offer solutions to make textile processing clean and eco-friendly.

Cellulases

Nature employs cellulases to break down cellulose produced by photosynthesis at a rate of at least 10^9 tons/year [50]. Most of the cellulose is broken down by cellulolytic organisms that use it as a source of carbon but degradation of cellulose by cellulases occurs through other means in nature. For example, trees shed leaves that are inefficient and unable to produce food through a process of senescence and leaf abscission. Near the end of the senescence process, abscission zones are created. Plant cells in this zone secrete pectinase and cellulase that degrade and reduce the strength of middle lamella and primary wall between cells, thus detaching leaves from the plant [51]. Plants also employ cellulases in the ripening of fruits.

An interest in this natural phenomenon of enzymatic hydrolysis of cellulose was generated during the World War II. After a brief period in a humid tropical climate, fungi ruined cotton gear, tents, uniforms and other military gear belonging to American units stationed in the South Pacific. Several laboratories within the Armed Services started investigating the nature of this "rotting". Later, it was established that the fungus responsible for the destruction was a cellulose-destroying fungus [52].

Interest in cellulose hydrolysis was rejuvenated in the 1970's when a global energy crisis emerged with the shortage of

fossil fuels. Cellulose was looked upon as a renewable source of energy as it can be broken down to glucose, which on fermentation gives ethanol - an alternative source of fuel. Mineral acids were used for the break down of cellulose, but the process is difficult to control, needs acid resistant containers and is not environmental friendly. Cellulases were employed to hydrolyze cellulose to glucose instead of acids [52]. Recent applications of cellulases include the selective modification of cellulosic materials and surface treatment without affecting mechanical properties of the substrate. Cellulase hydrolysis and action of cellulases on cotton fibers has been studied by researchers in the past [53] [54]. In these studies the effect of different factors affecting cellulose hydrolysis such as agitation and pretreatment of cotton has been studied [55, 56, 57]. The effect of enzymatic treatment on the hand properties, dyeability and tensile strength of cotton has also been studied in the past [58, 59, 60, 61, 62]. Knowledge of the properties of the substrate and enzymes makes it easy to understand the action of cellulases on cotton.

Structure of endoglucanases and exoglucanases

In 1986, Van Tilbeurgh[50] showed that exoglucanase I (CBH I) can be dissected into two functional domains by limited proteolysis. Later, other enzymes such as CBH II were also

found to have a similar arrangement. A general architecture of cellulases was then proposed based on these studies. A cellulase molecule can be divided into the following [50, 63]:

1. catalytic domain responsible for hydrolysis of cellulose
2. cellulose binding domain that promotes adsorption onto the insoluble cellulose
3. linker peptide that joins the two domains together

Structural investigations using small angle X-ray scattering (SAXS) later confirmed the two domain architecture of cellulases [50].

Catalytic Domains:

Based on the amino acid sequence, catalytic domains of cellulases can be grouped into 6 families, A-F. Each family of catalytic domain is characterized by a conserved fold and different folds are expected in different families. This implies that enzymes belonging to the same family will show a similarity in folding pattern resulting in an overall conservation of active site topology. The endo- or exo mode of action of cellulases is determined by the specific details of 3D structure. Endo and exo cellulases in the same family have a similar global 3D fold but differing substrate specificity due to minor details of the structure. For example: CBH II from *T. reesei* that belongs to Family B has a β/α barrel structure with the active site having a tunnel shape. Endoglucanases of the

same family were found to have similar overall fold but their active site was found to open into a cleft allowing random binding of the cellulose chains [50, 64].

Cellulose Binding Domain (CBD):

CBDs can be grouped into five distinct families (I - V) based on amino acid sequence similarities with their catalytic domain. Most CBD's are structurally well defined domains linked to either the N- or C-terminus of the catalytic domains. CBDs binding to crystalline cellulose may differ in size and topology but show a similar binding interaction [65].

The primary function of CBDs is believed to be attaching the cellulase on the surface of the substrate, but CBDs can take a more active part in cellulose degradation than the mere anchoring of cellulases to their substrate. CBDs are found to increase the concentration of the enzyme on the substrate and hence it may imply that the presence of CBDs increases the enzymatic activity of cellulases. Since catalytic domains are responsible for hydrolysis, the presence of CBDs may have little or no influence on activity of cellulases [66]. CBDs might help in loosening the individual cellulose chains from the cellulose surface prior to its actual hydrolysis. CBDs can penetrate the fibers at surface discontinuities thereby releasing noncovalently attached fragments and uncovering new cellulose chain ends [66].

The best-characterized type of CBD is type 2. CBDs of type 2 have been found exclusively among fungal cellulases. The domain is shaped like a wedge whose dimensions are approximately 30X18X10 °A. One side of the wedge is hydrophilic, and the other is hydrophobic. The hydrophilic side which presumably makes contact with the surface of the substrate contains three tyrosines. It has been proposed that the wedge-like shape of fungal CBDs not only promotes binding to the substrate, but also helps to peel off the cellulose chains from the top layer of cellulose microfibrils [66].

Linker Peptide:

The catalytic and cellulose binding domains in exoglucanases and endoglucanases are connected by a linker peptide. Since the cellulose binding domain may get attached to the cellulose substrate, the linker peptide provides a dynamic attachment of the catalytic domain on the surface of the substrate. The interdomain region may vary in length in different forms of endoglucanases and exoglucanases, thus affecting their enzymatic activity. The linker peptide is sensitive to proteolytic digestion and hence some culture solutions may contain enzyme forms both with and without CBDs [65].

Exoglucanases:

CBH I make up 60% and CBH II 20% of the total cellulolytic protein in cellulases from *Trichoderma reesei*, accounting for most of its cellulolytic activity. These two enzymes can achieve complete, although slow, solubilization of cellulose crystals even without the help of endoglucanase [63].

Exoglucanases have tunnel shaped active sites. Active sites are located in tunnels enclosed by two or four long loops [67]. The active sites of exoglucanases (CBH I and CBH II) are designed to bind a single glucose chain in a defined orientation [68]. For both enzymes, cellobiose is the major reaction product released from the non-reducing end of the chain by CBH II and from the reducing end of the chain by CBH I. It is thought that a single cellulose chain is fed into these tunnels from one end, followed by their threading through the entire length of the tunnel and subsequent bond cleavage and release of the cellobiose product from the far end of the tunnel. The remaining chain is still bound to the enzyme through interactions at a number of sub-sites. This creates the conditions for a processive mode of action in which several consecutive hydrolytic events occur without dissociation of the enzyme and the substrate.

CBH II has a shorter tunnel and hence falls off the substrate and reinitiates hydrolysis at an adjacent chain end.

CBH I which has a much longer tunnel would bind more tightly to the remaining glucan chain and is less likely to dissociate after each hydrolytic event. Therefore once bound to an available chain end, CBH I would continue hydrolyzing to the end. Cellobiohydrolases similar to CBHII, which have shorter active-site tunnels, may exhibit increasing degrees of endoglucanase type activity due to fewer loops with different lengths and/or mobilities.

Endoglucanases:

Endoglucanases preferentially attack internal glycosidic linkages of substrates such as acid swollen cellulose and soluble glucan, besides modified substrates such as CMC. Endoglucanases with open active sites can bind to and act in the middle of the glucan chains. The sequences corresponding to the active-site loops of exoglucanases are not present in homologous endoglucanases [69].

Structure of cotton fiber

A cotton fiber is a single cell fiber growing from a modified epidermal cell of cotton seed. Cotton is made of cellulosic and noncellulosic components. Table 2 lists the percentage composition of the cellulosic and noncellulosic components present in a typical cotton fiber [70]. Cellulose is present as fibrils organized in different layers whereas the noncellulosic

Table 2. Chemical composition of cotton fibers [70]

Constituents	Percent (dry basis)		
	Typical	Low	High
Cellulose	94.0	88.0	96.0
Protein	1.3	1.1	1.9
Pectic substances	0.9	0.7	1.2
Ash	1.2	0.7	1.6
Wax	0.6	0.4	1.0
Malic, citric and other organic acids	0.8	0.5	1.0
Total sugars	0.3		
Pigment	Trace		
Other	0.9		

substances such as pectin, fats and waxes are largely confined to the outer layers of the fiber. Figure 1 represents the fiber structure of cotton that is divided into the following layers [71]

1. cuticle
2. primary wall
3. Secondary wall

Cuticle: The cuticle of cotton contains waxes and pectin in about equal amounts and is so intimately bound to the primary wall that it may not be considered as a separate layer.

Primary wall: The primary wall is less than half a micrometer (μm) in thickness. The primary wall contains the bulk of the noncellulosic constituents of the cotton fiber. The cellulosic portion in this layer is a network of microfibrils interlaced in a fabric in which the general system of orientation is axial on the outer surface and transverse on the inner surface. Also, the cellulose fibrils of the inner surface of the primary wall are finer than those of the outer face [72].

Secondary wall: The secondary wall of cotton fiber is entirely cellulose. The secondary wall can be divided into the S-1 layer, S-2 layer and the lumen wall or S-3 layer. The S-1 layer is the very first layer of cellulose deposited after the cell grows to its full length. The S-1 layer is coarse and the fibrils arranged in a banded structure of alternate close and

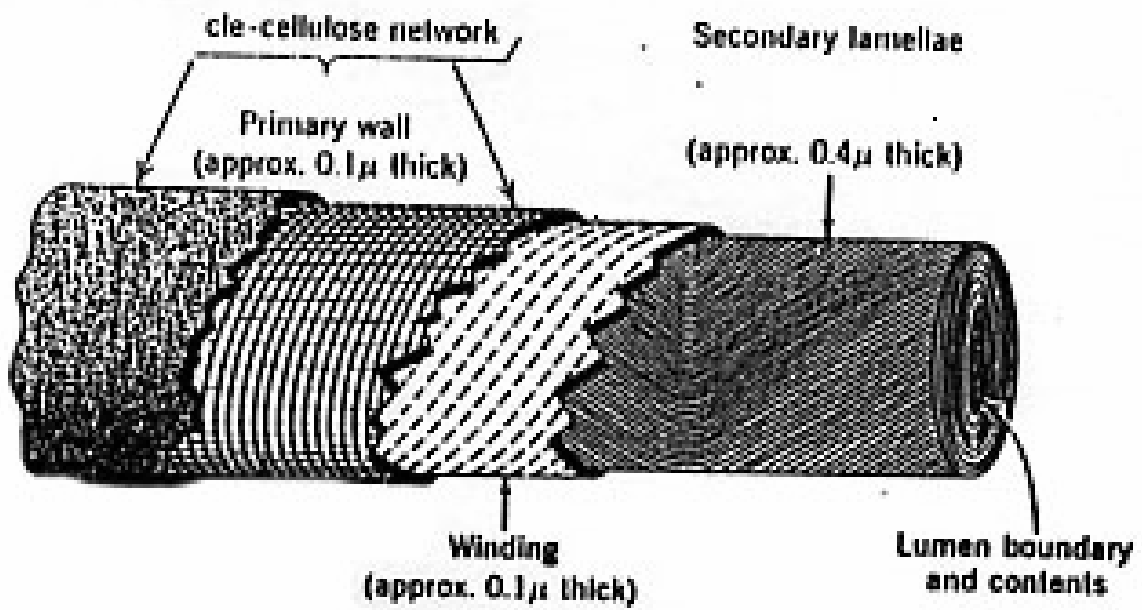


Figure 1. Structure of a Cotton Fiber [71]

open packing. The bands appear to spiral about the fiber axis at an angle of 45 degrees. The fibrils in the S-1 layer periodically reverse in direction and so a system of left and right (S and Z) spirals is set up along the length of the cotton fiber.

The S-2 layer is the main body of the fiber and can be as much as 4 to 5 μm in thickness. The layers in this part of the secondary wall can be separated fairly easily when swollen in a solvent for cellulose. The cellulose of the S-2 layer appears to occur in compact layers of closely aligned microfibrils which lie almost parallel to each other and at a slight angle to the fiber axis. The direction of the spiral in the S2 layer is opposite to that of the spiral in the S1 layer.

The lumen wall or the S-3 layer may or may not be present and is found to be more resistant to reagents [73]. The hollow space inside the collapsed fiber is known as the lumen.

Adsorption models

Adsorption isotherms traditionally used in adsorption of gases and liquids on surfaces has been used to model cellulase adsorption. The models most commonly used in the cellulose-cellulase system are [74, 75]:

- a. Linear Adsorption Isotherm
- b. Langmuir Adsorption Isotherm

c. Freundlich Adsorption Isotherm

d. Combined Langmuir-Freundlich Adsorption Isotherm

In most adsorption studies, microcrystalline cellulose has been used as the substrate. Adsorption studies have also been conducted on natural substrates such as bagasse, newspaper pulp and treated wood chips [76, 77, 78]. In some adsorption studies cellulase has been considered as a multicomponent adsorbate whereas in other studies purified cellulases have been used.

A. Linear Adsorption Isotherm

The linear adsorption isotherm is also known as the Nernst's adsorption isotherm. The isotherm can be represented by the equation as follows [79]:

$$[A] = K_d[E] \quad \text{Equation 1}$$

where, [A] = amount of enzyme adsorbed on cotton fibers

[E] = amount of enzyme present in the solution

K_d = constant given by the slope

In this model, the amount of enzyme adsorbed by the substrate is proportional to the concentration of enzyme initially present in the solution.

B. Langmuir Adsorption Isotherm

The Langmuir adsorption model is based on two assumptions

1. The substrate is composed of uniform binding sites and,
2. There is no interaction between the adsorbing molecules.

The Langmuir model assumes that a monolayer of adsorbents is formed and that the free energy of adsorption remains constant.

Peiterson, et.al.[74], were the first to suggest that cellulase adsorption on microcrystalline cellulose can be explained by the Langmuir isotherm. The authors studied adsorption of a mixture of cellulase (crude cellulase) on microcrystalline cellulose. Microcrystalline cellulose was treated in an aqueous bath. The mixture of enzymes and cellulose was then centrifuged and the protein content in the supernatant was determined. The amount of adsorbed protein can be obtained by the following equation:

$$P_{ads} = [P] \frac{K_p \cdot P_{ads,m}}{(1 + K_p \cdot P)} \quad \text{Equation 2}$$

where,

P = protein concentration in supernatant (mg/ml)

P_{ads} = adsorbed protein (mg of protein/mg of cellulose)

$P_{ads,m}$ = maximum amount of protein adsorbed (mg of protein/mg of cellulose)

K_p = constant (mg/ml)⁻¹

Kim et.al.[80-85] studied the adsorption of individual endoglucanase and exoglucanase components on microcrystalline cellulose. The equation that the authors used to describe the adsorption of various types of endoglucanase and exoglucanase on microcrystalline cellulose is another form of the Langmuir

Isotherm. The experimental data obtained to fit the following equation,

$$[A] = \frac{[A_{\max}] K_{\text{ad}} [E]}{1 + K_{\text{ad}} [E]} \quad \text{Equation 3}$$

where,

A_{\max} = maximum amount of enzyme adsorbed per unit weight of cellulose

K_{ad} = adsorption equilibrium constant

$[E]$ = concentration of enzyme in liquid phase at adsorption equilibrium

The term K_{ad} was used by the authors as a measure of the affinity of cellulase to cellulose and to calculate thermodynamic parameters of the adsorption process such as change in heat of enthalpy (ΔH_a), change in Gibbs free energy (ΔG_a) and change in entropy (ΔS_a) [81] [84] [85].

Another form of Langmuir adsorption isotherm was suggested by Medve et.al [75]. The authors attempted to fit the adsorption data they obtained in their experiments to different adsorption isotherms that were used to explain adsorption of other proteins. The Langmuir adsorption model used by Medve et.al, [75] is given as follows:

$$B = \frac{n[F]}{K_d + [F]} \quad \text{Equation 4}$$

where,

B = amount of the bound enzyme ($\mu\text{mol/g}$ of substrate)

F = concentration of free enzyme in solution ($\mu\text{mol/L}$)

n = number of binding sites per gram of substrate ($\mu\text{mol/g}$)

K_d = dissociation constant of the adsorbent-adsorbate complex ($\mu\text{mol/L}$)

The authors characterized the affinity of the adsorbate (cellulase) to the adsorbent (cellulose) by a term called the distribution coefficient - α . Alpha is given by the initial slope of the Langmuir isotherm, which is given as

$$\alpha = \frac{n}{K_d} \quad \text{Equation 5}$$

The authors used a modified Langmuir adsorption model that assumed the presence of more than one kind of binding site written as follows

$$B = \frac{n_1[F]}{K_{d1} + [F]} + \frac{n_2[F]}{K_{d2} + [F]} \quad \text{Equation 6}$$

n_1 and n_2 = ($\mu\text{mol/g}$) number of adsorption sites/g of substrate

K_{d1} and K_{d2} = ($\mu\text{mol/L}$) dissociation constant for the two sites

Distribution coefficient of the mixture of adsorbent is given as the sum of the distribution coefficient for the two sites,

$$\alpha = \frac{n_1}{K_{d1}} + \frac{n_2}{K_{d2}} \quad \text{Equation 7}$$

and the total number of sites is given as $n = n_1 + n_2$

C. Freundlich Adsorption Isotherm

In an attempt to account for structural heterogeneity of cellulose substrates, the Freundlich isotherm was used to describe adsorption of cellulases [75]. Freundlich isotherm has been used to describe adsorption of α -amylase to starch granules.

The Freundlich equation can be derived from a Langmuir equation by assuming a certain distribution function for sites having different free energy values for the adsorption process and assuming Langmuir adsorption at each type of site [75]. Freundlich isotherm for adsorption to heterogeneous surfaces is given as follows:

$$B = K[F]^{1/m} \quad \text{Equation 8}$$

where,

K = Freundlich Equilibrium constant

m = power term of the Freundlich Isotherm ($m > 1$)

n = ($\mu\text{mol/g}$) number of binding sites/g of substrate

m and K are empirical parameters.

D. Combined Langmuir-Freundlich Model

The combined Langmuir-Freundlich isotherm used to describe adsorption to a heterogeneous surface is given as follows:

$$B = \frac{n[F]^{1/m}}{K + [F]^{1/m}} \quad \text{Equation 9}$$

where,

m = empirical constant

K = apparent adsorption constant.

Equation 9 can describe experimental data for adsorption of cellulases as accurately as the 2-site Langmuir model with fewer parameters, but interpretation of the derived parameters is difficult [86].

Apart from the models described, other models have been used to describe cellulase adsorption on cellulose [75]. The Langmuir isotherm however is by far the most widely used isotherm to describe adsorption of cellulases to cellulose.

Adsorption of Exoglucanases

From the adsorption equilibrium constants, Kim et.al, [81] suggested that CBH I exhibits a greater affinity to cellulose compared to CBH II.

Kim et.al, [74][81] found that a mixture of CBH I and CBH II in the weight ratio of 1:1 adsorbed more spontaneously than either CBH I or CBH II individually. The authors suggested that

CBH I and CBH II partially form a CBH I-II complex which has a higher affinity for cellulose than the individual components. The Langmuir adsorption isotherm was found adequate to explain the adsorption data. On the other hand, Medve et.al., [75] [86] reported that the combined Langmuir-Freundlich adsorption or the 2-site Langmuir adsorption model described the adsorption of CBH I and II, together from a mixture, with a greater correlation than what was obtained from the Langmuir isotherm.

Adsorption of Endoglucanases

Adsorption of mixtures of endoglucanases as well as the individual endoglucanases such as EG I and EG II has been investigated by many researchers [59] [82] [83] [84] [85] [86]. Beldman et.al, [87] identified and studied the adsorption of six endoglucanases as EG I-VI. Based on values of the initial slope of adsorption isotherms plotted, EG I, III and V adsorbed more strongly than EG II, IV and VI. Results suggested that enzymes that adsorbed strongly are not necessarily less random in hydrolytic activity than the weakly adsorbed enzymes. Contradictory to this classification of endoglucanases reported, Kim, et.al, [82] found that EG I and II have higher affinity than EG III and IV based on their adsorption equilibrium constants. Kim et.al., [82] also suggested that the endoglucanases with high affinity to cellulose bind at sites

where the crystalline structure of the substrate is disturbed. It was suggested that endoglucanases induced defibrillation of the crystallites and opened new sites for the action of exoglucanases. Adsorption of crude and different types of endoglucanases followed Langmuir adsorption [83, 84, 85].

Adsorption of Mixtures of Endoglucanases and Exoglucanases

A mixture of crude endoglucanases and exoglucanases in weight ratio of 1:1 was found to show synergistic action on cellulose hydrolysis. An optimal ratio of endoglucanase and exoglucanase should be maintained during the course of hydrolysis, for maximum hydrolytic action. However, it has been suggested that the ratio may not be maintained, for example, because of a decrease in number of sites for endoglucanases and increase in number of sites for exoglucanases and individual nature of adsorption of endoglucanases and exoglucanases [85, 88].

The composition of a mixture of various endoglucanases and exoglucanases would affect the adsorption behavior due to differences in adsorption characteristics of types of endoglucanases and exoglucanases. Mixtures of individual types of endoglucanase or exoglucanases in different combinations have been studied. Some components of endoglucanase and exoglucanase show synergism in hydrolytic action while others do not show any

synergism. For example, CBH II shows synergistic action when present in weight ratio of 1:1 with EG I or EG II [82, 83, 84]. A mixture of EG I and CBH II hydrolyzes microcrystalline cellulose to a greater extent than EG I or CBH II individually. CBH II showed synergistic action in the presence of EG II but in presence of EG I, CBH II showed preferential adsorption on microcrystalline cellulose. CBH II was found to be inactive on microcrystalline cellulose when used alone.

Summary

Most of the studies described in the literature survey are conducted on cellulose substrates such as microcrystalline cellulose. Since there may be differences in the properties of microcrystalline cellulose and cotton, the adsorption characteristics of cellulases on cotton fibers may be different. In addition, the temperatures at which most studies described in the survey of literature were performed are not a representation of the temperature of textile wet processing conditions. Hence it is important to study the adsorption characteristics of purified cellulases on cotton under practical conditions of temperature and concentration. A study of the adsorption of purified cellulases will be helpful in understanding the action of cellulases on cotton and also in developing cellulase formulations that are more efficient.

Chapter 3

MATERIALS AND METHODS

Materials

Substrate

Raw cotton fibers (Texas Grade 52, Low Middling Light Spot Cotton) were obtained from Avondale Mills, Monroe, GA. The fibers were then cleaned and used in this study.

Enzymes

Celluclast[®] a commercial mixture of endoglucanases and exoglucanases produced from *Trichoderma reesei*, a fungus mainly used for commercial production of cellulases, was obtained from Novozymes NA, Franklinton, North Carolina. Endoglucanases and exoglucanases required for the adsorption experiments were purified from this commercial mixture using Fast Protein Liquid Chromatography (FPLC).

Spectrophotometer

A Spectronic Genesys 2 UV-VIS spectrophotometer was used for single spectrophotometric measurements. For large numbers of spectrophotometric measurements, a microplate spectrophotometer, 340 ATTC, was used.

Reagents

The following reagents were used in the purification of enzymes:

1. Ammonium sulfate, Reagent grade supplied by J.T.Baker
2. Tris(hydroxymethyl)aminomethane also known as Trizma Base or Tris, Reagent grade supplied by Sigma
3. NaCl, Reagent grade supplied by J.T.Baker
4. Hydrochloric acid, Reagent grade supplied by J.T. Baker

The following reagents were used in the adsorption experiments:

1. Sodium acetate, Reagent grade supplied by J.T. Baker
2. Acetic acid, Reagent grade supplied by J.T. Baker
3. Cellobiose, Reagent grade supplied by J.T. Baker
4. 2,4 - dinitro salicylic acid (DNS), Reagent grade supplied by Sigma

Methods

Substrate Purification

Impurities such as vegetable matter, seed fragments and very short fibers were removed by passing the fibers through a Shirley Analyzer. Cotton fibers were then scoured at 80 °C for 60 minutes. The material to liquor ratio used was 1:40. Scouring solution was prepared using the following ingredients:

- Sodium Carbonate: 5 gpl
- Wetting Agent: 1 gpl (Clavodene MCM-2)

- Chelating Agent: 1 gpl (Barapon C-108)

The scouring solution was heated to 60 °C and the cotton fibers were introduced into the bath. The bath was stirred with a glass rod intermittently. After 60 minutes the cotton fibers were collected and washed with hot and cold water. The fibers were then washed with deionised water until the pH of wash water pH was 7. The fibers were then air dried and stored in a standard atmosphere at 65% R.H. and 21±1 °C.

Enzyme Purification

Purification of Enzyme:

Endoglucanases and exoglucanases were purified from a commercial cellulase mixture, Cellulclast, using Fast Protein Liquid Chromatography (FPLC) system [89] [90]. The purification of the endoglucanases and exoglucanases involved the following steps:

1. Sample Preparation
2. Separation of enzyme components on DEAE Sepharose CL-6B column
3. Separation of enzyme fractions on SP-Sepharose column
4. Separation of enzyme fractions on Mono S column HR 10/10

Different columns were packed for the separation of endoglucanases and exoglucanases using the FPLC system. The packing resins to be packed in columns for the separation of

enzymes were obtained from Amersham Pharmacia Biotech, Piscataway, NJ. The properties of the packing resins in different columns are listed below:

1. HiPrep 26/10 Desalting column packed with Sephadex G-25 Fine. Sephadex G-25 Fine is commercially available bead-formed gel prepared by cross-linking dextran with epichlorohydrin. This column available as a packed column was obtained from Amersham Pharmacia Biotech, Piscataway, NJ, USA.
2. DEAE Sepharose CL-6B column is packed with DEAE Sepharose CL-6B, a commercially available weak anion exchanger with diethylaminoethyl as the ion exchange group.
3. SP Sepharose XL column is packed with SP Sepharose XL, a commercially available strong anion exchanger based on a highly cross-linked, bead formed 6% agarose matrix. Dextran chains are covalently coupled to the agarose matrix and sulphopropyl ion exchange groups are attached to this dextran through chemically stable ether bonds.
4. Mono S HR 10/10 is a commercially available strong cation exchange column based on a beaded hydrophilic resin with a particle size of 10 μm . The charged group on the gel is methyl sulphonate. The capacity of the gel is 0.14-0.18 mmol/ml and substances with molecular weights up to 10^7 can be successfully separated. The protein capacity is in the

range of 20-50 mg/ml gel. Mono S HR 10/10 is a column readily available with Amersham Pharmacia Biotech, Piscataway, NJ.

1. Sample Preparation: Celluclast was prepared prior to loading on the DEAE-Sepharose column for separation of the individual endoglucanases and exoglucanases. In this process, diluents in Celluclast mixture such as sorbitol and sodium chloride and preservatives such as potassium sorbate were separated from the proteins. The proteins in the commercial cellulase mixture solution were precipitated with 75% by weight of ammonium sulfate. After precipitation, the solution was centrifuged and the precipitate was collected by decanting the supernatant salt solution. The precipitate was dissolved in a minimum volume of tris-HCl buffer. In the initial trials three different methods were employed to remove any residual ammonium sulfate from the solution - dialysis, ultrafiltration and desalting column.

The dialysis bags are made of modified cellulose and hence are easily hydrolyzed by cellulase solution resulting in the breaking of dialysis bags. Repeated trials using dialysis bags indicated that short periods of dialysis were effective and did not result in breaking of the dialysis bags. Traces of ammonium sulfate from the enzyme solution could also be removed by ultrafiltration using a polyethersulfone membrane, PM30 manufactured by Millipore, Billerica, Massachusetts. The

enzyme solution was placed in the ultrafiltration cell and nitrogen gas passed through the ultrafiltration cell to prevent denaturation of the protein. The salt solution was filtered through the membrane under vacuum and discarded after testing for the presence of protein by the Bradford method []. During ultrafiltration, the protein solution was stirred continuously to prevent clogging of the ultrafiltration membrane.

Ultrafiltration of highly concentrated protein solutions was difficult since the membrane got clogged over time and had to be removed and washed periodically. Desalting of the protein solution was tried using a desalting column HiPrep 26/30. The gel filtration column separates the components of the solution into based on their molecular weights. The high molecular weight compounds such as the protein molecules are excluded from the gel and so eluted first whereas the low molecular weight substances such as ammonium sulfate enter the pores and are thus eluted later. The desalting column was found to be very efficient with protein solutions of low concentrations. The use of desalting columns did not require precipitation of protein from commercial mixture using ammonium sulfate.

Desalting of Celluclast was repeated using small volumes each time until sufficient quantities of desalted protein solution was collected.

2. Separation of Enzyme Components on DEAE- Sepharose CL-6B

Column:

DEAE Sepharose 20 mM Tris-HCl buffer (Buffer A) at pH 7.5 was used as a starting buffer. The buffer was used to load the column with the desalted protein solution. The flow rate of the starting buffer was maintained at 5 ml/min. The eluting buffer used was 20 mM Tris-HCl solution at pH 7.5 with 1 M NaCl. A salt gradient of 0-100% salt was provided by the 1 M NaCl present in the eluting buffer.

When the mixture of enzymes was injected, CBH I was adsorbed onto the column and later eluted by the elution buffer. CBH II and EG II did not adsorb to the DEAE Sepharose column and were eluted with the starting buffer A [89, 90]. CBH I was then concentrated by precipitation with ammonium sulfate and dialysis. The purity of the enzyme was later checked by SDS Page Gel Electrophoresis.

3. Separation of enzyme fractions on SP-Sepharose XL column

The unadsorbed fraction of the enzyme mixture containing CBH II and EG II was concentrated by precipitation with 75% ammonium sulfate followed by desalting by using the Fast Desalting Column (Pharmacia). The concentrated fraction was then injected into a SP-Sepharose column using a 10 mM sodium acetate starting buffer at pH 4.0 (Buffer A). EG II did not adsorb on the SP-Sepharose XL column and was eluted along with

the starting buffer [90]. The eluting buffer used was 10mM sodium acetate buffer with 0.5 M NaCl. The adsorbed CBH II was eluted with the elution buffer and concentrated by precipitation and dialysis. The fraction unadsorbed in SP-Sepharose contained more than one enzyme and hence had to be purified to homogeneity.

4. Separation of enzyme fractions on Mono S HR 10/10 column

The unadsorbed fraction was then injected into a Mono S column HR 10/10 using a starting buffer of 10 mM sodium acetate at pH 3.0. The eluting buffer used was 10 mM sodium acetate and 1M NaCl. All the protein was adsorbed on the column and then eluted with low salt whereas the other fraction was eluted at high salt concentration. EG II fraction was then checked for purity by preparing a SDS PAGE Gel.

At the end of the enzyme purification process, pure fractions of CBH I, CBH II and EG II were obtained.

Measurement of Purity of Enzymes:

The purity of separated endoglucanases and exoglucanases was measured using Sodium Dodecyl Sulphonate- Polyacrylamide Gel Electrophoresis (SDS-PAGE) [91]. Electrophoresis is the process of moving charged molecules in solution by applying an electric field across the mixture. The migration of the molecules to a

charged pole is dependent on their charge, shape and size [92, 93].

Procedure:

1. Acrylamide Gel Preparation: The polyacrylamide matrix used in SDS-PAGE electrophoresis is divided into two different layers, the lower layer or the separating or resolving gel (12% acrylamide) and the upper layer or the stacking gel (4% acrylamide). The compositions of these layers are given in Table 3. The separating gel is responsible for separating the polypeptides by size and the stacking gel causes the samples to be compressed or stacked in order to have thin bands. Ammonium persulfate (PSA) is used as the indicator peroxide and N, N, N', N'- tetramethylethylenediamine (TEMED) is used as a catalyst to polymerize the acrylamide solution. The preparation of the acrylamide gel is a standard procedure and the details were obtained from the electrophoresis handbook provided by Amersham Biochem [94].

Enzyme denaturization: Enzyme solutions of 5 and 10 µg/µl of CBH I, II and EG II were prepared. The protein solutions were boiled with an equal amount of loading buffer. The loading buffer contained tris buffer (0.25 M, pH 6.8), 20% SDS, mercaptoethanol, glycerol and bromophenol blue. SDS, which is an anionic detergent, denatures proteins by wrapping the

Table 3. Composition of the separating and stacking gels in SDS PAGE Gel

	Separating	Stacking
Acrylamide	2.09 mL	0.33 mL
SDS (20%)	25 μ L	12.5 μ L
Tris	1.5 μ L	1.25 μ L
Temed	2.5 μ L	2.5 μ L
Water	1.8 mL	0.765 mL
PSA	125 μ L	125 μ L

(SDS stands for Sodium dodecyl sulfonate, Tris stands for tris(hydroxyl)aminomethane, temed stands for N, N, N', N'-tetramethylethylenediamine and PSA stands for ammonium persulfate)

hydrophobic tail around the polypeptide backbone. SDS confers a net negative charge to the polypeptide in proportion to its length.

2. Electrophoresis: The denatured enzyme solutions were then poured into the wells of the stacking gel. A low molecular weight marker was added to one of the wells on the stacking gel. The marker acts as a reference and may be used to estimate the molecular weight of the enzyme. The gel was then placed in an electrophoresis tank and the buffer was slowly poured into the tank. A continuous buffer system (0.25 M tris, 1.92 M glycine and 0.1% SDS) was used in this experiment. The electrodes were connected and a potential difference of 160 V was applied across the gel for 50 minutes. At the end of 50 minutes, the gel was removed from the buffer tank.
3. Staining and Destaining of the gel: The gel was removed from the stand and placed in 50 ml of staining solution for 2 hours. The staining solution was prepared by mixing 0.2% Coomassie Brilliant Blue R-250 dye, 45% methanol and 10% acetic acid. The stained gel was then placed in a container with destaining solution. The destaining solution was prepared by mixing 100 ml of glacial acetic acid, 200 ml of ethanol and 700 ml of water (v/v). Destaining was conducted overnight with shaking provided by placing the container on a shaker operated at 80 rpm. The gel was then removed from the

destaining solution, washed with deionized water, dried and photographed [94].

Enzyme Adsorption Procedure

Enzyme solutions for the following concentrations (mg/ml) were prepared from the concentrated solutions of pure enzymes;

CBH I: 0.493, 0.737, 0.976, 1.159, 1.412, 1.827, 2.472

CBH II: 0.477, 0.715, 0.929, 1.315, 1.335, 1.436, 1.774

EG II: 0.362, 0.690, 0.997, 1.140, 1.556, 1.857, 2.069

Adsorption of enzymes was conducted in clean test tubes. One milliliter of enzyme solution was taken in a 15 ml test tube. The test tubes were placed in a water shaker bath operated at 50 rpm and heated to the pre-determined temperature. After the enzyme solution reached the set temperature, 0.1 grams of cotton fibers were added to each test tube. The test tubes were incubated for 60 minutes after which the fibers were removed from the bath. The enzyme solution was centrifuged at 14000 rpm for one minute to remove any suspended particles. The supernatant was transferred to clean Eppendorf tubes and stored at 3 °C until the protein was measured.

Analytical Procedures

Determination of Protein in Solution

The amount of protein present in solutions was measured using two methods:

1. Bradford method using Coomassie Brilliant Blue G-250 was used for quick measurements [95].
2. Bicinchoninic acid method (BCA method) was used for large number of measurements [96].

Bradford Method

Principle:

The anionic form of Coomassie Brilliant Blue G-250 present in acidic conditions forms a complex with protein. The binding of the dye to the protein results in an absorbance shift from 465 to 595 nm. The absorbance of the dye-protein complex is proportional to the protein concentration. Bovine Serum Albumin solution is used as the standard for calculating the amount of protein present in solutions.

Spectrophotometric Measurement of Color:

Ten microliters of supernatant solution were added to 1 ml of Bradford reagent. After 15 minutes, the absorbance was measured at 595 nm. A calibration curve plotted for a range of Bovine Serum Albumin concentrations was used to determine the protein concentration.

BCA Method

Principle:

Cupric ions (Cu^{+2}) are reduced by protein in an alkaline medium to cuprous ions (Cu^{+1}). The cuprous ions (Cu^{+1}) can be selectively detected by a highly sensitive detection agent, bicinchoninic acid (BCA). Two molecules of BCA react with one cuprous ion (Cu^{+1}) and form a purple-colored complex that exhibits a strong absorbance at 562 nm.

Preparation of BCA Working Reagent (WR):

BCA reagent was prepared by mixing BCA reagent A and BCA reagent B in the ratio of 50:1 (Reagent A:B).

The following formula was used to determine the total volume of WR required:

$$(\# \text{ standards} + \# \text{ unknowns}) \times (\# \text{ replicates}) \times (\text{volume of WR per sample}) = \text{total volume of WR required.}$$

Microplate Procedure:

1. 10 microliters of each standard or unknown sample replicate were taken in a microplate well.
2. 200 microliters of the WR were added to each well and the solutions were mixed by placing the plate on a plate shaker for 30 seconds.
3. The plate was then covered and incubated at 37 °C for 30 minutes.

4. The plate was cooled to room temperature and the absorbance was measured at 562 nm on a plate reader.

Determination of Reducing Sugar

Activity of CBH I, CBH II and EG II on cotton fibers was estimated by determining the amount of reducing sugars present in supernatant by the dinitrosalicylic (DNS) acid method [97].

Principle of DNS Method:

The DNS method is used to test the presence of free carbonyl groups (C=O), or the reducing sugars as they are called, produced by the action of cellulolytic enzymes such as cellulases. A free carbonyl group present in a sugar, for example glucose, is oxidized to a carboxyl group and simultaneously 3,5-dinitrosalicylic acid (DNS) is reduced to 3-amino, 5-nitrosalicylic acid under alkaline conditions. The color produced by 3-amino, 5-nitrosalicylic acid is measured at 575 nm.

Preparation of DNS solution:

DNS solution used in detecting reducing sugars is prepared as follows:

1. Dinitrosalicylic acid Reagent Solution, 1%
 - a. Dinitrosalicylic acid - 10g
 - b. Potassium Sodium Tartrate Solution, 300g
 - c. Sodium hydroxide - 10g

d. Water to make the volume to 1 liter

Procedure:

1. 100 microliters of standard or unknown solution is measured and added to 100 microliters of deionized water and 400 microliters of DNS reagent in a 5 ml test tube.
2. The test tubes are covered with an aluminum foil to prevent loss of liquid due to evaporation and heated at 90 °C for 5 minutes.
3. After 5 minutes, the test tubes are cooled to room temperature in a cold water bath and the absorbance is measured at 575 nm.
4. The concentration of reducing sugars is determined using a standard curve obtained for a range of cellobiose solutions.

Measurement of Exoglucanase activity

Exoglucanase activity was measured by p-nitrophenyl- β -D-cellobioside method (p-NPC method) [98].

Principle of p-PNC method:

p-nitrophenyl- β -D-cellobioside (pNPC) can be used to measure the activity of exoglucanase. Exoglucanases act on an agluconic bond between p-nitrophenyl and the disaccharide moiety where as endoglucanases and β -glucosidase act on agluconic as well as

holosidic bonds. Exoglucanases act on pNPC molecule as shown in the reaction [98]:

$p\text{-NP-glu-glu} + \text{endoglucanase and } \beta\text{-glucosidase} = p\text{-NP} + \text{glu}$

$p\text{-NP-glu-glu} + \text{exoglucanase} = p\text{-NP} + \text{glu-glu}$

where glu is a single molecule of glucose. Interference due to endoglucanase can be compensated for by prior standardization of the assay procedure with a purified endoglucanase.

1. 0.5 milliliters of 1 mM solution of pNPC solution was mixed with a known volume of enzyme solution and incubated for 5 min at 50°C.
2. After 30 minutes 0.5 milliliters of 1% sodium carbonate was mixed with the above solution.
3. The amount of p-nitrophenol produced during the reaction is estimated spectrophotometrically at 410 nm. The color produced due to p-nitrophenol is proportional to the amount of exoglucanase present in the solution.

Application of adsorption isotherms

Different adsorption isotherms were tried to be fitted to the experimental adsorption data. Linear regression analysis was conducted to determine the goodness of fit of the different adsorption isotherms. For Linear adsorption isotherm, a plot of [E] v/s [A] was plotted. In the case of the Langmuir adsorption isotherm, plots of 1/ [E] against 1/ [A] and [E] v/s [E]/[A]

were plotted. For Freundlich isotherm, a plot of $\ln[E]$ against $\ln[A]$ was plotted. The last model that was investigated was the Combined Langmuir-Freundlich isotherm. In this case, a plot of $1/[E]^{(1/m)}$ against $1/[A]$ was plotted, where $m > 1$. In the present case the value of m used was 2.

Linear regression analysis was performed to determine whether the given model is a good fit of the experimental adsorption data. Based on the r-square values, the model that explains the experimental adsorption data the best was selected.

Chapter 4

RESULTS AND DISCUSSION

The results and discussion is discussed in two parts; purification of the enzymes is discussed in the first part and the adsorption of the enzymes on cotton fibers and the fitting of different adsorption models is discussed in the second part.

Part I

Separation of CBH I, II and EG II:

Celluclast was first desalted using a desalting column, HiPrep 26/10. A profile of the enzyme solution desalted on a desalting column is shown in figure 2. In figure 2, the x-axis is the volume of buffer solution collected in fractions during the desalting process and y-axis is the conductance of the solution and the optical density of the solution eluted from the HiPrep 26/10 column measured at 280 nm. The solid line on the plot is the amount of protein eluted from the column whereas the broken line represents the salt eluted from the column. The desalting column efficiently separated the protein from the impurities such as salt present in the solution. The desalting

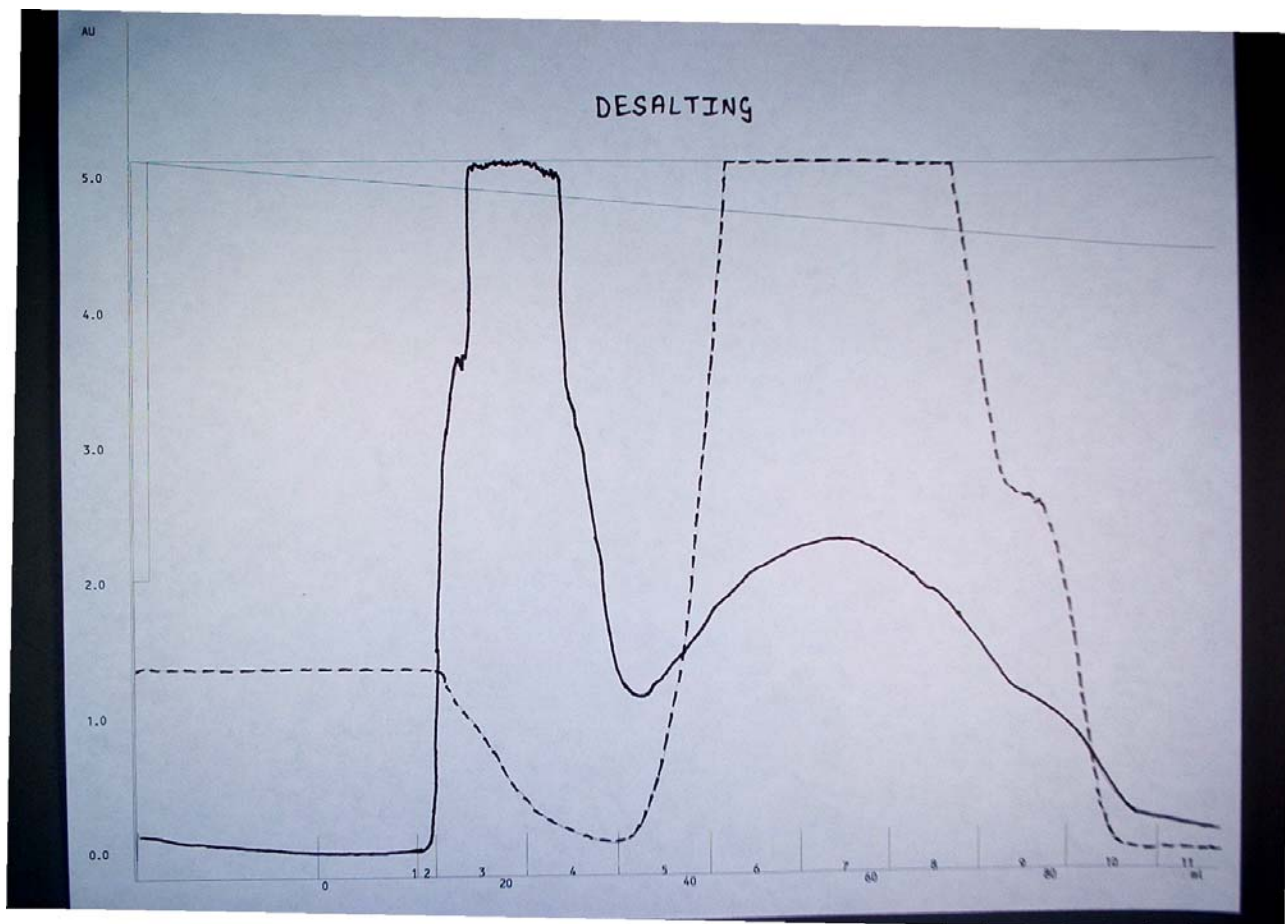


Figure 2. Profile of Enzyme Desalting Process on a Desalting Column

column was used at different stages of purification of the enzyme solution.

The desalted Celluclast was then injected onto a DEAE Sepharose CL-6B column. The enzyme fraction unadsorbed on the DEAE Sepharose column was eluted along with the starting buffer A. This enzyme fraction unadsorbed on the DEAE Sepharose column may contain CBH II and EG II. CBH I which was adsorbed on the DEAE Sepharose CL-6B column was eluted from the column by the salt gradient developed by buffer B.

Figure 3 is the elution profile of CBH I under the salt gradient created by buffer B. The fractions showing significant amounts of protein were collected, tested for exoglucanase activity using the p-nitrophenyl- β -D-cellobioside method and mixed together [98]. The eluted fraction was then precipitated, desalted and concentrated before storing at 3 °C.

The fraction unadsorbed on DEAE Sepharose CL-6B was collected, precipitated, desalted and then injected onto the SP Sepharose XL column. CBH II fraction was adsorbed on the SP Sepharose XL column while the EG II fraction is eluted along with buffer A. The adsorbed CBH II fraction was then eluted under a salt gradient, collected, concentrated by precipitation and dialysis and stored at 3 °C until used for adsorption experiments.

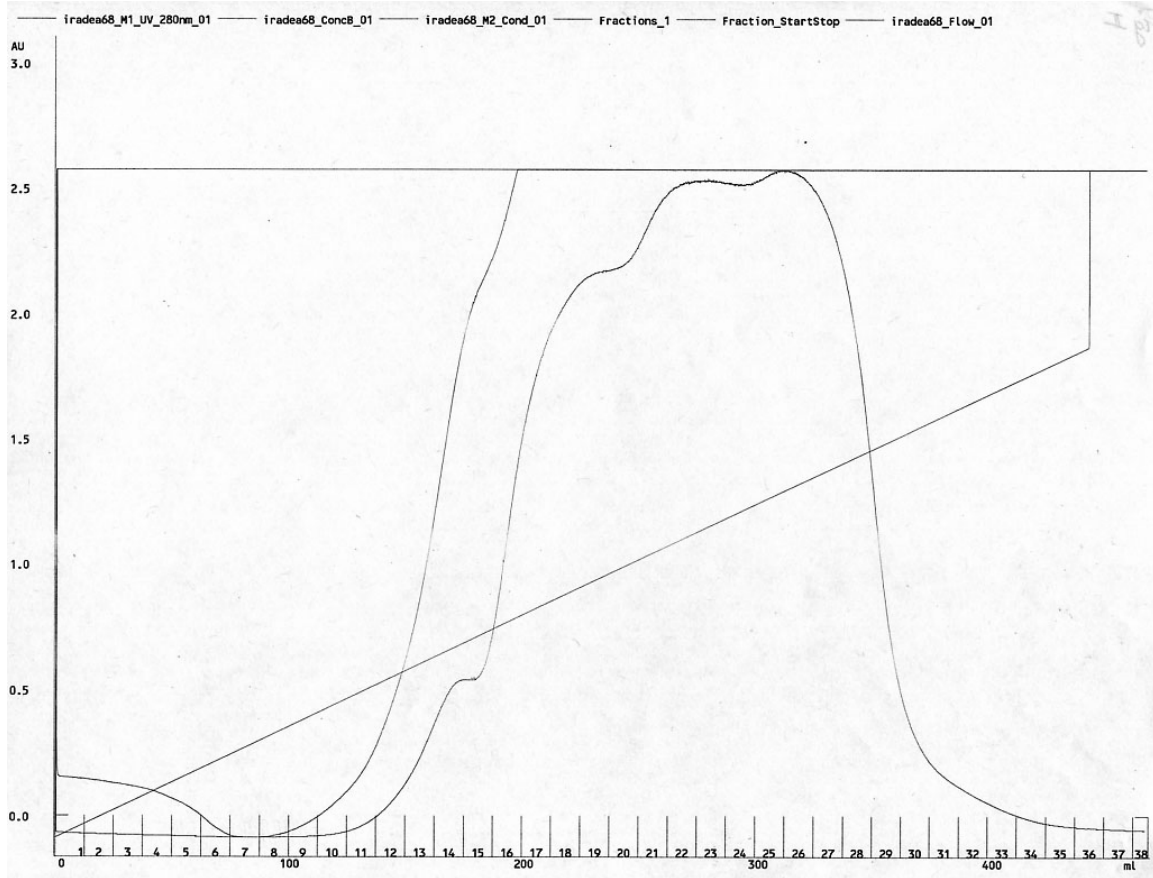


Figure 3. Profile of elution of adsorbed fraction (CBH I) on DEAE Sepharose CL-6B column.

EG II fraction unadsorbed on SP- Sepharose XL may have contained some impurities and hence was injected to Mono S HR 10/10 column. EG II fraction was then eluted at low salt concentration from the column. The fraction was collected, concentrated by precipitation and dialysis and stored at 3 °C until used for adsorption experiments.

Determination of the Purity of the enzymes:

The purity of the isolated enzymes was checked by SDS PAGE. The SDS PAGE was prepared as described in the materials and methods chapter. Figure 4 is a photograph of a SDS PAGE prepared for the three enzymes, CBH I, CBH II and EG II along with a low molecular weight marker protein that is a standard used in the SDS PAGE. A single solid band indicates that the protein is pure enough. From figure 4, indicates single bands for all the three enzymes, and hence that they are pure enzymes.

Part II

The adsorption data was evaluated by linear regression to determine the adsorption model that best explains the adsorption data. Four different adsorption models tested were: Linear, Langmuir, Freundlich and Combined Langmuir-Freundlich models. The following were plotted to test the fit of the four models to the experimental adsorption data obtained.

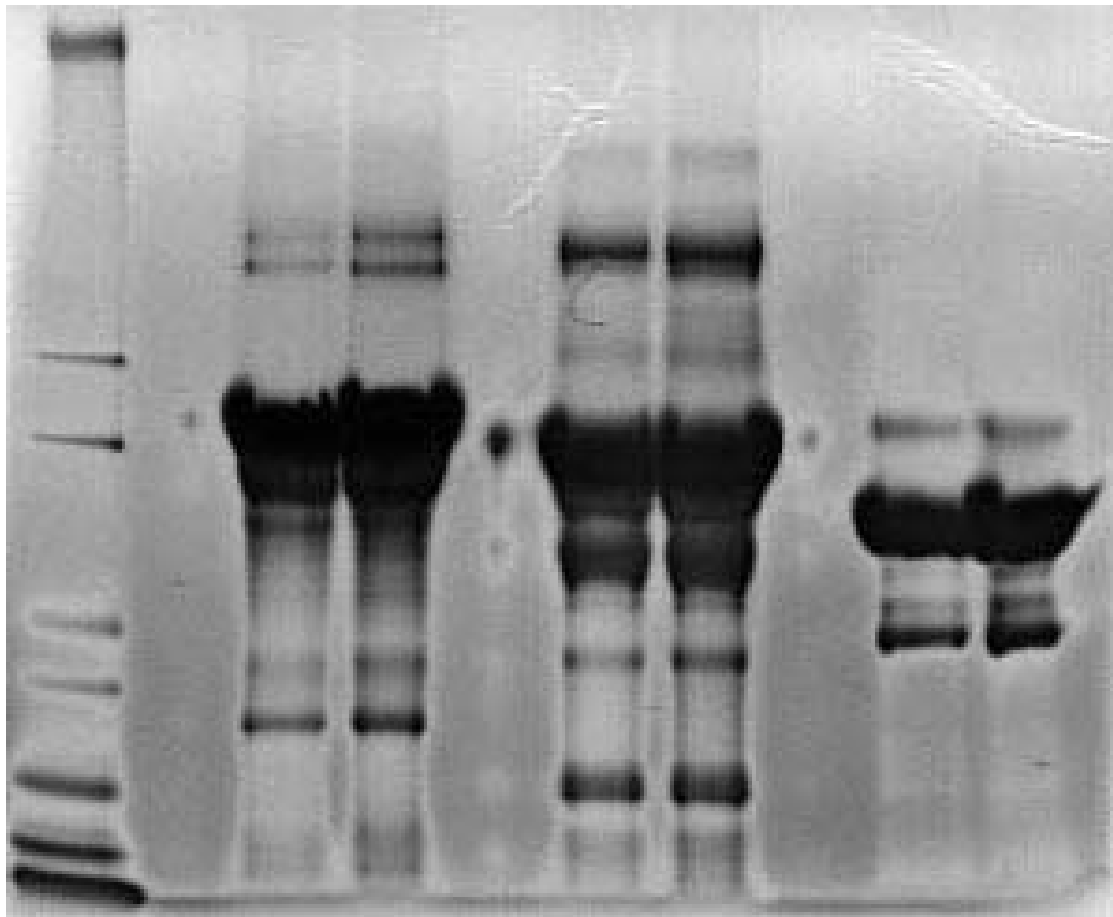


Figure 4. SDS PAGE to determine the purity of CBH I, CBH II and EG II enzymes.

1. $[E]$ v/s $[A]$ for Linear model
2. $[E]$ v/s $[E]/[A]$ for Langmuir model suggested by Kim et.al.
[81]
3. $1/[E]$ v/s $1/[A]$ for Langmuir model
4. $\ln[E]$ v/s $\ln[A]$ for Freundlich model
5. $1/[E]^{1/m}$ v/s $1/[A]$ for Combined Langmuir-Freundlich model

In the plots mentioned, $[E]$ is the amount of enzyme present in the solution before adsorption and $[A]$ is the amount of enzyme adsorbed on cotton fibers per gram of cotton fibers. The first plot was used to test whether the linear model was a good fit for the adsorption data of the CBH I, CBH II and EG II. Peitersen et.al [74] and Kim et.al.[81, 82] determined whether adsorption of cellulases on microcrystalline cellulose can be explained by Langmuir model by plotting the second plot. Hence in the present investigation the second plot was used to determine whether the model described by Peitersen et.al.[74] and Kim et.al.[82] can explain the adsorption data. The third plot in the above list determines whether the adsorption data of the enzymes is explained by Langmuir adsorption isotherm [99]. The fourth plot tested the goodness of fit of the Freundlich model and the last plot was plotted to test the combined Langmuir-Freundlich Model for the adsorption of CBH I, CBH II and EG II on cotton fibers.

The R-square values of the linear regression performed were used to determine whether the model was a good fit for the given experimental adsorption data. A R-square value close to 1 indicates a good fit by the model for the given experimental data whereas a R-square value near 0 indicates that the model is not a good fit for the given experimental data.

The results of the adsorption study are discussed separately for CBH I, CBH II and EG II. The adsorption data points for CBH I, CBH II and EG II are the mean values calculated from triplicate experimental measurements.

CBH I:

Figure 5 is a plot of the amount of enzyme adsorbed/ gram of cotton (A) against the enzyme present in the treatment bath (E in mg/ml). An increase in adsorption of CBH I was observed with an increase in concentration of CBH I present in the solution indicating that there are enough sites of adsorption on cotton fibers for CBH I. Increase in adsorption of CBH I with increase in concentration of CBH I in the solution was observed at all treatment temperatures studied, 25, 35, 45 and 55 °C. On plotting the amount of enzyme adsorbed on cotton fiber at different temperatures, as shown in figure 6, it can be seen that temperature of the adsorption experiments influences the adsorption of CBH I onto cotton fibers to a small extent. The

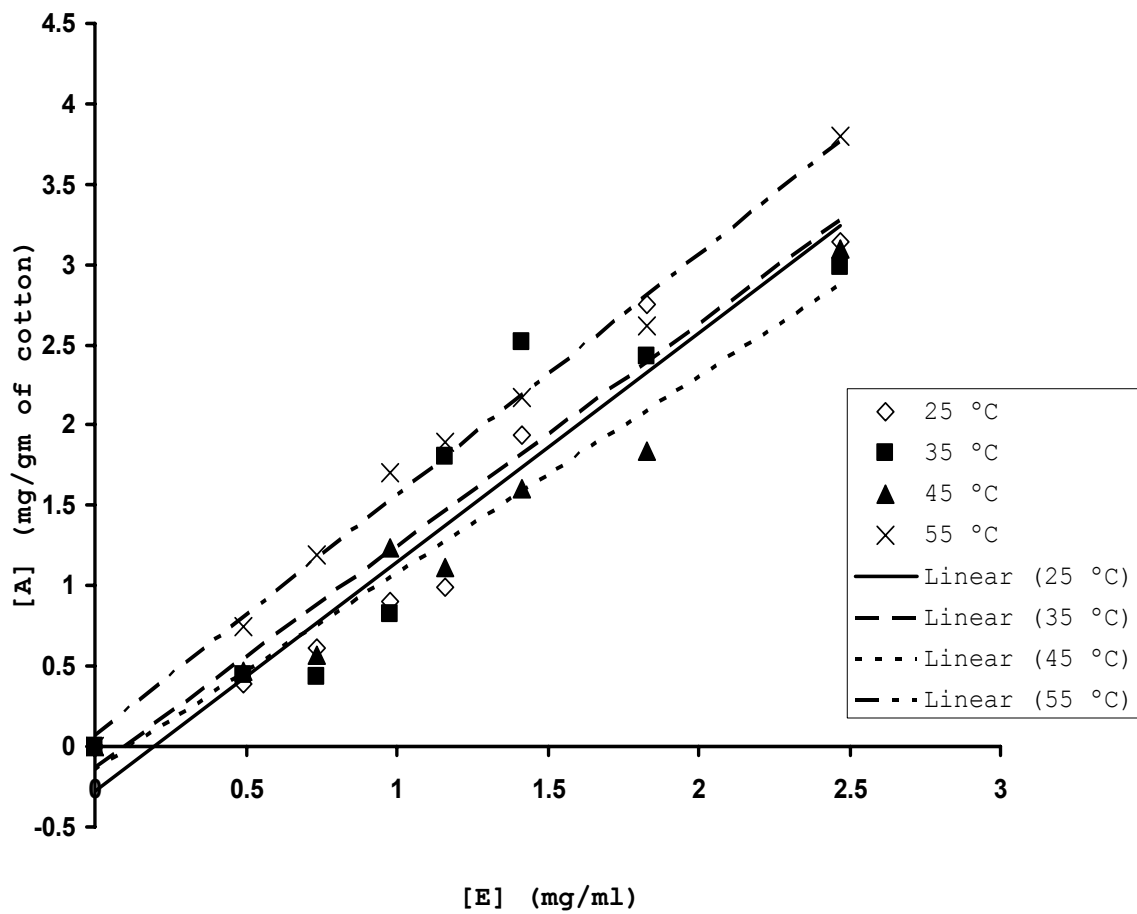


Figure 5. Linear plot for the adsorption isotherms of CBH I on cotton fibers.

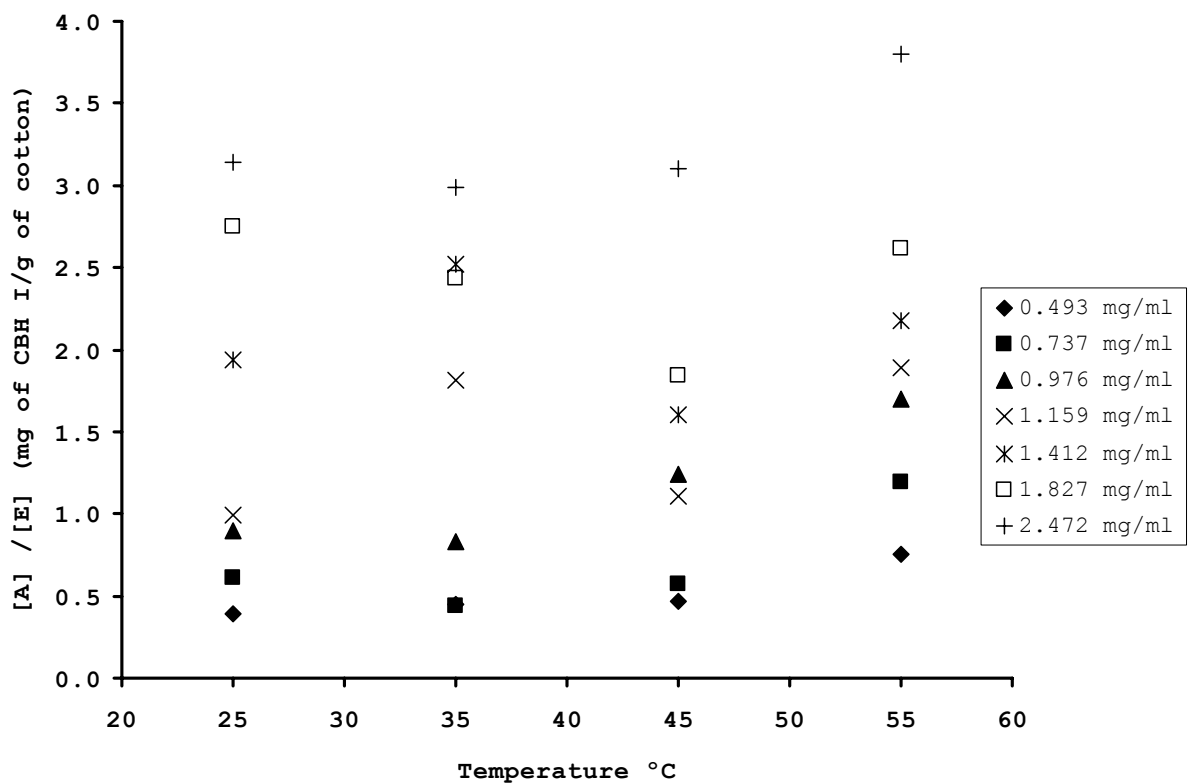


Figure 6. Effect of temperature on adsorption of CBH I on cotton fibers at a given concentration of CBH I

adsorption of CBH I on cotton fibers at 55 °C was higher as compared to the other temperatures studied at all given concentrations of CBH I in the treatment baths.

The ratio of the amount of CBH I adsorbed on cotton fibers [A] and CBH I remaining in the enzyme bath [Es] at different enzyme concentrations was plotted against temperature of adsorption in figure 7. The amount of CBH I adsorbed on cotton fibers at equilibrium is higher at 55 °C than the other adsorption temperatures studied irrespective of the concentration of CBH I in the treatment bath.

Linear trend lines for adsorption of CBH I at each temperature have been plotted in figure 5. Linear regression analysis was performed on the linear adsorption model given by equation 1. The output of the statistical analysis performed by SAS is included as Appendix A. The R-square values for the linear model are listed in Table 4. From the R-square values for the adsorption data at different temperatures, it can be seen that the R-square values are greater than 0.9 except for the adsorption of CBH I on cotton fibers at treatment temperature of 35 °C indicating that the linear adsorption model is a good fit for the experimental adsorption data.

Plot of [E] v/s [E]/ [A]

Studies involving adsorption of cellulases on microcrystalline cellulose used a plot of [E] against [E]/[A] to

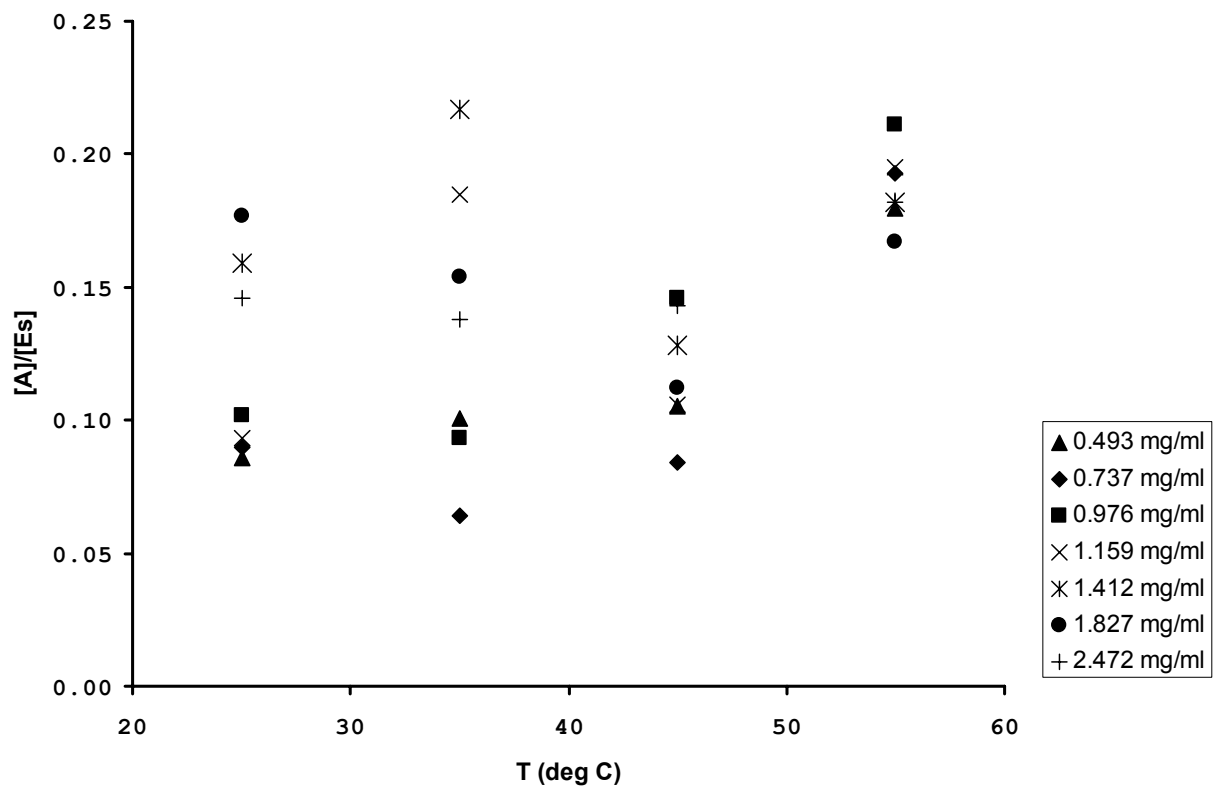


Figure 7. Effect of temperature on equilibrium adsorption of CBH I on cotton fibers at a given concentration of CBH I

Table 4. R-square values for the different plots for the experimental adsorption data of CBH I

Plot	25 °C	35 °C	45 °C	55 °C	Average
[E] v/s [A] (Linear)	0.9422	0.8829	0.9648	0.9917	0.9851
1/[E] v/s 1/[A] (Langmuir)	0.9868	0.8278	0.9349	0.9898	0.9802
[E] v/s [E]/[A] (Langmuir by Kim et.al.[81])	0.6717	0.2899	0.3054	0.1730	0.4928
Ln[E] v/s ln[A] (Freundlich)	0.9639	0.8614	0.9407	0.9877	0.9814
1/[E] ^{1/m} v/s 1/[A] Combined Langmuir- Freundlich)	0.9703	0.8361	0.9277	0.9491	0.9628

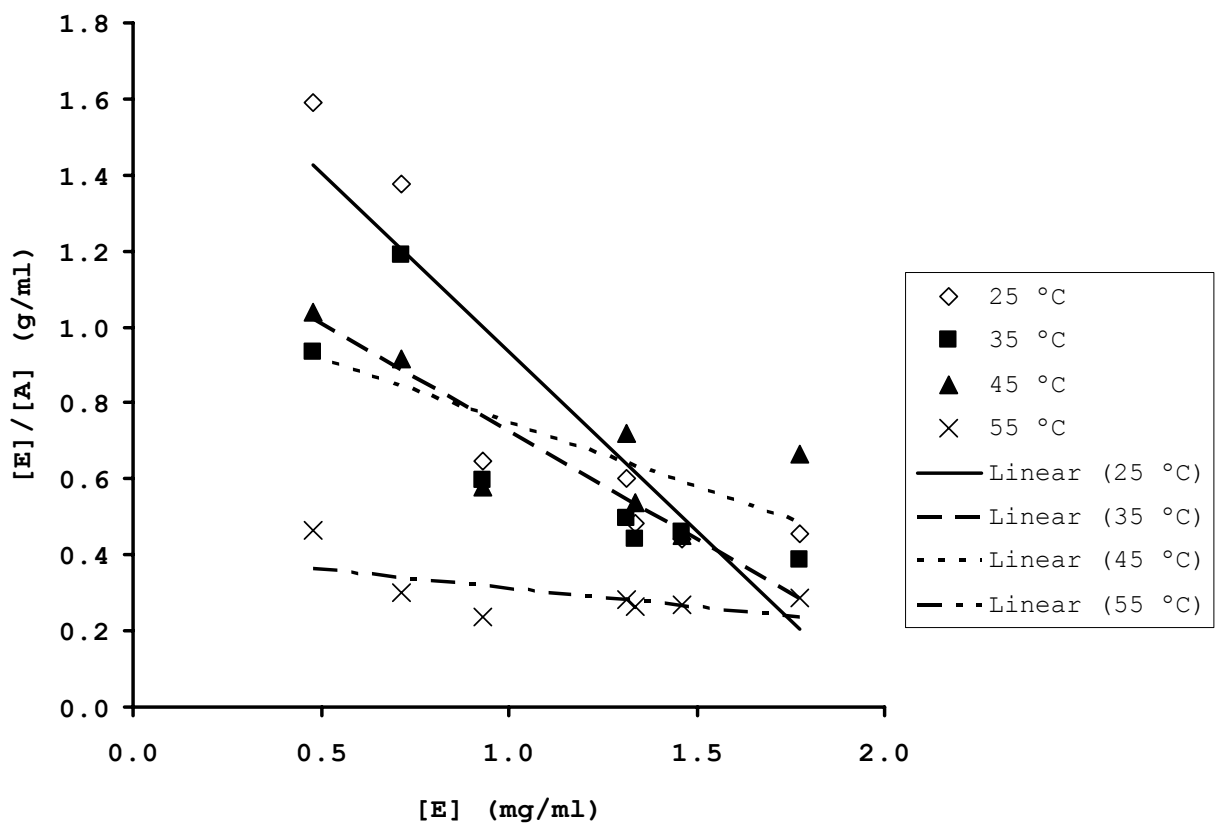


Figure 8. Langmuir plot for the adsorption isotherms of CBH I on cotton fibers (Method I)

determine if the adsorption data was explained by a Langmuir adsorption model [81]. In the present study, $[E]$ was plotted against $[E]/[A]$ was plotted as shown in figure 8. Adsorption data for CBH I on cotton fibers at 25, 35, 45 and 55 °C were plotted. The trend lines for the adsorption data of CBH I at different temperatures are also plotted in the figure 8.

Plot of $1/[E]$ v/s $1/[A]$

Another method to determine whether the adsorption data obtained can be explained by a Langmuir adsorption isotherm is to plot values of $1/[E]$ against $1/[A]$ [99]. A plot of $1/[E]$ against $1/[A]$ is given in figure 9 where $[E]$ is the amount of CBH I present in the bath before adsorption and $[A]$ is the amount of CBH I adsorbed on cotton fibers under the given conditions. Trend lines for the adsorption data at different temperatures are plotted in figure 9.

Plot of $\ln[E]$ v/s $\ln[A]$

Figure 10 shows the plots of natural logarithms of $[E]$ against $[A]$ for the adsorption of CBH I at different temperatures on cotton fibers. The trend lines for adsorption at the different temperatures studied are shown.

Plot of $1/[E]^{1/m}$ v/s $1/[A]$

A plot of $1/[E]^{1/m}$ v/s $1/[A]$ was plotted in order to determine whether the adsorption data can be explained by a combined Langmuir-Freundlich isotherm. The combined

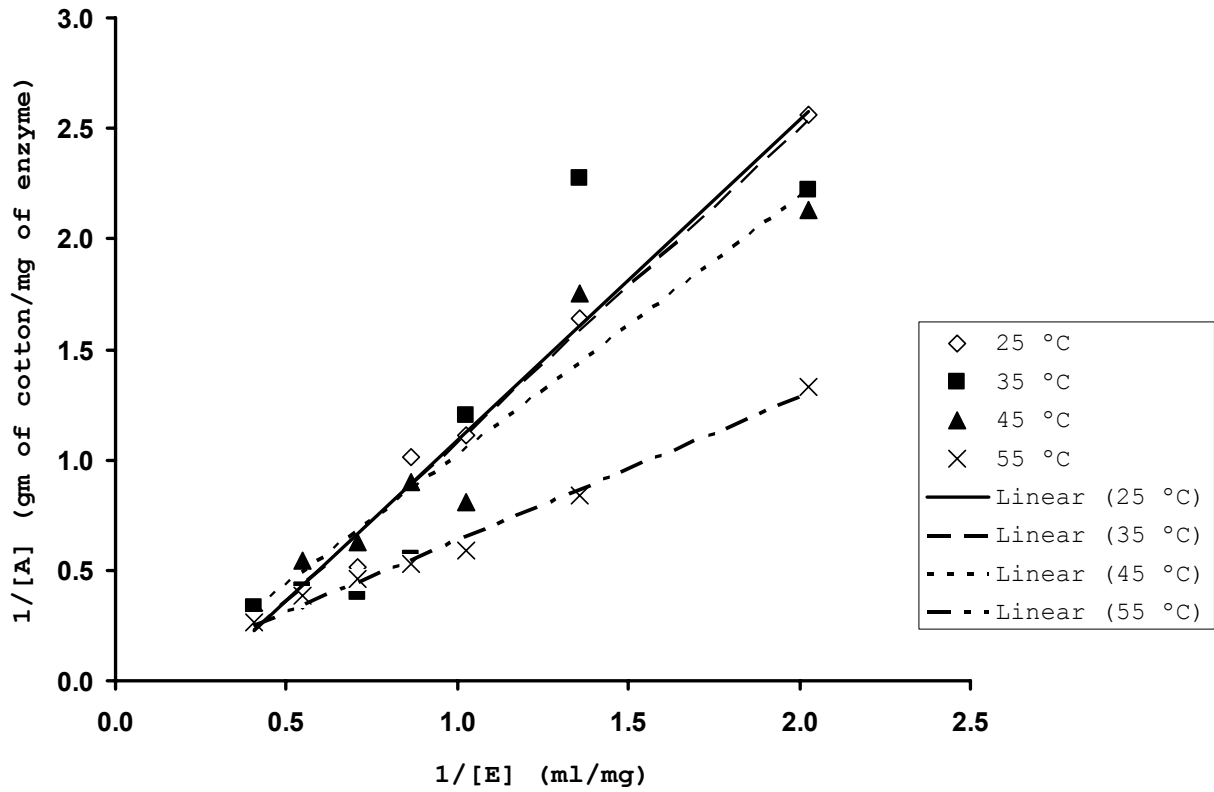


Figure 9. Langmuir plot for the adsorption isotherms of CBH I on cotton fibers (Method II)

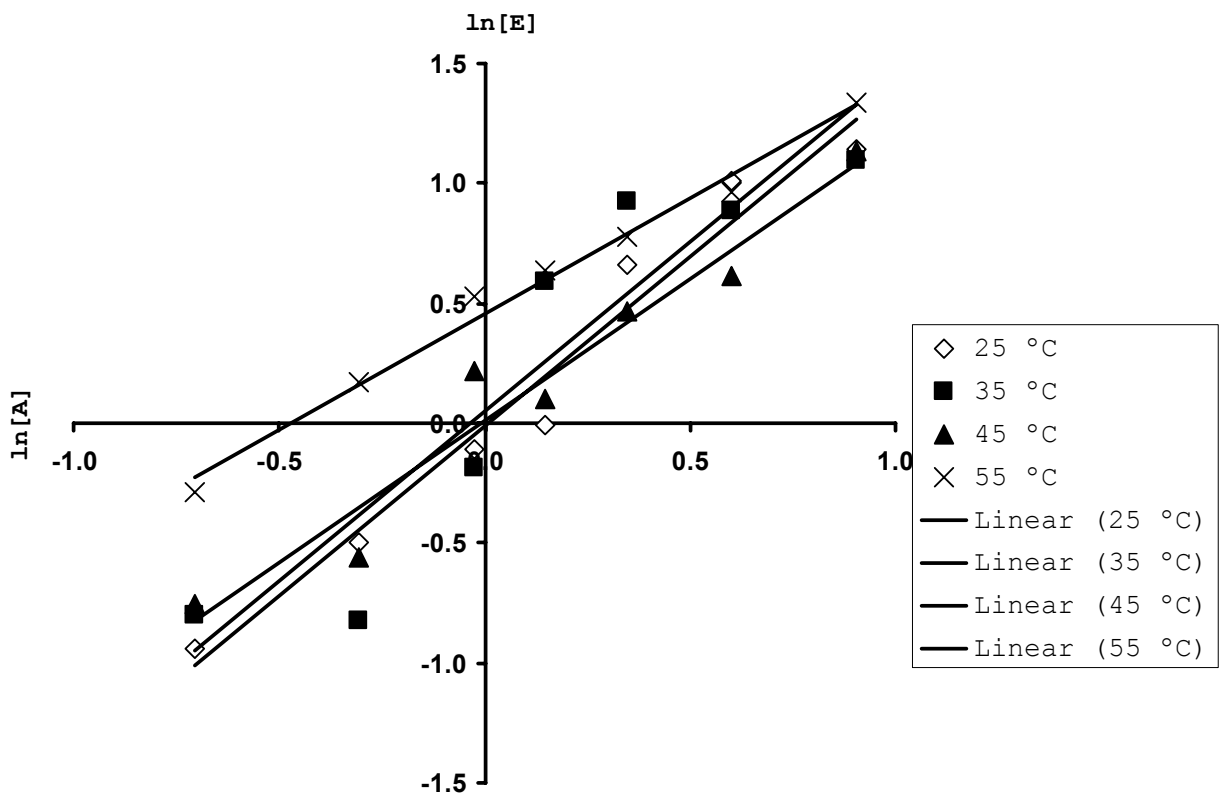


Figure 10. Freundlich plot for the adsorption isotherms of CBH I on cotton fibers

Langmuir-Freundlich isotherm tries to account for the inadequacies of the Langmuir and Freundlich models. The model is empirical and hence the parameters obtained from the model do not have any physical significance. Figure 11 is the plot of the values of $1/[E]^{1/m}$ against $1/[A]$ where $m=2$.

Linear regression was conducted on the adsorption data of CBH I to determine which of the adsorption models fit the experimental adsorption data. The R-square values for the different regressions models are listed in Table 4. The R-square values for the different models were used to determine if the model is a good fit of the present adsorption data. The regression analysis output for adsorption of CBH I at different temperatures is given as Appendix A.

For the linear adsorption model, the R-square values listed in Table 4 were found to be greater than 0.9 except for the plot at 35 deg C indicating that the linear adsorption model is a good fit for the experimental adsorption data. Hence the linear adsorption model could be used to describe the adsorption behavior of CBH I on cotton fibers within the given concentration range and experimental conditions.

The R-square values for Langmuir adsorption model based on the plots of $[E]$ against $[E]/[A]$ listed in Table 4 are found to be close to zero on a scale of 0 to 1 implying that the Langmuir adsorption model as described by Kim et.al.[81] is not a good

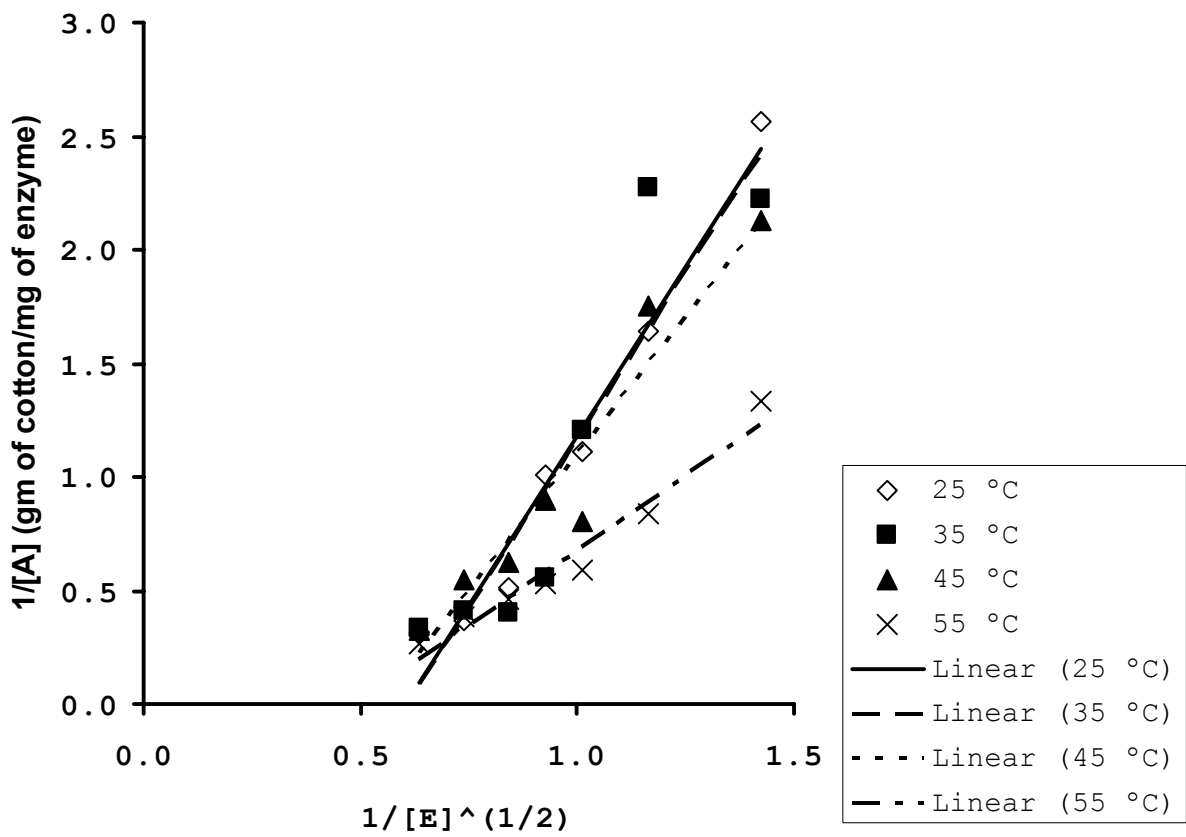


Figure 11. Combined Langmuir-Freundlich plot for the adsorption isotherms of CBH I on cotton fibers

fit for the experimental adsorption data of CBH I. Hence the present adsorption data cannot not be explained by Langmuir adsorption model described by Kim et. al. [81] based on the plots of $[E]$ against $[E]/[A]$. When linear regression was conducted using plots of $1/[E]$ against $1/[A]$, it was found that the R-square values were closer to 1 indicating that the Langmuir adsorption model is a good fit for the adsorption data of CBH I. Hence, the present experimental adsorption data for CBH I can be explained by Langmuir adsorption model based on the plots of $1/[E]$ against $1/[A]$.

The Freundlich adsorption model can be said to be a good fit for the given experimental adsorption data since the linear regression of $\ln[E]$ against $\ln[A]$ gave R-square values in the range of 0.8614 to 0.9877 for the four temperatures at which adsorption experiments were conducted.

The R-square values for the combined Langmuir-Freundlich model with $m=2$ are listed in Table 4. The values for the adsorption data at 25, 45 and 55 °C are greater than 0.9 whereas the R-square value for the adsorption data for 35 °C is 0.8361 indicating that the combined Langmuir-Freundlich model could be used to describe the adsorption data of CBH I.

In the adsorption studies conducted by Kim et al and Peiterson et al [74] [82], the authors tried to determine whether the experimental adsorption data could be explained by

Langmuir adsorption data alone. In the present study, the experimental adsorption data was tested for more than one type of adsorption models. Kim et al, [82] reported that the adsorption of CBH I on microcrystalline cellulose was explained by Langmuir adsorption isotherm. However, in the present data, though the range of enzyme concentration used in the present study is similar, the adsorption data could not be explained by the Langmuir adsorption model as employed by Kim et.al.[82] and Peitersen et.al [74], i.e. plot a graph of $[E]$ against $[E]/[A]$. But when a plot of the reciprocals of $[E]$ and $[A]$ were plotted, the adsorption data could be explained by Langmuir adsorption model. The experimental data is explained by linear, Freundlich and combined Langmuir-Freundlich models.

The average of the values of adsorption of CBH I at each given concentration on cotton fibers at different temperatures was plotted and linear regression analysis was conducted to determine the best fitting model for the data. In Table 4, it can be seen that the R-square values for all the models are greater than 0.9 except for the Langmuir model when tested by plotting $[E]$ against $[E]/[A]$ (model described by Kim et.al.[81]). R-square values close to 1 for more than one model indicate that the experimental adsorption data could be explained by different models at the given range of concentration.

CBH II:

Plot of [E] v/s [A]

Figure 12 is a plot of the amount of CBH II adsorbed per gram of cotton against the amount of CBH II present in the original solution. The amount of CBH II adsorbed on the cotton fibers increases with the concentration of CBH II initially present in the treatment bath. The adsorption of CBH II at the different temperatures studied, 25, 35, 45 and 55 °C was higher at 55 °C as compared to the adsorption at the other adsorption temperatures studied. Figure 13 shows the effect of temperature on adsorption of CBH II at a given concentration of CBH II. Linear trend lines are shown in each figure for adsorption of CBH II from solutions of different concentrations at each treatment temperature.

Figure 14 is a plot of the ratio of amount of CBH II adsorbed on cotton fibers [A] and the amount of CBH II remaining in the enzyme bath [Es] after adsorption at different enzyme concentrations against temperature of adsorption. From figure 14 it is evident that the amount of CBH II adsorbed on cotton fibers at equilibrium is higher at 55 °C compared to other treatment temperatures studied irrespective of the concentration of CBH II.

Plot of [E] v/s [E]/ [A]

Figure 15 is a plot of the amount of CBH II present in

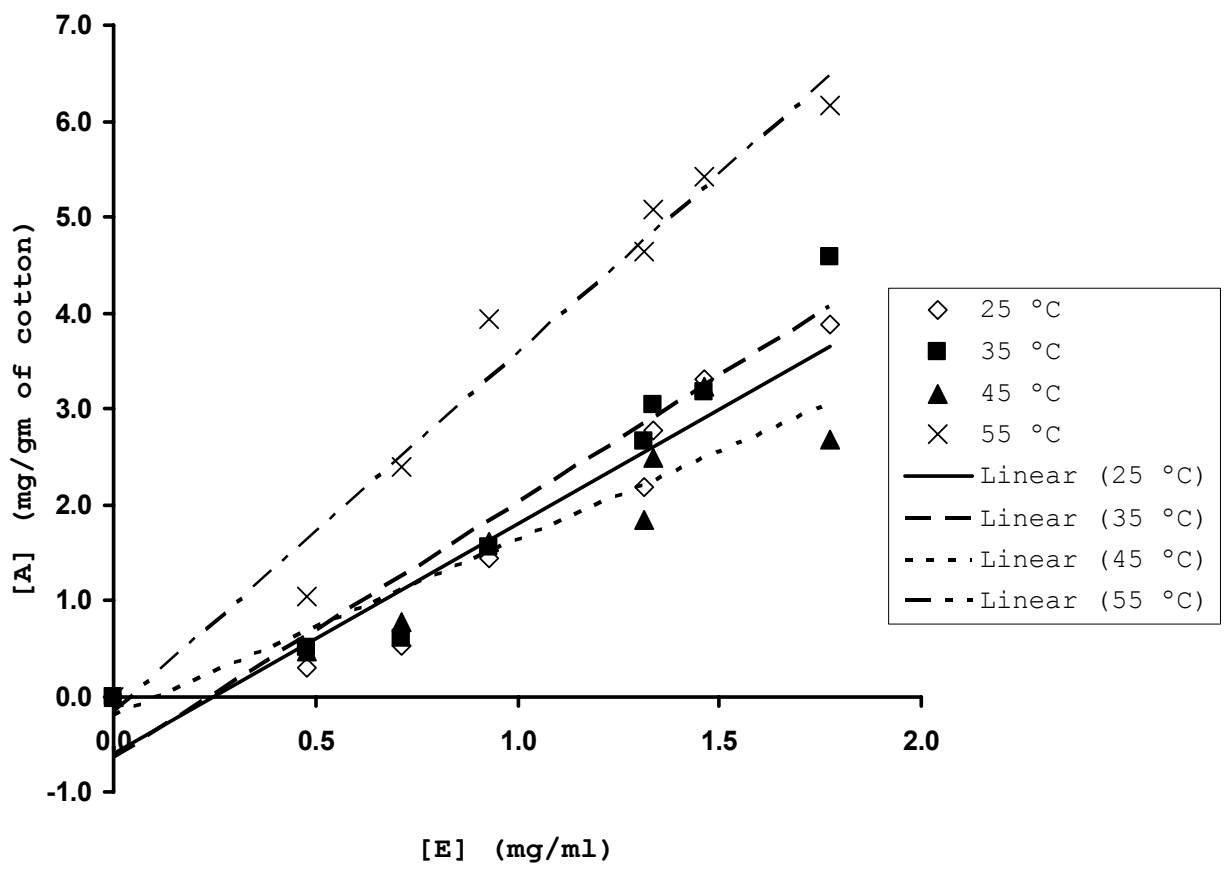


Figure 12. Linear plot for the adsorption isotherms of CBH II on cotton fibers.

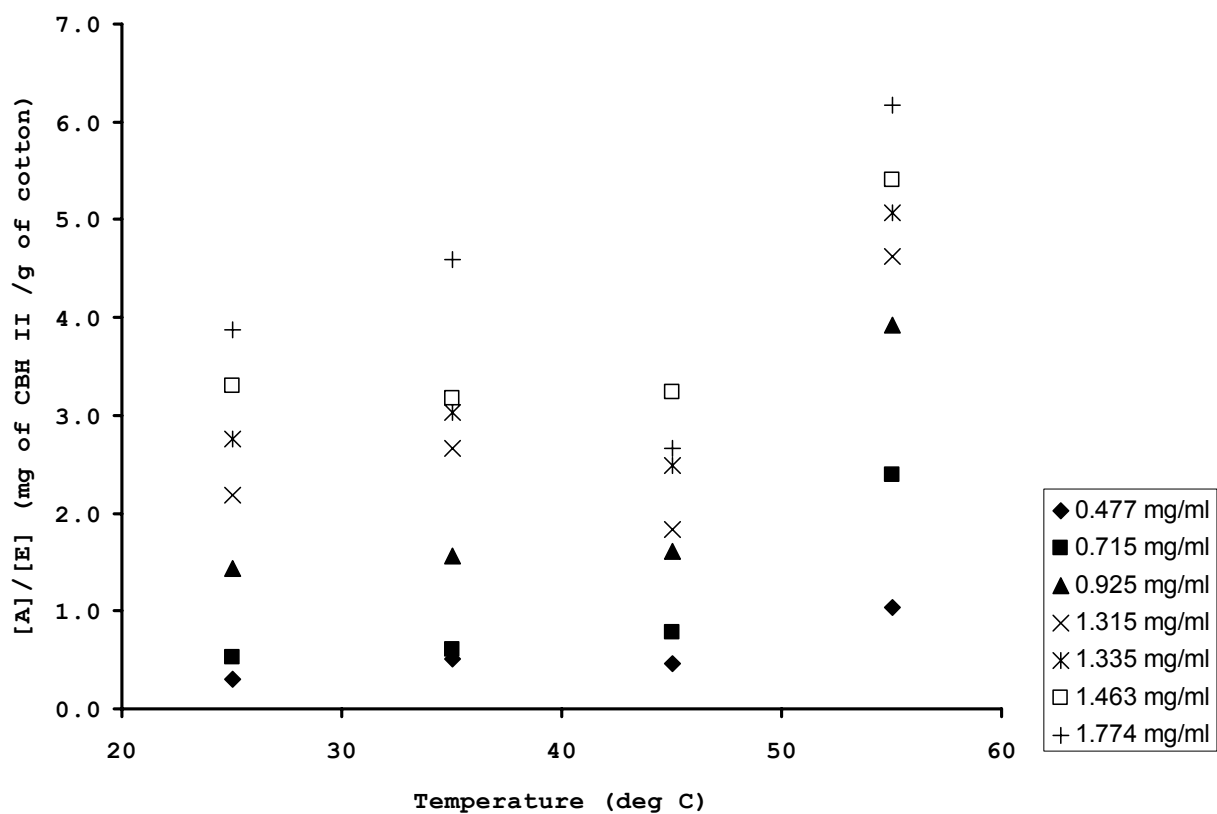


Figure 13. Effect of temperature on adsorption of CBH II on cotton fibers at a given concentration of CBH II

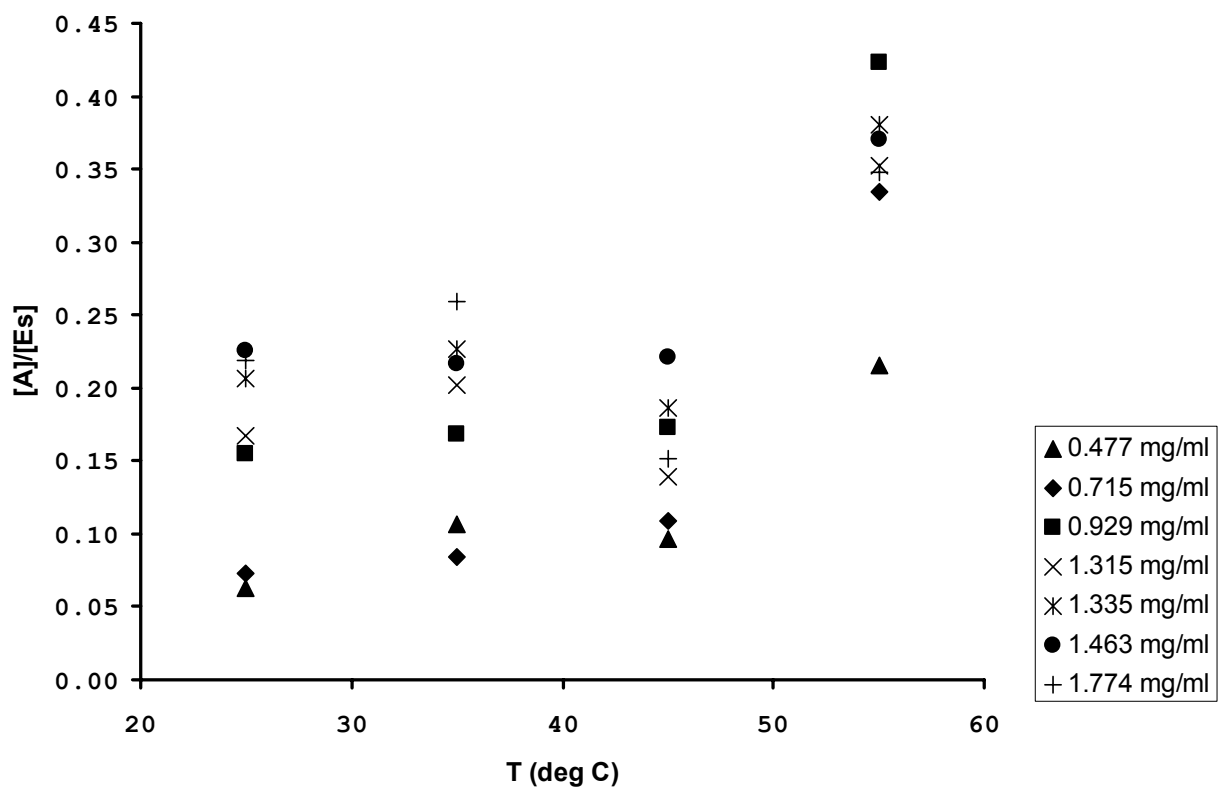


Figure 14. Effect of temperature on equilibrium adsorption of CBH II on cotton fibers at a given concentration of CBH II

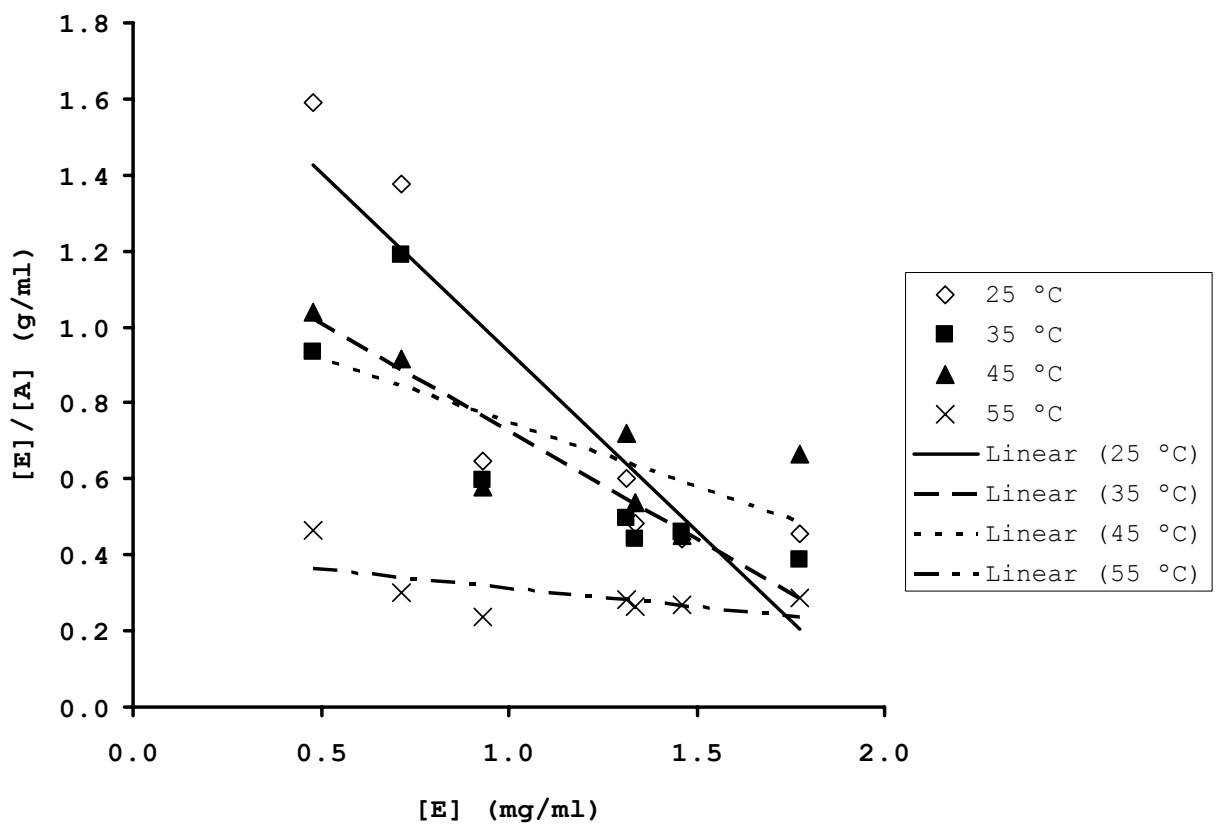


Figure 15. Langmuir plot for the adsorption isotherms of CBH II on cotton fibers (Method I)

solution against the ratio of the amount of CBH II present in solution to the amount of CBH II adsorbed per gram of cotton fibers. Plotting $[E]$ against $[E]/[A]$ is the method used by Kim et.al. [81] to determine whether the experimental adsorption data is explained by the Langmuir adsorption model.

Plot of $1/[E]$ v/s $1/[A]$

Figure 16 is a plot of the reciprocal of the amount of CBH II present in solution against the amount of CBH II adsorbed per gram of cotton fibers. By plotting the reciprocals of the $[E]$ and $[A]$, we can determine whether the experimental adsorption data of CBH II can be explained by the Langmuir adsorption model.

Plot of $\ln[E]$ v/s $\ln[A]$

Figure 17 is the log-log plot of amount of CBH II present in solution against the amount of CBH II adsorbed per gram of cotton fibers. The plots of natural logarithm values of $[E]$ and $[A]$ are plotted to determine if the experimental adsorption data can be explained by the Freundlich adsorption isotherm.

Plot of $1/[E]^{1/m}$ v/s $1/[A]$

Figure 18 is the plot of inverse of the amount of CBH II present in solution to the power of $1/m$ (where $m=2$) against the inverse of the amount of CBH II adsorbed per gram of cotton fibers. The linear regression output of the different models for the experimental adsorption data of CBH II is given as

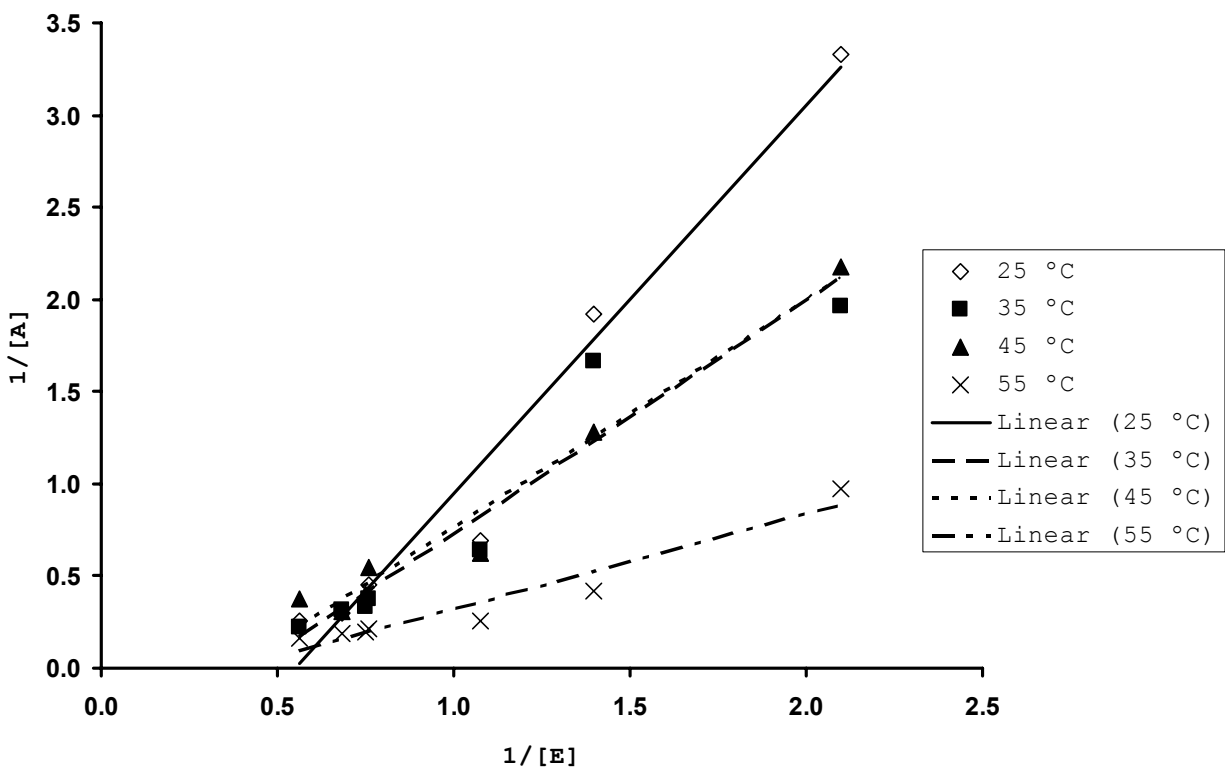


Figure 16. Langmuir plot for the adsorption isotherms of CBH II on cotton fibers (Method II)

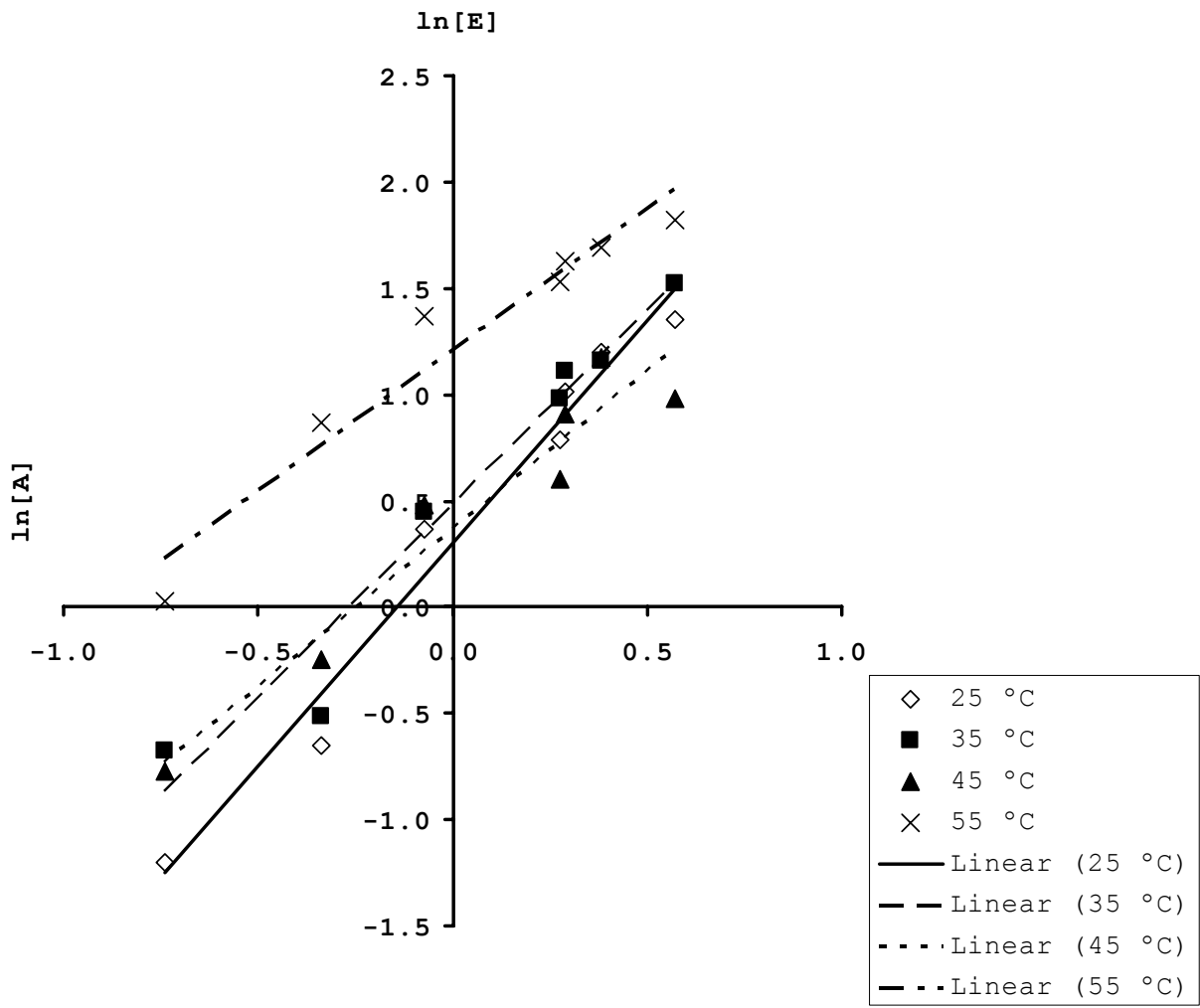


Figure 17. Freundlich plot for the adsorption isotherms of CBH II on cotton fibers

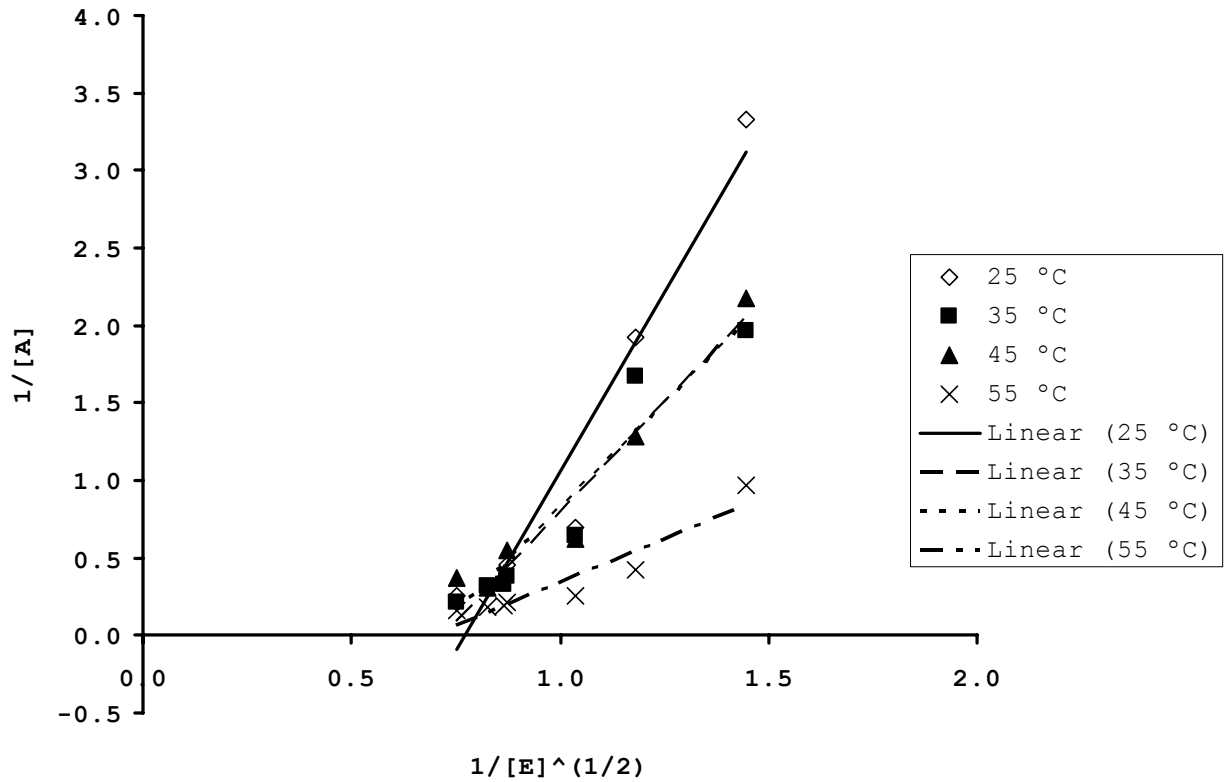


Figure 18. Combined Langmuir-Freundlich plot for the adsorption isotherms of CBH II on cotton fibers

Appendix B.

The R-square values for the regression analysis for the model used to fit the adsorption data for CBH II on cotton fibers are given in Table 5. The R-square values for the linear adsorption model are found to be greater than 0.9 except for adsorption at 45 °C, implying that the linear adsorption model is a good fit for the experimental adsorption data obtained at different temperatures of treatment of cotton fibers with CBH II.

The R-square values of the regression analysis for plots of $[E]$ against $[E]/[A]$ were in the range of 0.5387 and 0.8025 except for the plot of adsorption points at 55°C which was 0.3596 (Table 5). From the R-square values the Langmuir adsorption isotherm as described by Peitersen et.al. [74] and Kim et.al. [81], does not explain the experimental adsorption data. In contrast, plotting the reciprocal values of $[E]$ and $[A]$ resulted in R-square values greater than 0.9 at all treatment temperatures and indicated that the experimental adsorption data can be described by Langmuir adsorption isotherm.

The R-square values for the Freundlich plot, which is a natural logarithmic plot of $[E]$ and $[A]$, are the highest among the different models. The R-square values for the adsorption of CBH II at all the temperatures studied were found to be greater

Table 5. R-square values for the different plots of the experimental adsorption data of CBH II

Plot	25 °C	35 °C	45 °C	55 °C	Average
[E] v/s [A] (Linear)	0.9213	0.9303	0.8786	0.9723	0.9621
1/[E] v/s 1/[A] (Langmuir)	0.9689	0.9170	0.9660	0.9274	0.9729
[E] v/s [E]/[A] (Langmuir by Kim et.al. [81])	0.8025	0.7259	0.5387	0.3596	0.7228
Ln[E] v/s ln[A] (Freundlich)	0.9714	0.9543	0.9280	0.9402	0.9722
1/[E] ^{1/m} v/s 1/[A] Combined Langmuir- Freundlich)	0.9431	0.9255	0.9379	0.8793	0.9406

than 0.9. Hence the Freundlich adsorption model appears to be a good fit of the experimental adsorption data. Similarly, the R-square values for the adsorption plots of CBH II at the different temperatures are greater than 0.9 except the adsorption data at 55 °C, which was 0.8793 and is still acceptable. Hence the Combined Langmuir-Freundlich adsorption model with $m=2$ is a good fit for the experimental adsorption data.

The experimental adsorption values of CBH II at different concentrations were plotted to determine the effect of increasing temperature on adsorption of CBH II as shown in figure 12. From the figure, it could be said that temperature did not affect the adsorption of CBH II on cotton fibers at a given concentration except at 55 °C at which higher adsorption of CBH II on cotton fibers was observed at all given concentrations. The average values of adsorption values of CBH II on cotton fibers at different temperatures were then plotted. From the R-square values for the regression analysis using the average values (shown in table 5) the R-square values were greater than 0.9 for all plots except for the plot of $[E]$ against $[E]/[A]$ which is Peitersen et.al. [74], and Kim et.al's [82] method to determine whether the Langmuir adsorption model could be used to explain the experimental adsorption data. Hence, from the R-square values, that the experimental

adsorption data for CBH II on cotton fibers could be explained by more than one adsorption model.

EG II:

EG II was the third enzyme that was evaluated in this study. The adsorption studies were performed and the experimental adsorption data obtained was then subjected to linear regression. The output of the linear regression is included in the dissertation as Appendix C.

Plot of [E] v/s [A]

The amount of EG II adsorbed per gram of cotton plotted against the amount of EG II initially present in the solution indicates an increasing adsorption of EG II on cotton fibers with increasing concentration of EG II in solution (Figure 19). A graph of EG II adsorbed per gram of cotton for individual concentrations with increasing temperatures is shown in figure 20. This plot shows that the adsorption of EG II is not affected by temperature at low concentrations. At higher concentrations of EG II (1.557, 1.857 and 2.069 mg/ml), an increase in the adsorption of EG II on cotton fibers was observed with increase in temperature of treatment.

Figure 21 is a plot of the ratio of amount of CBH II adsorbed on cotton fibers [A] and the amount of EG II remaining in the enzyme bath [Es] after adsorption at different enzyme

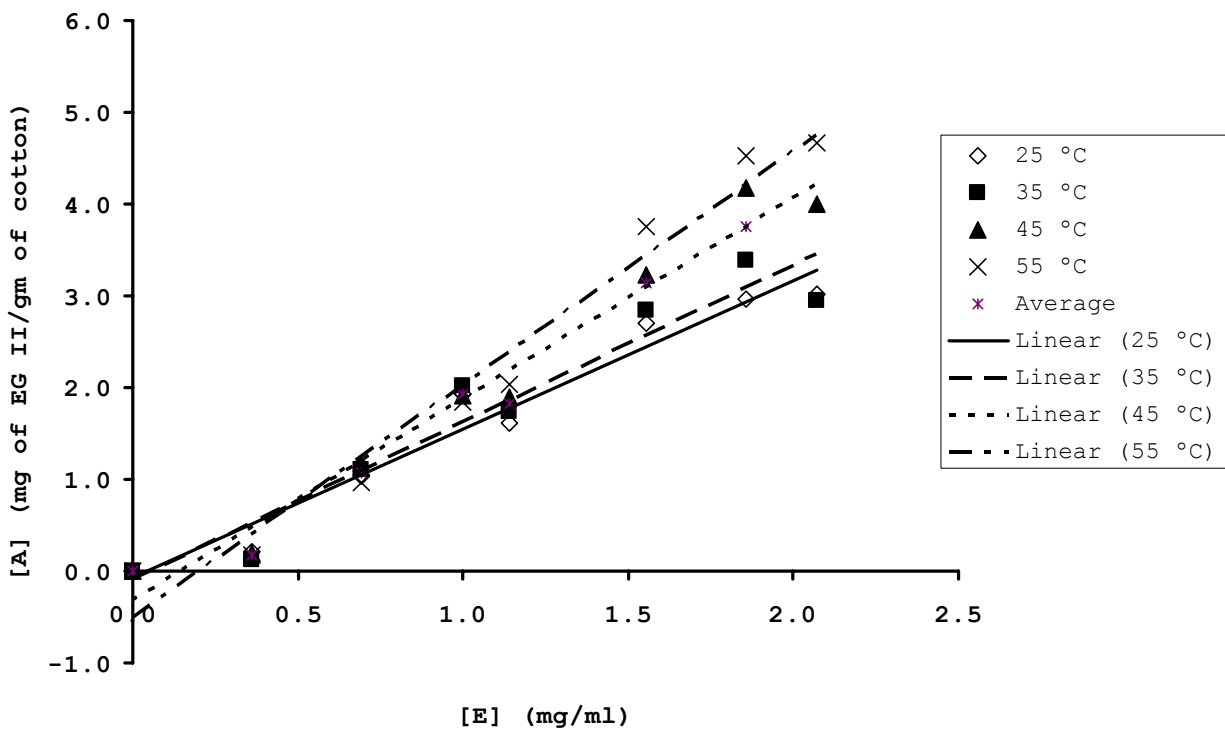


Figure 19. Linear plot for the adsorption isotherms of EG II on cotton fibers.

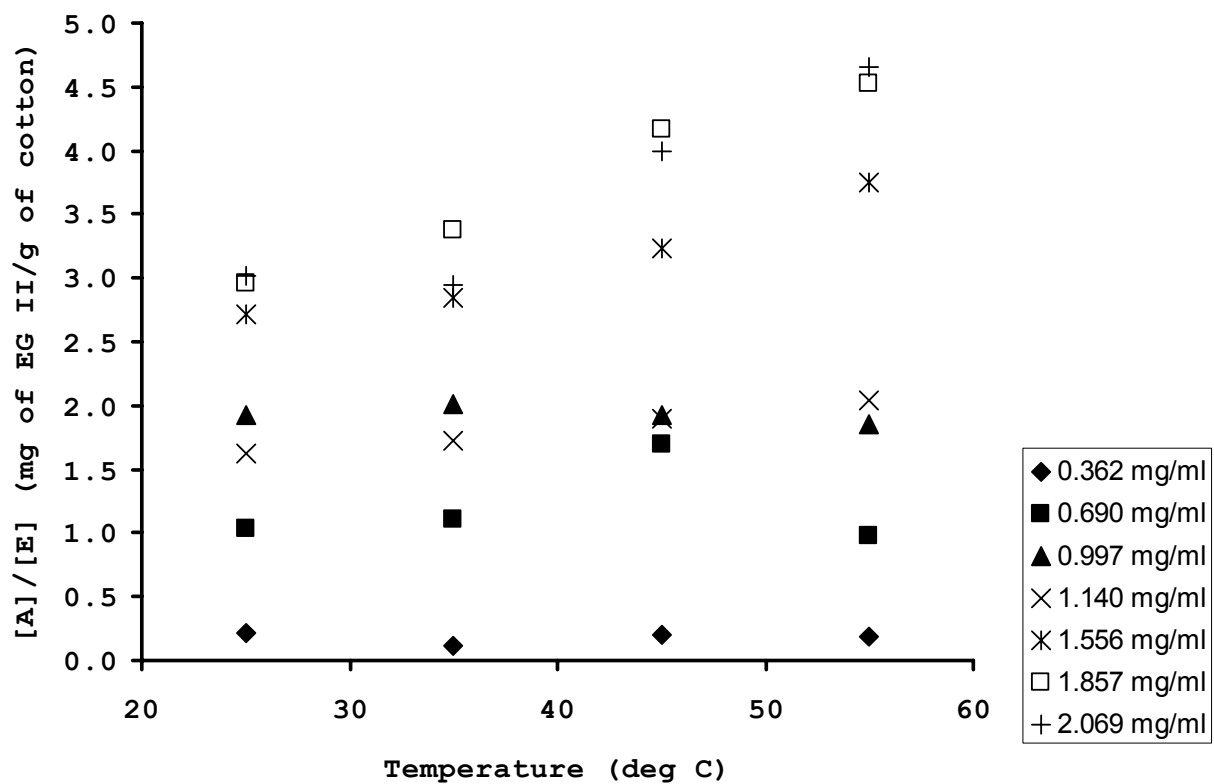


Figure 20. Effect of temperature on adsorption of EG II on cotton fibers at a given concentration of EG II

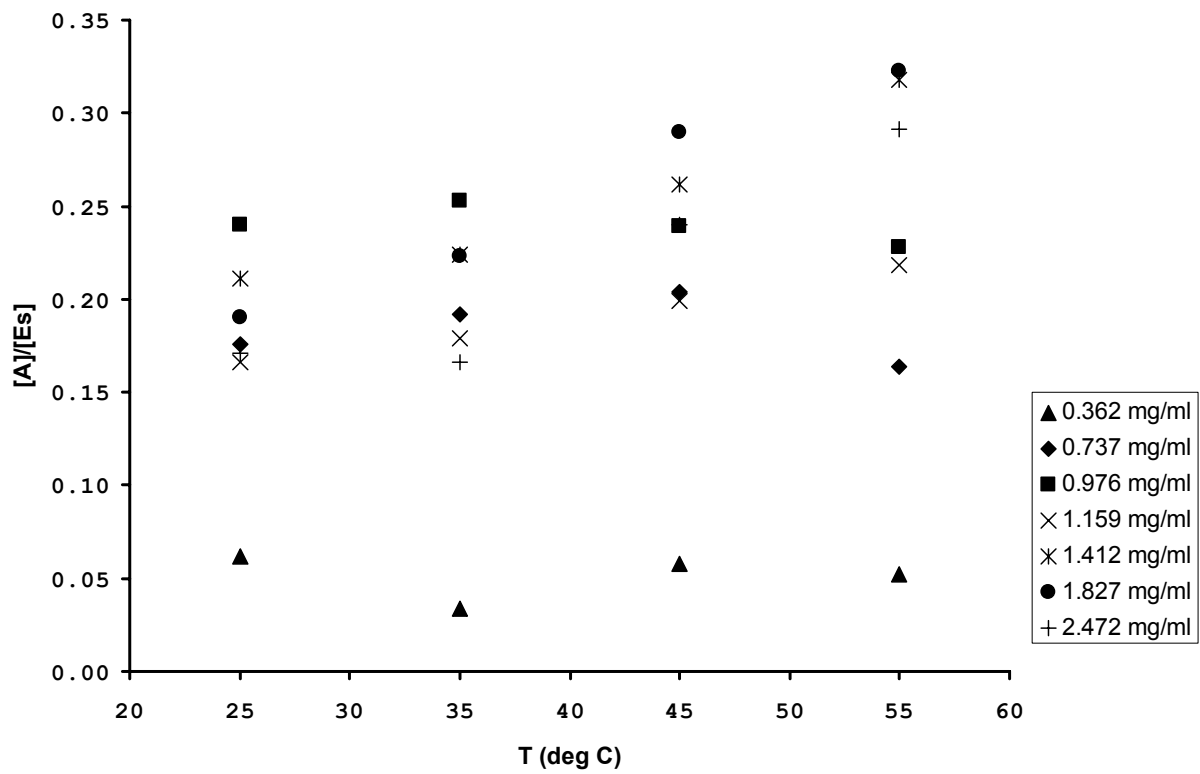


Figure 21. Effect of temperature on equilibrium adsorption of EG II on cotton fibers at a given concentration of EG II

concentrations against temperature of adsorption. From the figure 21 it is evident that the amount of EG II adsorbed on cotton fibers at equilibrium increases with the increase in temperature of adsorption. The increase in ratio, $[A]/[E_s]$ was higher at higher concentrations of EG II present in the adsorption bath.

Plot of $[E]$ v/s $[E]/[A]$

Figure 22 is a plot of the amount of EG II present in solution against the ratio of the amount of EG II present in solution to the amount of EG II adsorbed per gram of cotton fibers. This plot used by Peitersen et.al. [74] and Kim et.al.[81] to model the adsorption of cellulases on microcrystalline cellulose was used to determine the goodness of fit of the Langmuir model for the adsorption of EG II on cotton.

Plot of $1/[E]$ v/s $1/[A]$

Figure 23 is a plot of the inverse of the amount of EG II present in solution against the amount of EG II adsorbed per gram of cotton fibers. The plots of $[E]$ against $[E]/[A]$ and the reciprocal plots of $[E]$ against $[A]$ are the two methods employed to determine whether the experimental adsorption data can be explained by the Langmuir adsorption model.

Plot of $\ln[E]$ v/s $\ln[A]$

Figure 24 is the natural logarithmic plot of the amount of EG II present in solution against the amount of EG II adsorbed

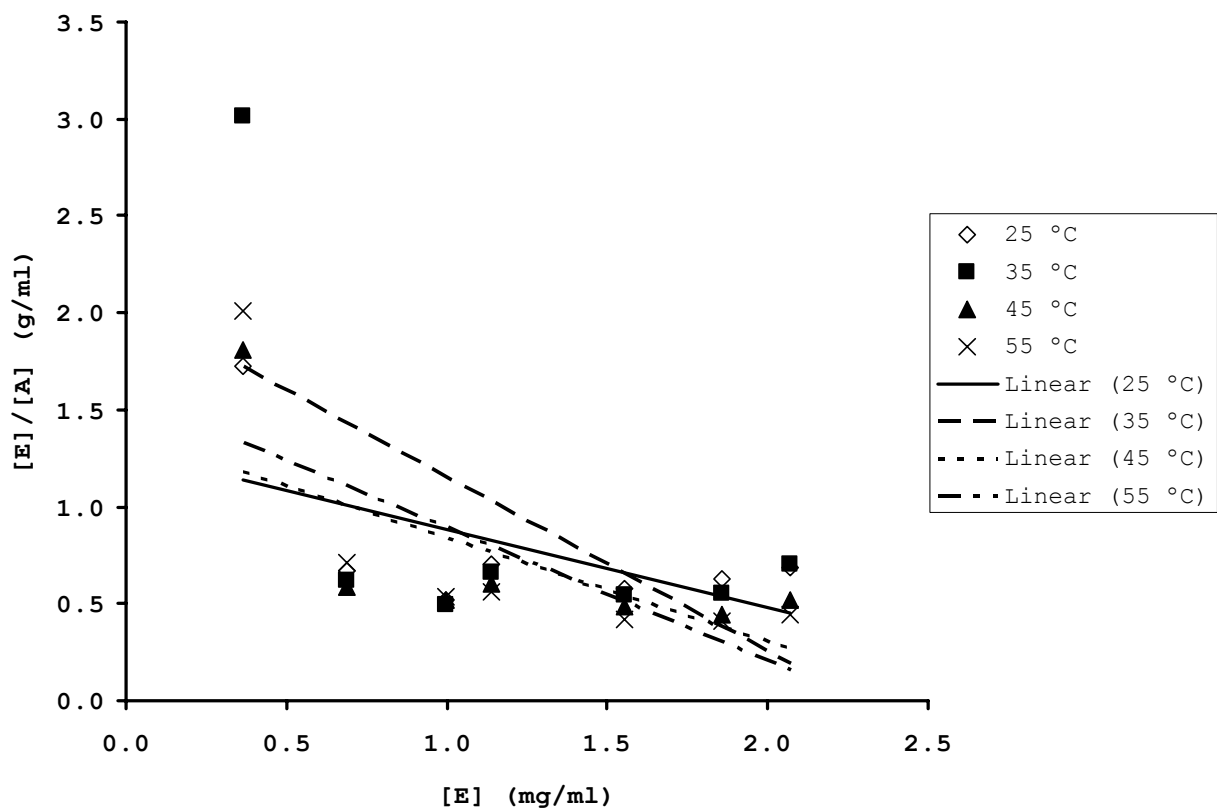


Figure 22. Langmuir plot for the adsorption isotherms of EG II on cotton fibers (Method I)

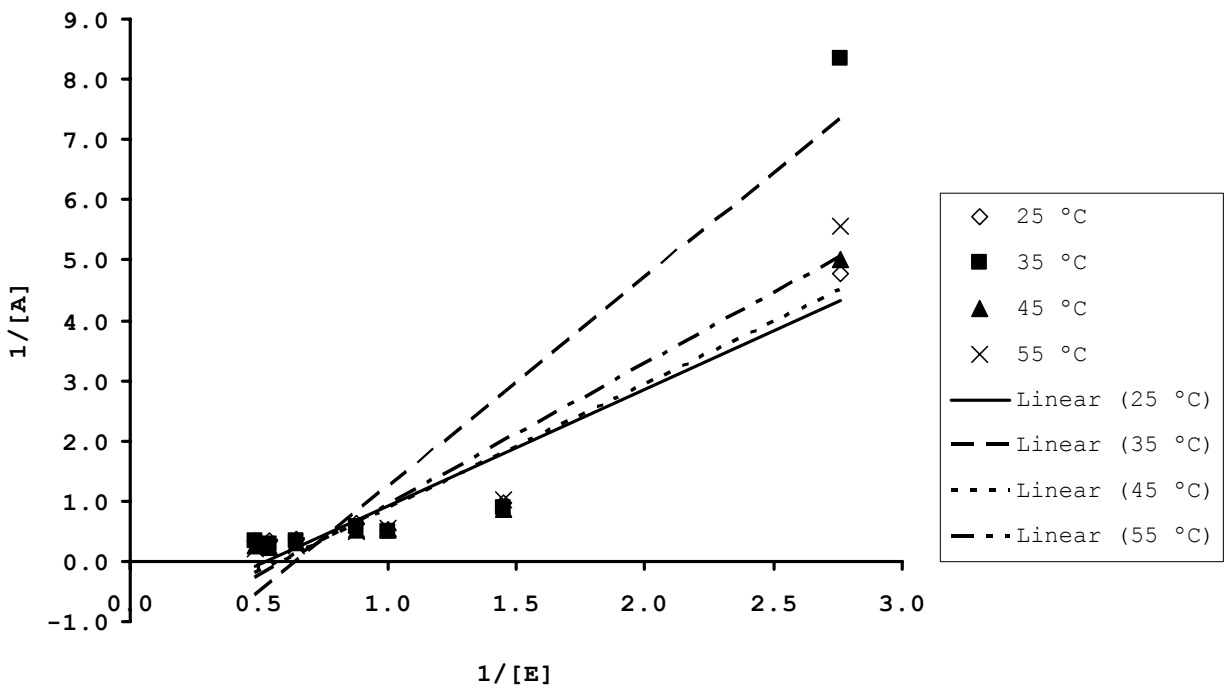


Figure 23. Langmuir plot for the adsorption isotherms of EG II on cotton fibers (Method II)

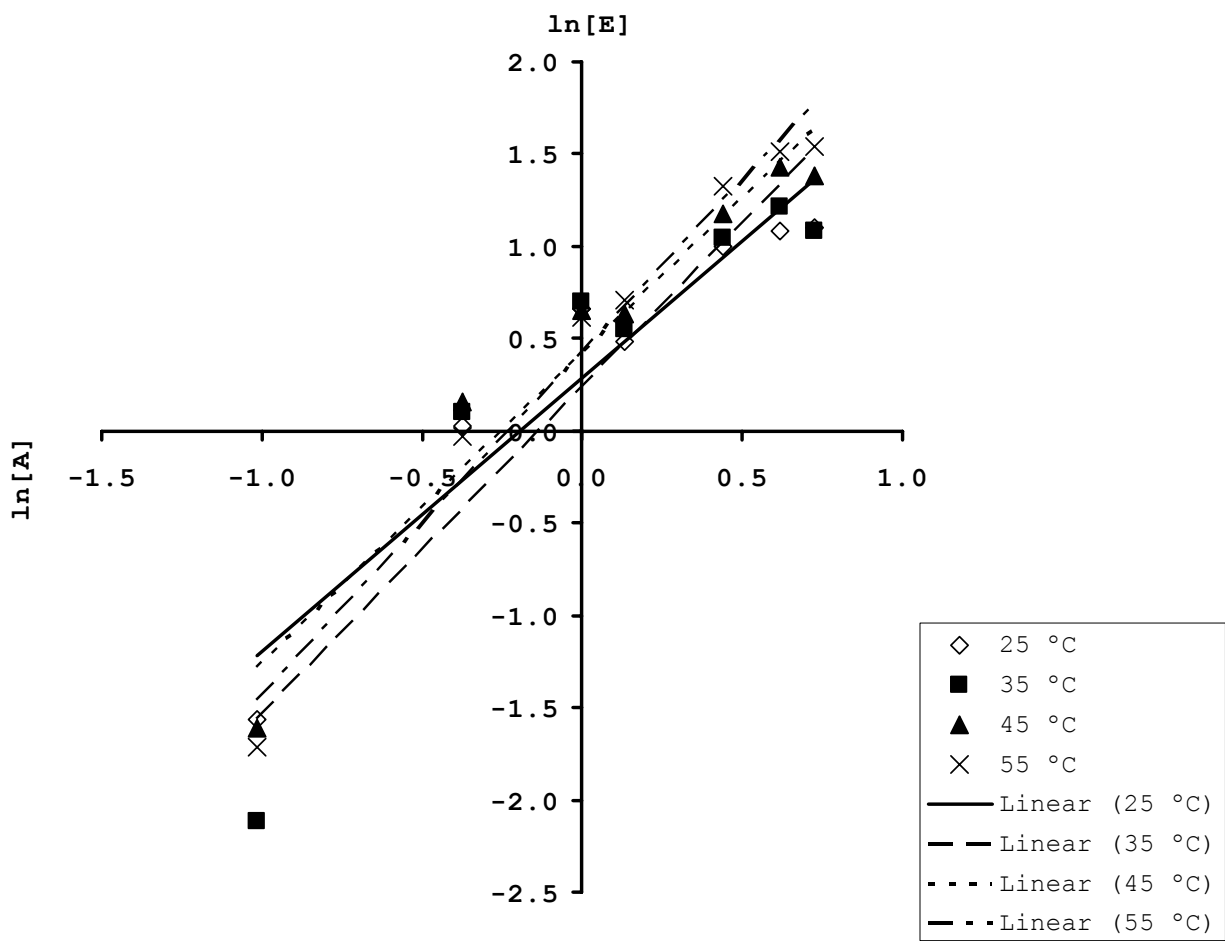


Figure 24. Freundlich plot for the adsorption isotherms of EG II on cotton fibers

per gram of cotton fibers. Plot of natural logarithms of [E] and [A] were made in order to determine whether the experimental adsorption data can be explained by Freundlich adsorption model. Plot of $1/[E]^{1/m}$ v/s $1/[A]$

Figure 25 is the plot of inverse of the amount of EG II present in solution to the power of $1/m$ where $m=2$ against the inverse of amount of EG II adsorbed per gram of cotton fibers. The plot of $1/[E]^{1/m}$ v/s $1/[A]$ was made in order to determine whether the experimental adsorption data can be explained by the combined Langmuir-Freundlich adsorption model.

The R-square values of the linear regressions for the different models are listed in Table 6. The R-square values for the linear adsorption model were found to be greater than 0.9, ranging from 0.9329 to 0.9722. Values of R-square closer to 1 indicate that the linear adsorption model is a good fit of the experimental adsorption data for EG II.

The R-square values for the plot of [E] against $[E]/[A]$ are found to be closer to 0 ranging from 0.3556 to 0.5461. From the R-square values, it may be said that the Langmuir adsorption model used by Peitersen et.al.[74] and Kim et.al. [81], may not be a good fit of the experimental adsorption data as per method I. But when the plots of the reciprocals of [E] and [A] were plotted, linear regression yielded R-square values that were closer to 1 indicating that Langmuir adsorption model

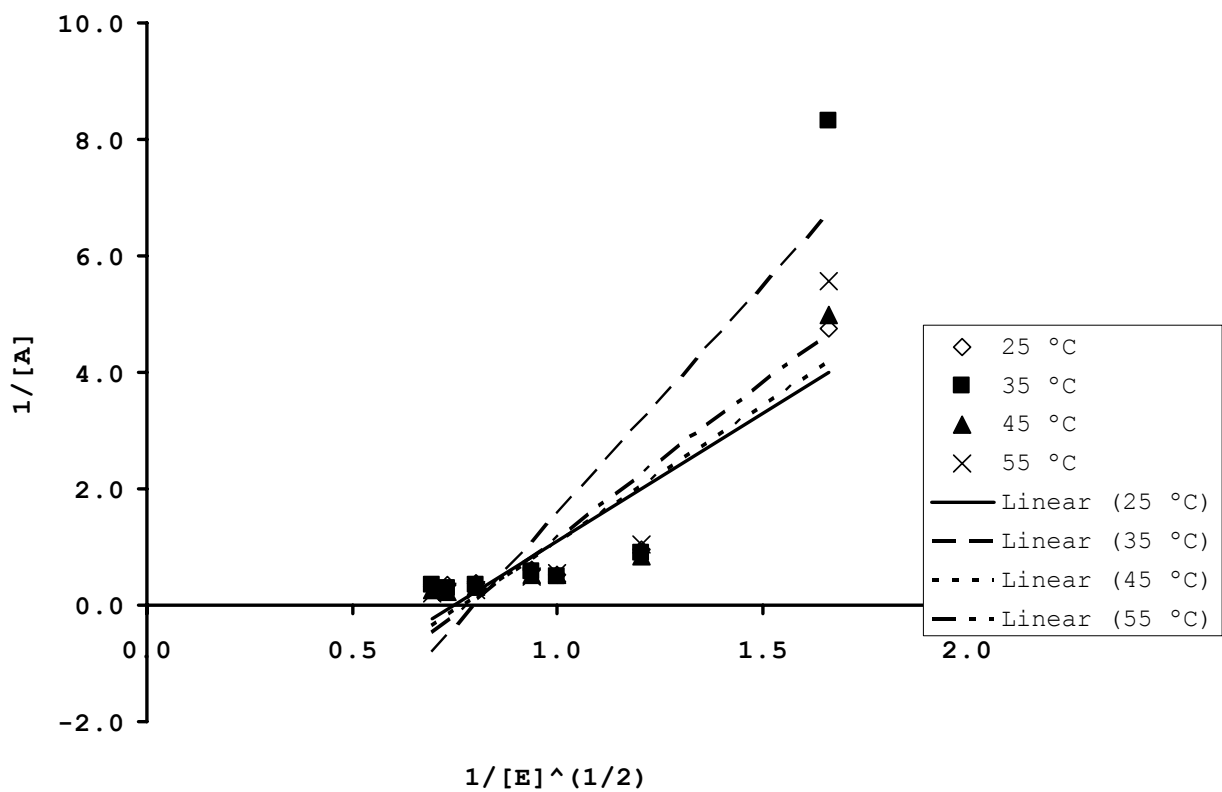


Figure 25. Combined Langmuir-Freundlich plot for the adsorption isotherms of EG II on cotton fibers

Table 6. R-square values for the different plots for the experimental adsorption data of EG II

Plot	25 °C	35 °C	45 °C	55 °C	Average
[E] v/s [A] (Linear)	0.9591	0.9329	0.9722	0.9697	0.9697
1/[E] v/s 1/[A] (Langmuir)	0.9164	0.8765	0.9112	0.9231	0.9078
[E] v/s [E]/[A] (Langmuir by Kim et.al.[81])	0.3589	0.3682	0.3556	0.5461	0.4430
Ln[E] v/s ln[A] (Freundlich)	0.9205	0.8735	0.9467	0.9719	0.9373
1/[E] ^{1/m} v/s 1/[A] Combined Langmuir- Freundlich)	0.8397	0.7892	0.8336	0.8482	0.8286

may be used for the experimental adsorption data as per Method II.

The R-square values for the Freundlich adsorption model obtained by the linear regression of the plot of natural logarithm of [E] against [A] were found to be close to 1 and were amongst highest R-square values for any model used to fit the adsorption data of EG II. Similarly, the R-square values for the combined Langmuir-Freundlich adsorption model were found to be close to 1 though the values were not as high as the R-square values for Freundlich adsorption model.

The R-square values for the average values of the adsorption of EG II at different temperatures at a given concentration are given in Table 6. The R-square values are all greater than 0.9 for all models except for the Langmuir model when tested by plotting [E] against [E]/[A]. For the combined Langmuir-Freundlich model the R-square value was found to be less than the R-square values for the other models.

The experimental adsorption data for CBH I, CBH II and EG II can be explained by more than one adsorption model. The range of enzyme concentrations used in the present study is not wide enough to determine whether a single adsorption model can explain the adsorption data. The determination of the enzyme concentration range used in this study was influenced by three factors:

1. The range of concentration of enzymes used in the adsorption experiments conducted by researchers on microcrystalline cellulose in the past studies[74] [81].
2. Limited quantities of pure enzymes available for the present work.
3. Practical range of concentration used in enzymatic processing of cotton fabrics with cellulases.

Thus, a typical enzyme concentration range used for enzymatic treatment of cotton may not be a very wide range to determine whether the adsorption data can be explained by a particular adsorption model. The enzyme concentration range in this research is similar to the range of enzyme concentration used in adsorption experiments conducted on microcrystalline cellulose by Kim et al [82] [83] to determine whether the Langmuir adsorption isotherm is a good fit for their adsorption data. Saturation of microcrystalline cellulose with the different enzymes used was observed in the study conducted by Kim et.al, [82] but in the present study, cotton was not saturated by the enzyme when treated by different purified enzymes in the given range of concentration of purified enzymes.

A possible explanation for the above mentioned observation may be more than those on cotton may be more as compared to the adsorption sites on microcrystalline cellulose. Cotton, which is highly crystalline, may have a large number of sites for

adsorption of enzymes in the form or surface irregularities of crystal surfaces. With the hydrolysis of cotton, the irregularities on the surface of the crystal increase due to enzyme action and hence the adsorption sites for enzymes also increase. As a result of the increase in adsorption sites, the saturation of cotton fibers with the enzymes does not take place. Therefore, more than one models can explain the adsorption data of CBH I, CBH II and EG II.

Based on the structure of cellulases, the catalytic domain is responsible for the hydrolytic activity whereas the cellulose binding domain is responsible for the adsorption of cellulases on cellulose. Therefore, it may be possible that after inhibition of the enzyme by the cellulose hydrolysis product such as glucose or cellobiose the activity of purified enzymes would decrease but the enzyme may still get adsorbed resulting in non-hydrolytic adsorption of the enzyme.

CHAPTER 5

CONCLUSIONS AND RECOMMENDATIONS

Conclusions

From the results of the experiments conducted in the present investigation, it is evident that experimental adsorption data for the adsorption of purified CBH I, CBH II and EG II on cotton fibers in this research can be explained by more than one adsorption model. The experimental adsorption data of purified CBH I, CBH II and EG II on cotton fibers obtained at different temperatures could be explained by the linear, Langmuir, Freundlich and combined Langmuir-Freundlich adsorption models.

The general concentration range of CBH I, CBH II and EG II used in the present investigation was in the range of 0.3 to 2.5 mg/ml which is the practical range of concentration of cellulases used in treatment of cotton with cellulases. The present range of concentration of purified enzymes was sufficient to saturate microcrystalline cellulose as observed in the studies reported [82] [74] but a similar range of concentration was not sufficient to saturate cotton fibers with enzymes. At this concentration range of enzymes, adsorption of

purified enzymes was explained by more than one adsorption model, in the absence of saturation of the substrate with enzyme. Hence a wider range of enzyme concentration should to be used to determine the adsorption model that may best explain the adsorption of CBH I, II and EG II.

Effect of temperature on adsorption of enzymes

From figure 6, which is a plot of the adsorption of CBH I on cotton fibers at different adsorption temperatures at the different concentrations of CBH I studied, it can be said that the adsorption of CBH I on cotton fibers is affected the most at 55 °C. At lower temperatures, effect of temperature of adsorption is minimal in the range of concentration CBH I studied.

Similarly, when the adsorption of CBH II on cotton fibers was plotted against temperature at different concentrations as shown in figure 12, it was found that adsorption of CBH II is affected the most at 55 °C in the given range of concentration CBH II.

Adsorption of EG II on cotton fibers at low concentrations of EG II in the treatment bath was not affected by temperature of adsorption. At higher concentrations of EG II in the treatment bath studied, the amount of EG II adsorbed on cotton fibers increased slightly with increase in temperature of treatment.

Recommendations

Further research is recommended in the following areas:

1. Since the range of enzyme concentration used in the present study is not wide enough for a single adsorption model to explain the experimental adsorption data, higher concentrations of enzyme solutions are recommended in future studies. Though the range of concentration of purified enzymes used in the present study was enough to saturate microcrystalline cellulose, it was not enough to saturate cotton with the enzyme.
2. It is known that agitation of the treatment bath has a significant effect on the extent of cellulose hydrolysis. The effect of agitation on the adsorption of purified cellulases on cotton fibers may be investigated. Such a study may provide some valuable insight on the proven effect of agitation of treatment bath on cellulose hydrolysis.
3. When a mixture of two different enzymes was used in adsorption studied conducted on microcrystalline cellulose, the adsorption of one enzyme is influenced by the presence of the other enzyme in the mixture. Hence adsorption patterns of different mixtures of purified enzymes on cotton fibers such as CBH I and EG II, CBH II and EG II and CBH I and CBH II may be studied.

4. Pretreatment of the cotton fibers such as alkali treatment may influence the adsorption of purified cellulases on cotton fibers. Hence it may be of interest to study the adsorption pattern of purified cellulases on alkali treated cotton fibers.
5. The ionic strength of the enzymatic solution has been said to influence the adsorption pattern of purified cellulases on microcrystalline cellulose [80]. The effect of additives in enzymatic treatment of cotton such as wetting agents and salts present in the buffer system may also affect the adsorption pattern of purified cellulases on cotton.

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Appendix A

Statistical Analysis for Experimental Adsorption Data of
CBH I to fit Adsorption Models

SAS CODE

```
%let path=D:\Dole Files\;
libname bus "&path.";
/* THIS SAS FILE IS FOR REGRESSION FOR DATA ON CBH1a FILE */

/* This step makes new variables for calculating different models */

data temp;
set bus.CBH1a;
m=2;
if E>0.001 then do;
EbyA=E/A;
Einv=1/E;
Ainv=1/A;
LNE=log(E);
LNA=log(A);
CMBLNGE=1/(E**(1/m));
drop m;
end;
run;

/*****
/* regression models ..... FOR TEMPERATURE=25 */
*****/

/* Selecting Cases */

data temp25;
set temp;
if TEMP=25 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT AbyE*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
MODEL Ainv=Einv;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
```

```

PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

/*****
/* regression models ..... FOR TEMPERATURE=35 */
*****/

/* Selecting Cases */

data temp35;
set temp;
if TEMP=35 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL Ainv=Einvs;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;

```

```

PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

/*****
/* regression models ..... FOR TEMPERATURE=45 */
*****/

/* Selecting Cases */

data temp45;
set temp;
if TEMP=45 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL Ainv=Einv;

```

```

OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

/*****
/* regression models ..... FOR TEMPERATURE=55 */
*****/

/* Selecting Cases */

data temp55;
set temp;
if TEMP=55 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
MODEL Ainv=Einv;

```



```

OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

```

SAS OUTPUT

A versus E REGRESSION on TEMPERATURE=25

1

18:08 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	8.59412	8.59412	97.82	<.0001
Error	6	0.52714	0.08786		
Corrected Total	7	9.12126			

Root MSE	0.29641	R-Square	0.9422
Dependent Mean	1.33963	Adj R-Sq	0.9326
Coeff Var	22.12601		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.27850	0.19429	-1.43	0.2017
E	1	1.42613	0.14419	9.89	<.0001

A versus E REGRESSION on TEMPERATURE=25

2

18:08 Saturday, October

25, 2003

Obs	Temp	E	A	EbyA	EinV
1	25	0.001	0.001	.	.
2	25	0.493	0.39	1.26410	2.02840
3	25	0.737	0.61	1.20820	1.35685
4	25	0.976	0.9	1.08444	1.02459
5	25	1.159	0.99	1.17071	0.86281
6	25	1.412	1.94	0.72784	0.70822
7	25	1.827	2.746	0.66533	0.54735
8	25	2.472	3.14	0.78726	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.27707	0.27807
2	2.56410	-0.70725	-0.94161	1.42422	0.42458	-0.03458
3	1.63934	-0.30517	-0.49430	1.16484	0.77256	-0.16256
4	1.11111	-0.02429	-0.10536	1.01222	1.11340	-0.21340
5	1.01010	0.14756	-0.01005	0.92888	1.37439	-0.38439
6	0.51546	0.34501	0.66269	0.84156	1.73520	0.20480
7	0.36417	0.60268	1.01015	0.73983	2.32704	0.41896
8	0.31847	0.90503	1.14422	0.63603	3.24690	-0.10690

EbyA versus E REGRESSION on TEMPERATURE=25

3

18:08 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.25489	0.25489	10.23	0.0240
Error	5	0.12460	0.02492		
Corrected Total	6	0.37949			

Root MSE	0.15786	R-Square	0.6717
Dependent Mean	0.98684	Adj R-Sq	0.6060
Coeff Var	15.99637		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.38109	0.13695	10.08	0.0002
E	1	-0.30407	0.09507	-3.20	0.0240

EbyA versus E REGRESSION on TEMPERATURE=25

4

18:08 Saturday, October

25, 2003

Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.493	0.39	1.26410	2.02840
3	25	0.737	0.61	1.20820	1.35685
4	25	0.976	0.9	1.08444	1.02459
5	25	1.159	0.99	1.17071	0.86281
6	25	1.412	1.94	0.72784	0.70822
7	25	1.827	2.746	0.66533	0.54735
8	25	2.472	3.14	0.78726	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	1.38078	.
2	2.56410	-0.70725	-0.94161	1.42422	1.23118	0.03292
3	1.63934	-0.30517	-0.49430	1.16484	1.15699	0.05121
4	1.11111	-0.02429	-0.10536	1.01222	1.08432	0.00013
5	1.01010	0.14756	-0.01005	0.92888	1.02867	0.14204
6	0.51546	0.34501	0.66269	0.84156	0.95174	-0.22391
7	0.36417	0.60268	1.01015	0.73983	0.82555	-0.16022
8	0.31847	0.90503	1.14422	0.63603	0.62943	0.15783

LANGMUIR REGRESSION on TEMPERATURE=25

5

18:08 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	3.88024	3.88024	373.95	<.0001
Error	5	0.05188	0.01038		
Corrected Total	6	3.93213			

Root MSE	0.10187	R-Square	0.9868
Dependent Mean	1.07468	Adj R-Sq	0.9842

Coeff Var 9.47864

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.36030	0.08360	-4.31	0.0076
Einv	1	1.44890	0.07493	19.34	<.0001

LANGMUIR REGRESSION on TEMPERATURE=25

6

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25, 2003

Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.493	0.39	1.26410	2.02840
3	25	0.737	0.61	1.20820	1.35685
4	25	0.976	0.9	1.08444	1.02459
5	25	1.159	0.99	1.17071	0.86281
6	25	1.412	1.94	0.72784	0.70822
7	25	1.827	2.746	0.66533	0.54735
8	25	2.472	3.14	0.78726	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.56410	-0.70725	-0.94161	1.42422	2.57865	-0.01454
3	1.63934	-0.30517	-0.49430	1.16484	1.60564	0.03370
4	1.11111	-0.02429	-0.10536	1.01222	1.12423	-0.01312
5	1.01010	0.14756	-0.01005	0.92888	0.88983	0.12027
6	0.51546	0.34501	0.66269	0.84156	0.66583	-0.15037
7	0.36417	0.60268	1.01015	0.73983	0.43275	-0.06858
8	0.31847	0.90503	1.14422	0.63603	0.22583	0.09265

FREUNDLICH REGRESSION on TEMPERATURE=25

7

18:08 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	3.54933	3.54933	133.69	<.0001
Error	5	0.13275	0.02655		
Corrected Total	6	3.68208			

Root MSE 0.16294 R-Square 0.9639
 Dependent Mean 0.18082 Adj R-Sq 0.9567
 Coeff Var 90.11203

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.01332	0.06383	-0.21	0.8430
LNE	1	1.41036	0.12198	11.56	<.0001

FREUNDLICH REGRESSION on TEMPERATURE=25

8

25, 2003

18:08 Saturday, October

Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.493	0.39	1.26410	2.02840
3	25	0.737	0.61	1.20820	1.35685
4	25	0.976	0.9	1.08444	1.02459
5	25	1.159	0.99	1.17071	0.86281
6	25	1.412	1.94	0.72784	0.70822
7	25	1.827	2.746	0.66533	0.54735
8	25	2.472	3.14	0.78726	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.56410	-0.70725	-0.94161	1.42422	-1.01079	0.06918
3	1.63934	-0.30517	-0.49430	1.16484	-0.44371	-0.05058
4	1.11111	-0.02429	-0.10536	1.01222	-0.04758	-0.05778
5	1.01010	0.14756	-0.01005	0.92888	0.19479	-0.20484
6	0.51546	0.34501	0.66269	0.84156	0.47327	0.18942
7	0.36417	0.60268	1.01015	0.73983	0.83667	0.17347
8	0.31847	0.90503	1.14422	0.63603	1.26310	-0.11887

COMBINED LANGMUIR REGRESSION on TEMPERATURE=25

9

25, 2003

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The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	3.81548	3.81548	163.55	<.0001
Error	5	0.11664	0.02333		
Corrected Total	6	3.93213			

Root MSE	0.15274	R-Square	0.9703
Dependent Mean	1.07468	Adj R-Sq	0.9644
Coeff Var	14.21244		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-1.80169	0.23221	-7.76	0.0006
CMBLNGE	1	2.98397	0.23333	12.79	<.0001

COMBINED LANGMUIR REGRESSION on TEMPERATURE=25

10

25, 2003

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Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.493	0.39	1.26410	2.02840
3	25	0.737	0.61	1.20820	1.35685
4	25	0.976	0.9	1.08444	1.02459
5	25	1.159	0.99	1.17071	0.86281
6	25	1.412	1.94	0.72784	0.70822

7	25	1.827	2.746	0.66533	0.54735	
8	25	2.472	3.14	0.78726	0.40453	
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.56410	-0.70725	-0.94161	1.42422	2.44814	0.11596
3	1.63934	-0.30517	-0.49430	1.16484	1.67417	-0.03482
4	1.11111	-0.02429	-0.10536	1.01222	1.21875	-0.10764
5	1.01010	0.14756	-0.01005	0.92888	0.97006	0.04004
6	0.51546	0.34501	0.66269	0.84156	0.70949	-0.19403
7	0.36417	0.60268	1.01015	0.73983	0.40594	-0.04178
8	0.31847	0.90503	1.14422	0.63603	0.09620	0.22227

A versus E REGRESSION on TEMPERATURE=35

11

18:08 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	8.03863	8.03863	45.24	0.0005
Error	6	1.06609	0.17768		
Corrected Total	7	9.10472			

Root MSE	0.42152	R-Square	0.8829
Dependent Mean	1.43375	Adj R-Sq	0.8634
Coeff Var	29.40006		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.13121	0.27630	-0.47	0.6517
E	1	1.37927	0.20506	6.73	0.0005

A versus E REGRESSION on TEMPERATURE=35

12

18:08 Saturday, October

25, 2003

Obs	Temp	E	A	EbyA	Einv	
1	35	0.001	0.001	.	.	
2	35	0.493	0.45	1.09556	2.02840	
3	35	0.737	0.44	1.67500	1.35685	
4	35	0.976	0.83	1.17590	1.02459	
5	35	1.159	1.81	0.64033	0.86281	
6	35	1.412	2.516	0.56121	0.70822	
7	35	1.827	2.434	0.75062	0.54735	
8	35	2.472	2.989	0.82703	0.40453	
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.12983	0.13083
2	2.22222	-0.70725	-0.79851	1.42422	0.54877	-0.09877
3	2.27273	-0.30517	-0.82098	1.16484	0.88532	-0.44532
4	1.20482	-0.02429	-0.18633	1.01222	1.21496	-0.38496
5	0.55249	0.14756	0.59333	0.92888	1.46737	0.34263
6	0.39746	0.34501	0.92267	0.84156	1.81633	0.69967
7	0.41085	0.60268	0.88954	0.73983	2.38872	0.04528

8 0.33456 0.90503 1.09494 0.63603 3.27835 -0.28935

EbyA versus E REGRESSION on TEMPERATURE=35

13

18:08 Saturday, October

25, 2003

The REG Procedure
Model: MODEL1
Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.26063	0.26063	2.04	0.2124
Error	5	0.63833	0.12767		
Corrected Total	6	0.89895			

Root MSE	0.35730	R-Square	0.2899
Dependent Mean	0.96081	Adj R-Sq	0.1479
Coeff Var	37.18788		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.35946	0.30998	4.39	0.0071
E	1	-0.30747	0.21519	-1.43	0.2124

EbyA versus E REGRESSION on TEMPERATURE=35

14

18:08 Saturday, October

25, 2003

Obs	Temp	E	A	EbyA	Einvs
1	35	0.001	0.001	.	.
2	35	0.493	0.45	1.09556	2.02840
3	35	0.737	0.44	1.67500	1.35685
4	35	0.976	0.83	1.17590	1.02459
5	35	1.159	1.81	0.64033	0.86281
6	35	1.412	2.516	0.56121	0.70822
7	35	1.827	2.434	0.75062	0.54735
8	35	2.472	2.989	0.82703	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	1.35916	.
2	2.22222	-0.70725	-0.79851	1.42422	1.20788	-0.11232
3	2.27273	-0.30517	-0.82098	1.16484	1.13286	0.54214
4	1.20482	-0.02429	-0.18633	1.01222	1.05937	0.11653
5	0.55249	0.14756	0.59333	0.92888	1.00311	-0.36277
6	0.39746	0.34501	0.92267	0.84156	0.92532	-0.36411
7	0.41085	0.60268	0.88954	0.73983	0.79772	-0.04710
8	0.33456	0.90503	1.09494	0.63603	0.59940	0.22763

LANGMUIR REGRESSION on TEMPERATURE=35

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The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	3.71403	3.71403	24.04	0.0045
Error	5	0.77252	0.15450		
Corrected Total	6	4.48655			

Root MSE 0.39307 R-Square 0.8278
 Dependent Mean 1.05645 Adj R-Sq 0.7934
 Coeff Var 37.20677

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.34746	0.32259	-1.08	0.3306
Einvs	1	1.41753	0.28912	4.90	0.0045

LANGMUIR REGRESSION on TEMPERATURE=35

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Obs	Temp	E	A	EbyA	Einvs
1	35	0.001	0.001	.	.
2	35	0.493	0.45	1.09556	2.02840
3	35	0.737	0.44	1.67500	1.35685
4	35	0.976	0.83	1.17590	1.02459
5	35	1.159	1.81	0.64033	0.86281
6	35	1.412	2.516	0.56121	0.70822
7	35	1.827	2.434	0.75062	0.54735
8	35	2.472	2.989	0.82703	0.40453

Obs	Ainvs	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.22222	-0.70725	-0.79851	1.42422	2.52785	-0.30563
3	2.27273	-0.30517	-0.82098	1.16484	1.57591	0.69681
4	1.20482	-0.02429	-0.18633	1.01222	1.10492	0.09990
5	0.55249	0.14756	0.59333	0.92888	0.87560	-0.32311
6	0.39746	0.34501	0.92267	0.84156	0.65645	-0.25900
7	0.41085	0.60268	0.88954	0.73983	0.42841	-0.01757
8	0.33456	0.90503	1.09494	0.63603	0.22597	0.10859

FREUNDLICH REGRESSION on TEMPERATURE=35

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The REG Procedure
 Model: MODEL1
 Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	3.55737	3.55737	31.08	0.0026
Error	5	0.57223	0.11445		
Corrected Total	6	4.12960			

Root MSE 0.33830 R-Square 0.8614
 Dependent Mean 0.24209 Adj R-Sq 0.8337
 Coeff Var 139.73843

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.04774	0.13253	0.36	0.7334
LNE	1	1.41196	0.25325	5.58	0.0026

FREUNDLICH REGRESSION on TEMPERATURE=35

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Obs	Temp	E	A	EbyA	Einv
1	35	0.001	0.001	.	.
2	35	0.493	0.45	1.09556	2.02840
3	35	0.737	0.44	1.67500	1.35685
4	35	0.976	0.83	1.17590	1.02459
5	35	1.159	1.81	0.64033	0.86281
6	35	1.412	2.516	0.56121	0.70822
7	35	1.827	2.434	0.75062	0.54735
8	35	2.472	2.989	0.82703	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.22222	-0.70725	-0.79851	1.42422	-0.95087	0.15236
3	2.27273	-0.30517	-0.82098	1.16484	-0.38315	-0.43783
4	1.20482	-0.02429	-0.18633	1.01222	0.01343	-0.19976
5	0.55249	0.14756	0.59333	0.92888	0.25608	0.33725
6	0.39746	0.34501	0.92267	0.84156	0.53487	0.38780
7	0.41085	0.60268	0.88954	0.73983	0.89869	-0.00915
8	0.33456	0.90503	1.09494	0.63603	1.32560	-0.23066

COMBINED LANGMUIR REGRESSION on TEMPERATURE=35

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The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	3.75107	3.75107	25.50	0.0039
Error	5	0.73548	0.14710		
Corrected Total	6	4.48655			

Root MSE	0.38353	R-Square	0.8361
Dependent Mean	1.05645	Adj R-Sq	0.8033
Coeff Var	36.30394		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-1.79554	0.58308	-3.08	0.0275
CMBLNGE	1	2.95868	0.58590	5.05	0.0039

COMBINED LANGMUIR REGRESSION on TEMPERATURE=35

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Obs	Temp	E	A	EbyA	Einv
1	35	0.001	0.001	.	.
2	35	0.493	0.45	1.09556	2.02840
3	35	0.737	0.44	1.67500	1.35685
4	35	0.976	0.83	1.17590	1.02459
5	35	1.159	1.81	0.64033	0.86281
6	35	1.412	2.516	0.56121	0.70822
7	35	1.827	2.434	0.75062	0.54735
8	35	2.472	2.989	0.82703	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.22222	-0.70725	-0.79851	1.42422	2.41827	-0.19604
3	2.27273	-0.30517	-0.82098	1.16484	1.65085	0.62188
4	1.20482	-0.02429	-0.18633	1.01222	1.19930	0.00552
5	0.55249	0.14756	0.59333	0.92888	0.95271	-0.40023
6	0.39746	0.34501	0.92267	0.84156	0.69435	-0.29690
7	0.41085	0.60268	0.88954	0.73983	0.39338	0.01747
8	0.33456	0.90503	1.09494	0.63603	0.08626	0.24830

A versus E REGRESSION on TEMPERATURE=45

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The REG Procedure
 Model: MODEL1
 Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	6.31416	6.31416	164.69	<.0001
Error	6	0.23004	0.03834		
Corrected Total	7	6.54420			

Root MSE	0.19581	R-Square	0.9648
Dependent Mean	1.24150	Adj R-Sq	0.9590
Coeff Var	15.77174		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.14548	0.12835	-1.13	0.3003
E	1	1.22241	0.09525	12.83	<.0001

A versus E REGRESSION on TEMPERATURE=45

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Obs	Temp	E	A	EbyA	Einv
1	45	0.001	0.001	.	.
2	45	0.493	0.47	1.04894	2.02840
3	45	0.737	0.57	1.29298	1.35685
4	45	0.976	1.24	0.78710	1.02459
5	45	1.159	1.11	1.04414	0.86281
6	45	1.412	1.6	0.88250	0.70822
7	45	1.827	1.841	0.99240	0.54735

8		45	2.472	3.1	0.79742	0.40453
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.14425	0.14525
2	2.12766	-0.70725	-0.75502	1.42422	0.45717	0.01283
3	1.75439	-0.30517	-0.56212	1.16484	0.75544	-0.18544
4	0.80645	-0.02429	0.21511	1.01222	1.04760	0.19240
5	0.90090	0.14756	0.10436	0.92888	1.27130	-0.16130
6	0.62500	0.34501	0.47000	0.84156	1.58057	0.01943
7	0.54318	0.60268	0.61031	0.73983	2.08787	-0.24687
8	0.32258	0.90503	1.13140	0.63603	2.87632	0.22368

EbyA versus E REGRESSION on TEMPERATURE=45

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The REG Procedure
 Model: MODEL1
 Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.05711	0.05711	2.20	0.1982
Error	5	0.12989	0.02598		
Corrected Total	6	0.18700			

Root MSE	0.16118	R-Square	0.3054
Dependent Mean	0.97792	Adj R-Sq	0.1665
Coeff Var	16.48137		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.16455	0.13983	8.33	0.0004
E	1	-0.14393	0.09707	-1.48	0.1982

EbyA versus E REGRESSION on TEMPERATURE=45

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Obs	Temp	E	A	EbyA	Einv	
1	45	0.001	0.001	.	.	
2	45	0.493	0.47	1.04894	2.02840	
3	45	0.737	0.57	1.29298	1.35685	
4	45	0.976	1.24	0.78710	1.02459	
5	45	1.159	1.11	1.04414	0.86281	
6	45	1.412	1.6	0.88250	0.70822	
7	45	1.827	1.841	0.99240	0.54735	
8	45	2.472	3.1	0.79742	0.40453	
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	1.16440	.
2	2.12766	-0.70725	-0.75502	1.42422	1.09359	-0.04465
3	1.75439	-0.30517	-0.56212	1.16484	1.05847	0.23452
4	0.80645	-0.02429	0.21511	1.01222	1.02407	-0.23697
5	0.90090	0.14756	0.10436	0.92888	0.99773	0.04642
6	0.62500	0.34501	0.47000	0.84156	0.96131	-0.07881
7	0.54318	0.60268	0.61031	0.73983	0.90158	0.09082
8	0.32258	0.90503	1.13140	0.63603	0.80874	-0.01132

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LANGMUIR REGRESSION on TEMPERATURE=45

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The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.51971	2.51971	71.76	0.0004
Error	5	0.17557	0.03511		
Corrected Total	6	2.69528			

Root MSE	0.18739	R-Square	0.9349
Dependent Mean	1.01145	Adj R-Sq	0.9218
Coeff Var	18.52666		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.14490	0.15379	-0.94	0.3893
Einv	1	1.16757	0.13783	8.47	0.0004

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LANGMUIR REGRESSION on TEMPERATURE=45

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Obs	Temp	E	A	EbyA	Einv
1	45	0.001	0.001	.	.
2	45	0.493	0.47	1.04894	2.02840
3	45	0.737	0.57	1.29298	1.35685
4	45	0.976	1.24	0.78710	1.02459
5	45	1.159	1.11	1.04414	0.86281
6	45	1.412	1.6	0.88250	0.70822
7	45	1.827	1.841	0.99240	0.54735
8	45	2.472	3.1	0.79742	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.12766	-0.70725	-0.75502	1.42422	2.22340	-0.09574
3	1.75439	-0.30517	-0.56212	1.16484	1.43932	0.31507
4	0.80645	-0.02429	0.21511	1.01222	1.05138	-0.24493
5	0.90090	0.14756	0.10436	0.92888	0.86249	0.03841
6	0.62500	0.34501	0.47000	0.84156	0.68199	-0.05699
7	0.54318	0.60268	0.61031	0.73983	0.49416	0.04902
8	0.32258	0.90503	1.13140	0.63603	0.32742	-0.00484

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FREUNDLICH REGRESSION on TEMPERATURE=45

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The REG Procedure
Model: MODEL1
Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.47736	2.47736	96.22	0.0002
Error	5	0.12873	0.02575		
Corrected Total	6	2.60609			

Root MSE 0.16046 R-Square 0.9506
 Dependent Mean 0.17343 Adj R-Sq 0.9407
 Coeff Var 92.51701

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.01124	0.06286	0.18	0.8651
LNE	1	1.17829	0.12012	9.81	0.0002

FREUNDLICH REGRESSION on TEMPERATURE=45

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Obs	Temp	E	A	EbyA	EinV
1	45	0.001	0.001	.	.
2	45	0.493	0.47	1.04894	2.02840
3	45	0.737	0.57	1.29298	1.35685
4	45	0.976	1.24	0.78710	1.02459
5	45	1.159	1.11	1.04414	0.86281
6	45	1.412	1.6	0.88250	0.70822
7	45	1.827	1.841	0.99240	0.54735
8	45	2.472	3.1	0.79742	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.12766	-0.70725	-0.75502	1.42422	-0.82210	0.06708
3	1.75439	-0.30517	-0.56212	1.16484	-0.34833	-0.21379
4	0.80645	-0.02429	0.21511	1.01222	-0.01738	0.23249
5	0.90090	0.14756	0.10436	0.92888	0.18511	-0.08075
6	0.62500	0.34501	0.47000	0.84156	0.41776	0.05224
7	0.54318	0.60268	0.61031	0.73983	0.72137	-0.11106
8	0.32258	0.90503	1.13140	0.63603	1.07763	0.05378

COMBINED LANGMUIR REGRESSION on TEMPERATURE=45

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The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.50052	2.50052	64.20	0.0005
Error	5	0.19476	0.03895		
Corrected Total	6	2.69528			

Root MSE 0.19736 R-Square 0.9277
 Dependent Mean 1.01145 Adj R-Sq 0.9133
 Coeff Var 19.51271

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-1.31710	0.30005	-4.39	0.0071
CMBLNGE	1	2.41566	0.30150	8.01	0.0005

COMBINED LANGMUIR REGRESSION on TEMPERATURE=45

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Obs	Temp	E	A	EbyA	EinV
1	45	0.001	0.001	.	.
2	45	0.493	0.47	1.04894	2.02840
3	45	0.737	0.57	1.29298	1.35685
4	45	0.976	1.24	0.78710	1.02459
5	45	1.159	1.11	1.04414	0.86281
6	45	1.412	1.6	0.88250	0.70822
7	45	1.827	1.841	0.99240	0.54735
8	45	2.472	3.1	0.79742	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.12766	-0.70725	-0.75502	1.42422	2.12333	0.00433
3	1.75439	-0.30517	-0.56212	1.16484	1.49676	0.25762
4	0.80645	-0.02429	0.21511	1.01222	1.12809	-0.32163
5	0.90090	0.14756	0.10436	0.92888	0.92676	-0.02586
6	0.62500	0.34501	0.47000	0.84156	0.71582	-0.09082
7	0.54318	0.60268	0.61031	0.73983	0.47008	0.07311
8	0.32258	0.90503	1.13140	0.63603	0.21933	0.10325

A versus E REGRESSION on TEMPERATURE=55

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The REG Procedure
Model: MODEL1
Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	9.45160	9.45160	713.29	<.0001
Error	6	0.07950	0.01325		
Corrected Total	7	9.53111			

Root MSE	0.11511	R-Square	0.9917
Dependent Mean	1.76550	Adj R-Sq	0.9903
Coeff Var	6.52007		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.06857	0.07545	0.91	0.3985
E	1	1.49559	0.05600	26.71	<.0001

A versus E REGRESSION on TEMPERATURE=55

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Obs	Temp	E	A	EbyA	EinV
1	55	0.001	0.001	.	.
2	55	0.493	0.75	0.65733	2.02840
3	55	0.737	1.19	0.61933	1.35685
4	55	0.976	1.7	0.57412	1.02459
5	55	1.159	1.89	0.61323	0.86281
6	55	1.412	2.173	0.64979	0.70822
7	55	1.827	2.62	0.69733	0.54735
8	55	2.472	3.8	0.65053	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	0.07007	-0.06907
2	1.33333	-0.70725	-0.28768	1.42422	0.80589	-0.05589
3	0.84034	-0.30517	0.17395	1.16484	1.17082	0.01918
4	0.58824	-0.02429	0.53063	1.01222	1.52826	0.17174
5	0.52910	0.14756	0.63658	0.92888	1.80195	0.08805
6	0.46019	0.34501	0.77611	0.84156	2.18034	-0.00734
7	0.38168	0.60268	0.96317	0.73983	2.80101	-0.18101
8	0.26316	0.90503	1.33500	0.63603	3.76566	0.03434

EbyA versus E REGRESSION on TEMPERATURE=55

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18:08 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.00160	0.00160	1.05	0.3533
Error	5	0.00763	0.00153		
Corrected Total	6	0.00923			

Root MSE	0.03907	R-Square	0.1730
Dependent Mean	0.63738	Adj R-Sq	0.0076
Coeff Var	6.13011		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.60617	0.03390	17.88	<.0001
E	1	0.02407	0.02353	1.02	0.3533

EbyA versus E REGRESSION on TEMPERATURE=55

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25, 2003

Obs	Temp	E	A	EbyA	EinV
1	55	0.001	0.001	.	.
2	55	0.493	0.75	0.65733	2.02840
3	55	0.737	1.19	0.61933	1.35685
4	55	0.976	1.7	0.57412	1.02459
5	55	1.159	1.89	0.61323	0.86281
6	55	1.412	2.173	0.64979	0.70822
7	55	1.827	2.62	0.69733	0.54735
8	55	2.472	3.8	0.65053	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	0.60620	.
2	1.33333	-0.70725	-0.28768	1.42422	0.61804	0.039295
3	0.84034	-0.30517	0.17395	1.16484	0.62391	-0.004583
4	0.58824	-0.02429	0.53063	1.01222	0.62966	-0.055546
5	0.52910	0.14756	0.63658	0.92888	0.63407	-0.020840
6	0.46019	0.34501	0.77611	0.84156	0.64016	0.009636
7	0.38168	0.60268	0.96317	0.73983	0.65015	0.047182
8	0.26316	0.90503	1.33500	0.63603	0.66567	-0.015144

LANGMUIR REGRESSION on TEMPERATURE=55

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The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.76800	0.76800	486.96	<.0001
Error	5	0.00789	0.00158		
Corrected Total	6	0.77589			

Root MSE	0.03971	R-Square	0.9898
Dependent Mean	0.62801	Adj R-Sq	0.9878
Coeff Var	6.32367		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.01040	0.03259	-0.32	0.7625
Einv	1	0.64460	0.02921	22.07	<.0001

LANGMUIR REGRESSION on TEMPERATURE=55

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Obs	Temp	E	A	EbyA	Einv
1	55	0.001	0.001	.	.
2	55	0.493	0.75	0.65733	2.02840
3	55	0.737	1.19	0.61933	1.35685
4	55	0.976	1.7	0.57412	1.02459
5	55	1.159	1.89	0.61323	0.86281
6	55	1.412	2.173	0.64979	0.70822
7	55	1.827	2.62	0.69733	0.54735
8	55	2.472	3.8	0.65053	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	1.33333	-0.70725	-0.28768	1.42422	1.29710	0.036231
3	0.84034	-0.30517	0.17395	1.16484	0.86423	-0.023889
4	0.58824	-0.02429	0.53063	1.01222	0.65005	-0.061814
5	0.52910	0.14756	0.63658	0.92888	0.54577	-0.016667
6	0.46019	0.34501	0.77611	0.84156	0.44611	0.014079
7	0.38168	0.60268	0.96317	0.73983	0.34242	0.039262
8	0.26316	0.90503	1.33500	0.63603	0.25036	0.012798

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25, 2003

FREUNDLICH REGRESSION on TEMPERATURE=55

18:08 Saturday, October

The REG Procedure
Model: MODEL1
Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	1.65734	1.65734	400.33	<.0001
Error	5	0.02070	0.00414		
Corrected Total	6	1.67804			

Root MSE	0.06434	R-Square	0.9877
Dependent Mean	0.58968	Adj R-Sq	0.9852
Coeff Var	10.91142		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.45702	0.02521	18.13	<.0001
LNE	1	0.96375	0.04817	20.01	<.0001

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25, 2003

FREUNDLICH REGRESSION on TEMPERATURE=55

18:08 Saturday, October

Obs	Temp	E	A	EbyA	Einv
1	55	0.001	0.001	.	.
2	55	0.493	0.75	0.65733	2.02840
3	55	0.737	1.19	0.61933	1.35685
4	55	0.976	1.7	0.57412	1.02459
5	55	1.159	1.89	0.61323	0.86281
6	55	1.412	2.173	0.64979	0.70822
7	55	1.827	2.62	0.69733	0.54735
8	55	2.472	3.8	0.65053	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	1.33333	-0.70725	-0.28768	1.42422	-0.22459	-0.063096
3	0.84034	-0.30517	0.17395	1.16484	0.16292	0.011038
4	0.58824	-0.02429	0.53063	1.01222	0.43361	0.097021
5	0.52910	0.14756	0.63658	0.92888	0.59923	0.037350
6	0.46019	0.34501	0.77611	0.84156	0.78952	-0.013409
7	0.38168	0.60268	0.96317	0.73983	1.03784	-0.074670
8	0.26316	0.90503	1.33500	0.63603	1.32924	0.005766

39
25, 2003

COMBINED LANGMUIR REGRESSION on TEMPERATURE=55

18:08 Saturday, October

The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Sum of	Mean
--------	------

Source	DF	Squares	Square	F Value	Pr > F
Model	1	0.74295	0.74295	112.77	0.0001
Error	5	0.03294	0.00659		
Corrected Total	6	0.77589			

Root MSE	0.08117	R-Square	0.9575
Dependent Mean	0.62801	Adj R-Sq	0.9491
Coeff Var	12.92472		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.64125	0.12340	-5.20	0.0035
CMBLNGE	1	1.31674	0.12400	10.62	0.0001

COMBINED LANGMUIR REGRESSION on TEMPERATURE=55

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18:08 Saturday, October

25, 2003

Obs	Temp	E	A	EbyA	Einv
1	55	0.001	0.001	.	.
2	55	0.493	0.75	0.65733	2.02840
3	55	0.737	1.19	0.61933	1.35685
4	55	0.976	1.7	0.57412	1.02459
5	55	1.159	1.89	0.61323	0.86281
6	55	1.412	2.173	0.64979	0.70822
7	55	1.827	2.62	0.69733	0.54735
8	55	2.472	3.8	0.65053	0.40453

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	1.33333	-0.70725	-0.28768	1.42422	1.23407	0.09926
3	0.84034	-0.30517	0.17395	1.16484	0.89254	-0.05220
4	0.58824	-0.02429	0.53063	1.01222	0.69158	-0.10335
5	0.52910	0.14756	0.63658	0.92888	0.58184	-0.05274
6	0.46019	0.34501	0.77611	0.84156	0.46686	-0.00667
7	0.38168	0.60268	0.96317	0.73983	0.33291	0.04877
8	0.26316	0.90503	1.33500	0.63603	0.19623	0.06692

Appendix B

Statistical Analysis for Experimental Adsorption Data of
CBH II to fit Adsorption Models

SAS CODE

```
%let path=D:\Dole Files\;
libname bus "&path.";
/* THIS SAS FILE IS FOR REGRESSION FOR DATA ON CBHIIa FILE */

/* This step makes new variables for calculating different models */

data temp;
set bus.CBHIIa;
if E>0.001 then do;
m=2;
EbyA=E/A;
Einv=1/E;
Ainv=1/A;
LNE=log(E);
LNA=log(A);
CMBLNGE=1/(E**(1/m));
drop m;
end;
run;

/*****
/* regression models ..... FOR TEMPERATURE=25 */
*****/

/* Selecting Cases */

data temp25;
set temp;
if TEMP=25 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
  MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
  MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
```

```

MODEL Ainv=Einvs;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einvs='*' PREDICTED.*Einvs='o' RESIDUAL.*Einvs='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

/*****
/* regression models ..... FOR TEMPERATURE=35 */
*****/

/* Selecting Cases */

data temp35;
set temp;
if TEMP=35 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;

```

```

MODEL Ainv=Einv;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

/*****
/* regression models ..... FOR TEMPERATURE=45 */
*****/

/* Selecting Cases */

data temp45;
set temp;
if TEMP=45 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on TEMPERATURE=45';

```

```

PROC REG data=temp45;
  MODEL Ainv=Einv;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

/*****
/* regression models ..... FOR TEMPERATURE=55 */
*****/

/* Selecting Cases */

data temp55;
set temp;
if TEMP=55 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
  MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
  MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on TEMPERATURE=55';

```

```

PROC REG data=temp55;
  MODEL Ainv=Einv;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

```


SAS OUTPUT

A versus E REGRESSION on TEMPERATURE=25

1

18:32 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	13.75884	13.75884	70.23	0.0002
Error	6	1.17549	0.19592		
Corrected Total	7	14.93433			

Root MSE	0.44262	R-Square	0.9213
Dependent Mean	1.80025	Adj R-Sq	0.9082
Coeff Var	24.58677		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.60548	0.32695	-1.85	0.1135
E	1	2.40302	0.28675	8.38	0.0002

A versus E REGRESSION on TEMPERATURE=25

2

18:32 Saturday, October

25, 2003

Obs	Temp	E	A	EbyA	EinV
1	25	0.001	0.001	.	.
2	25	0.477	0.3	1.59000	2.09644
3	25	0.715	0.52	1.37500	1.39860
4	25	0.929	1.44	0.64514	1.07643
5	25	1.315	2.19	0.60046	0.76046
6	25	1.335	2.763	0.48317	0.74906
7	25	1.463	3.308	0.44226	0.68353
8	25	1.774	3.88	0.45722	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.60307	0.60407
2	3.33333	-0.74024	-1.20397	1.44791	0.54077	-0.24077
3	1.92308	-0.33547	-0.65393	1.18262	1.11268	-0.59268
4	0.69444	-0.07365	0.36464	1.03751	1.62693	-0.18693
5	0.45662	0.27384	0.78390	0.87204	2.55450	-0.36450
6	0.36193	0.28893	1.01632	0.86548	2.60256	0.16044
7	0.30230	0.38049	1.19634	0.82676	2.91015	0.39785
8	0.25773	0.57324	1.35584	0.75080	3.65749	0.22251

EbyA versus E REGRESSION on TEMPERATURE=25

3

18:32 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	1.09498	1.09498	20.32	0.0064
Error	5	0.26940	0.05388		
Corrected Total	6	1.36438			

Root MSE	0.23212	R-Square	0.8025
Dependent Mean	0.79903	Adj R-Sq	0.7631
Coeff Var	29.05000		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.87426	0.25413	7.38	0.0007
E	1	-0.93988	0.20849	-4.51	0.0064

EbyA versus E REGRESSION on TEMPERATURE=25

4

18:32 Saturday, October

25, 2003

Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.477	0.3	1.59000	2.09644
3	25	0.715	0.52	1.37500	1.39860
4	25	0.929	1.44	0.64514	1.07643
5	25	1.315	2.19	0.60046	0.76046
6	25	1.335	2.763	0.48317	0.74906
7	25	1.463	3.308	0.44226	0.68353
8	25	1.774	3.88	0.45722	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	1.87332	.
2	3.33333	-0.74024	-1.20397	1.44791	1.42594	0.16406
3	1.92308	-0.33547	-0.65393	1.18262	1.20224	0.17276
4	0.69444	-0.07365	0.36464	1.03751	1.00111	-0.35597
5	0.45662	0.27384	0.78390	0.87204	0.63832	-0.03786
6	0.36193	0.28893	1.01632	0.86548	0.61952	-0.13635
7	0.30230	0.38049	1.19634	0.82676	0.49921	-0.05695
8	0.25773	0.57324	1.35584	0.75080	0.20691	0.25031

LANGMUIR REGRESSION on TEMPERATURE=25

5

18:32 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	7.86240	7.86240	155.92	<.0001
Error	5	0.25213	0.05043		
Corrected Total	6	8.11453			

Root MSE	0.22456	R-Square	0.9689
Dependent Mean	1.04706	Adj R-Sq	0.9627

Coeff Var 21.44637

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-1.16414	0.19637	-5.93	0.0019
Einv	1	2.11217	0.16915	12.49	<.0001

LANGMUIR REGRESSION on TEMPERATURE=25

6

18:32 Saturday, October

25, 2003

Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.477	0.3	1.59000	2.09644
3	25	0.715	0.52	1.37500	1.39860
4	25	0.929	1.44	0.64514	1.07643
5	25	1.315	2.19	0.60046	0.76046
6	25	1.335	2.763	0.48317	0.74906
7	25	1.463	3.308	0.44226	0.68353
8	25	1.774	3.88	0.45722	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	3.33333	-0.74024	-1.20397	1.44791	3.26389	0.06945
3	1.92308	-0.33547	-0.65393	1.18262	1.78994	0.13313
4	0.69444	-0.07365	0.36464	1.03751	1.10945	-0.41501
5	0.45662	0.27384	0.78390	0.87204	0.44207	0.01455
6	0.36193	0.28893	1.01632	0.86548	0.41801	-0.05608
7	0.30230	0.38049	1.19634	0.82676	0.27958	0.02271
8	0.25773	0.57324	1.35584	0.75080	0.02648	0.23125

FREUNDLICH REGRESSION on TEMPERATURE=25

7

18:32 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	5.59482	5.59482	170.12	<.0001
Error	5	0.16444	0.03289		
Corrected Total	6	5.75925			

Root MSE 0.18135 R-Square 0.9714
 Dependent Mean 0.40845 Adj R-Sq 0.9657
 Coeff Var 44.39924

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.29873	0.06906	4.33	0.0075
LNE	1	2.09190	0.16038	13.04	<.0001

FREUNDLICH REGRESSION on TEMPERATURE=25

8

25, 2003

18:32 Saturday, October

Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.477	0.3	1.59000	2.09644
3	25	0.715	0.52	1.37500	1.39860
4	25	0.929	1.44	0.64514	1.07643
5	25	1.315	2.19	0.60046	0.76046
6	25	1.335	2.763	0.48317	0.74906
7	25	1.463	3.308	0.44226	0.68353
8	25	1.774	3.88	0.45722	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	3.33333	-0.74024	-1.20397	1.44791	-1.24978	0.04580
3	1.92308	-0.33547	-0.65393	1.18262	-0.40304	-0.25088
4	0.69444	-0.07365	0.36464	1.03751	0.14467	0.21997
5	0.45662	0.27384	0.78390	0.87204	0.87157	-0.08767
6	0.36193	0.28893	1.01632	0.86548	0.90315	0.11317
7	0.30230	0.38049	1.19634	0.82676	1.09468	0.10166
8	0.25773	0.57324	1.35584	0.75080	1.49789	-0.14205

COMBINED LANGMUIR REGRESSION on TEMPERATURE=25

9

25, 2003

18:32 Saturday, October

The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	7.65270	7.65270	82.85	0.0003
Error	5	0.46183	0.09237		
Corrected Total	6	8.11453			

Root MSE	0.30392	R-Square	0.9431
Dependent Mean	1.04706	Adj R-Sq	0.9317
Coeff Var	29.02578		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-3.54018	0.51689	-6.85	0.0010
CMBLNGE	1	4.59833	0.50518	9.10	0.0003

COMBINED LANGMUIR REGRESSION on TEMPERATURE=25

10

25, 2003

18:32 Saturday, October

Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.477	0.3	1.59000	2.09644
3	25	0.715	0.52	1.37500	1.39860
4	25	0.929	1.44	0.64514	1.07643
5	25	1.315	2.19	0.60046	0.76046
6	25	1.335	2.763	0.48317	0.74906

7	25	1.463	3.308	0.44226	0.68353	
8	25	1.774	3.88	0.45722	0.56370	
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	3.33333	-0.74024	-1.20397	1.44791	3.11777	0.21556
3	1.92308	-0.33547	-0.65393	1.18262	1.89792	0.02516
4	0.69444	-0.07365	0.36464	1.03751	1.23063	-0.53619
5	0.45662	0.27384	0.78390	0.87204	0.46975	-0.01313
6	0.36193	0.28893	1.01632	0.86548	0.43960	-0.07768
7	0.30230	0.38049	1.19634	0.82676	0.26152	0.04078
8	0.25773	0.57324	1.35584	0.75080	-0.08776	0.34550

A versus E REGRESSION on TEMPERATURE=35

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The REG Procedure
 Model: MODEL1
 Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	16.70985	16.70985	80.05	0.0001
Error	6	1.25238	0.20873		
Corrected Total	7	17.96223			

Root MSE	0.45687	R-Square	0.9303
Dependent Mean	2.01625	Adj R-Sq	0.9187
Coeff Var	22.65942		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.63494	0.33748	-1.88	0.1089
E	1	2.64822	0.29598	8.95	0.0001

A versus E REGRESSION on TEMPERATURE=35

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Obs	Temp	E	A	EbyA	Einv	
1	35	0.001	0.001	.	.	
2	35	0.477	0.51	0.93529	2.09644	
3	35	0.715	0.6	1.19167	1.39860	
4	35	0.929	1.56	0.59551	1.07643	
5	35	1.315	2.66	0.49436	0.76046	
6	35	1.335	3.03	0.44059	0.74906	
7	35	1.463	3.179	0.46021	0.68353	
8	35	1.774	4.59	0.38649	0.56370	
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.63230	0.63330
2	1.96078	-0.74024	-0.67334	1.44791	0.62825	-0.11825
3	1.66667	-0.33547	-0.51083	1.18262	1.25853	-0.65853
4	0.64103	-0.07365	0.44469	1.03751	1.82525	-0.26525
5	0.37594	0.27384	0.97833	0.87204	2.84746	-0.18746
6	0.33003	0.28893	1.10856	0.86548	2.90042	0.12958
7	0.31456	0.38049	1.15657	0.82676	3.23939	-0.06039

8 0.21786 0.57324 1.52388 0.75080 4.06299 0.52701

EbyA versus E REGRESSION on TEMPERATURE=35

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25, 2003

The REG Procedure
Model: MODEL1
Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.39997	0.39997	13.24	0.0149
Error	5	0.15102	0.03020		
Corrected Total	6	0.55100			

Root MSE 0.17380 R-Square 0.7259
Dependent Mean 0.64345 Adj R-Sq 0.6711
Coeff Var 27.01010

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.29329	0.19028	6.80	0.0010
E	1	-0.56805	0.15610	-3.64	0.0149

EbyA versus E REGRESSION on TEMPERATURE=35

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Obs	Temp	E	A	EbyA	Einvs
1	35	0.001	0.001	.	.
2	35	0.477	0.51	0.93529	2.09644
3	35	0.715	0.6	1.19167	1.39860
4	35	0.929	1.56	0.59551	1.07643
5	35	1.315	2.66	0.49436	0.76046
6	35	1.335	3.03	0.44059	0.74906
7	35	1.463	3.179	0.46021	0.68353
8	35	1.774	4.59	0.38649	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	1.29272	.
2	1.96078	-0.74024	-0.67334	1.44791	1.02233	-0.08704
3	1.66667	-0.33547	-0.51083	1.18262	0.88714	0.30453
4	0.64103	-0.07365	0.44469	1.03751	0.76558	-0.17006
5	0.37594	0.27384	0.97833	0.87204	0.54631	-0.05195
6	0.33003	0.28893	1.10856	0.86548	0.53495	-0.09436
7	0.31456	0.38049	1.15657	0.82676	0.46224	-0.00203
8	0.21786	0.57324	1.52388	0.75080	0.28558	0.10092

LANGMUIR REGRESSION on TEMPERATURE=35

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The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.84078	2.84078	55.27	0.0007
Error	5	0.25701	0.05140		
Corrected Total	6	3.09779			

Root MSE 0.22672 R-Square 0.9170
 Dependent Mean 0.78670 Adj R-Sq 0.9004
 Coeff Var 28.81929

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.54244	0.19826	-2.74	0.0410
Einv	1	1.26961	0.17078	7.43	0.0007

LANGMUIR REGRESSION on TEMPERATURE=35

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Obs	Temp	E	A	EbyA	Einv
1	35	0.001	0.001	.	.
2	35	0.477	0.51	0.93529	2.09644
3	35	0.715	0.6	1.19167	1.39860
4	35	0.929	1.56	0.59551	1.07643
5	35	1.315	2.66	0.49436	0.76046
6	35	1.335	3.03	0.44059	0.74906
7	35	1.463	3.179	0.46021	0.68353
8	35	1.774	4.59	0.38649	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	1.96078	-0.74024	-0.67334	1.44791	2.11921	-0.15843
3	1.66667	-0.33547	-0.51083	1.18262	1.23324	0.43343
4	0.64103	-0.07365	0.44469	1.03751	0.82420	-0.18317
5	0.37594	0.27384	0.97833	0.87204	0.42304	-0.04710
6	0.33003	0.28893	1.10856	0.86548	0.40858	-0.07854
7	0.31456	0.38049	1.15657	0.82676	0.32537	-0.01081
8	0.21786	0.57324	1.52388	0.75080	0.17324	0.04463

FREUNDLICH REGRESSION on TEMPERATURE=35

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The REG Procedure
 Model: MODEL1
 Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	4.23757	4.23757	104.51	0.0002
Error	5	0.20274	0.04055		
Corrected Total	6	4.44032			

Root MSE 0.20137 R-Square 0.9543
 Dependent Mean 0.57541 Adj R-Sq 0.9452
 Coeff Var 34.99557

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.47992	0.07668	6.26	0.0015
LNE	1	1.82057	0.17809	10.22	0.0002

FREUNDLICH REGRESSION on TEMPERATURE=35

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Obs	Temp	E	A	EbyA	Einv
1	35	0.001	0.001	.	.
2	35	0.477	0.51	0.93529	2.09644
3	35	0.715	0.6	1.19167	1.39860
4	35	0.929	1.56	0.59551	1.07643
5	35	1.315	2.66	0.49436	0.76046
6	35	1.335	3.03	0.44059	0.74906
7	35	1.463	3.179	0.46021	0.68353
8	35	1.774	4.59	0.38649	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	1.96078	-0.74024	-0.67334	1.44791	-0.86773	0.19439
3	1.66667	-0.33547	-0.51083	1.18262	-0.13083	-0.38000
4	0.64103	-0.07365	0.44469	1.03751	0.34584	0.09884
5	0.37594	0.27384	0.97833	0.87204	0.97846	-0.00013
6	0.33003	0.28893	1.10856	0.86548	1.00594	0.10262
7	0.31456	0.38049	1.15657	0.82676	1.17263	-0.01606
8	0.21786	0.57324	1.52388	0.75080	1.52354	0.00034

COMBINED LANGMUIR REGRESSION on TEMPERATURE=35

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The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.86710	2.86710	62.14	0.0005
Error	5	0.23069	0.04614		
Corrected Total	6	3.09779			

Root MSE	0.21480	R-Square	0.9255
Dependent Mean	0.78670	Adj R-Sq	0.9106
Coeff Var	27.30381		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-2.02110	0.36532	-5.53	0.0026
CMBLNGE	1	2.81458	0.35705	7.88	0.0005

COMBINED LANGMUIR REGRESSION on TEMPERATURE=35

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25, 2003

Obs	Temp	E	A	EbyA	Einv
1	35	0.001	0.001	.	.
2	35	0.477	0.51	0.93529	2.09644
3	35	0.715	0.6	1.19167	1.39860
4	35	0.929	1.56	0.59551	1.07643
5	35	1.315	2.66	0.49436	0.76046
6	35	1.335	3.03	0.44059	0.74906
7	35	1.463	3.179	0.46021	0.68353
8	35	1.774	4.59	0.38649	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	1.96078	-0.74024	-0.67334	1.44791	2.05416	-0.09337
3	1.66667	-0.33547	-0.51083	1.18262	1.30750	0.35917
4	0.64103	-0.07365	0.44469	1.03751	0.89906	-0.25803
5	0.37594	0.27384	0.97833	0.87204	0.43333	-0.05739
6	0.33003	0.28893	1.10856	0.86548	0.41488	-0.08485
7	0.31456	0.38049	1.15657	0.82676	0.30588	0.00869
8	0.21786	0.57324	1.52388	0.75080	0.09208	0.12578

A versus E REGRESSION on TEMPERATURE=45

21

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	8.03851	8.03851	43.41	0.0006
Error	6	1.11098	0.18516		
Corrected Total	7	9.14949			

Root MSE	0.43031	R-Square	0.8786
Dependent Mean	1.63263	Adj R-Sq	0.8583
Coeff Var	26.35665		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.20621	0.31786	-0.65	0.5405
E	1	1.83677	0.27877	6.59	0.0006

A versus E REGRESSION on TEMPERATURE=45

22

25, 2003

Obs	Temp	E	A	EbyA	Einv
1	45	0.001	0.001	.	.
2	45	0.477	0.46	1.03696	2.09644
3	45	0.715	0.78	0.91667	1.39860
4	45	0.929	1.61	0.57702	1.07643
5	45	1.315	1.83	0.71858	0.76046
6	45	1.335	2.48	0.53831	0.74906
7	45	1.463	3.23	0.45294	0.68353

8		45	1.774	2.67	0.66442	0.56370
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.20437	0.20537
2	2.17391	-0.74024	-0.77653	1.44791	0.66993	-0.20993
3	1.28205	-0.33547	-0.24846	1.18262	1.10708	-0.32708
4	0.62112	-0.07365	0.47623	1.03751	1.50015	0.10985
5	0.54645	0.27384	0.60432	0.87204	2.20914	-0.37914
6	0.40323	0.28893	0.90826	0.86548	2.24588	0.23412
7	0.30960	0.38049	1.17248	0.82676	2.48098	0.74902
8	0.37453	0.57324	0.98208	0.75080	3.05222	-0.38222

EbyA versus E REGRESSION on TEMPERATURE=45

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The REG Procedure
Model: MODEL1
Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.14244	0.14244	5.84	0.0604
Error	5	0.12196	0.02439		
Corrected Total	6	0.26440			

Root MSE	0.15618	R-Square	0.5387
Dependent Mean	0.70070	Adj R-Sq	0.4465
Coeff Var	22.28886		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.08850	0.17099	6.37	0.0014
E	1	-0.33899	0.14028	-2.42	0.0604

EbyA versus E REGRESSION on TEMPERATURE=45

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Obs	Temp	E	A	EbyA	Einv	
1	45	0.001	0.001	.	.	
2	45	0.477	0.46	1.03696	2.09644	
3	45	0.715	0.78	0.91667	1.39860	
4	45	0.929	1.61	0.57702	1.07643	
5	45	1.315	1.83	0.71858	0.76046	
6	45	1.335	2.48	0.53831	0.74906	
7	45	1.463	3.23	0.45294	0.68353	
8	45	1.774	2.67	0.66442	0.56370	
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	1.08817	.
2	2.17391	-0.74024	-0.77653	1.44791	0.92681	0.11015
3	1.28205	-0.33547	-0.24846	1.18262	0.84613	0.07054
4	0.62112	-0.07365	0.47623	1.03751	0.77358	-0.19656
5	0.54645	0.27384	0.60432	0.87204	0.64273	0.07585
6	0.40323	0.28893	0.90826	0.86548	0.63595	-0.09764
7	0.30960	0.38049	1.17248	0.82676	0.59256	-0.13962
8	0.37453	0.57324	0.98208	0.75080	0.48713	0.17729

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LANGMUIR REGRESSION on TEMPERATURE=45

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The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.69862	2.69862	142.22	<.0001
Error	5	0.09487	0.01897		
Corrected Total	6	2.79349			

Root MSE	0.13775	R-Square	0.9660
Dependent Mean	0.81584	Adj R-Sq	0.9592
Coeff Var	16.88429		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.47961	0.12046	-3.98	0.0105
Einv	1	1.23743	0.10376	11.93	<.0001

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LANGMUIR REGRESSION on TEMPERATURE=45

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Obs	Temp	E	A	EbyA	Einv
1	45	0.001	0.001	.	.
2	45	0.477	0.46	1.03696	2.09644
3	45	0.715	0.78	0.91667	1.39860
4	45	0.929	1.61	0.57702	1.07643
5	45	1.315	1.83	0.71858	0.76046
6	45	1.335	2.48	0.53831	0.74906
7	45	1.463	3.23	0.45294	0.68353
8	45	1.774	2.67	0.66442	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.17391	-0.74024	-0.77653	1.44791	2.11459	0.05933
3	1.28205	-0.33547	-0.24846	1.18262	1.25106	0.03099
4	0.62112	-0.07365	0.47623	1.03751	0.85239	-0.23128
5	0.54645	0.27384	0.60432	0.87204	0.46140	0.08505
6	0.40323	0.28893	0.90826	0.86548	0.44730	-0.04408
7	0.30960	0.38049	1.17248	0.82676	0.36621	-0.05661
8	0.37453	0.57324	0.98208	0.75080	0.21793	0.15661

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FREUNDLICH REGRESSION on TEMPERATURE=45

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The REG Procedure
Model: MODEL1
Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.81331	2.81331	64.42	0.0005
Error	5	0.21836	0.04367		
Corrected Total	6	3.03167			

Root MSE 0.20898 R-Square 0.9280
 Dependent Mean 0.44548 Adj R-Sq 0.9136
 Coeff Var 46.91012

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.36768	0.07958	4.62	0.0057
LNE	1	1.48340	0.18482	8.03	0.0005

FREUNDLICH REGRESSION on TEMPERATURE=45

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25, 2003

Obs	Temp	E	A	EbyA	Einv
1	45	0.001	0.001	.	.
2	45	0.477	0.46	1.03696	2.09644
3	45	0.715	0.78	0.91667	1.39860
4	45	0.929	1.61	0.57702	1.07643
5	45	1.315	1.83	0.71858	0.76046
6	45	1.335	2.48	0.53831	0.74906
7	45	1.463	3.23	0.45294	0.68353
8	45	1.774	2.67	0.66442	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.17391	-0.74024	-0.77653	1.44791	-0.73039	-0.04614
3	1.28205	-0.33547	-0.24846	1.18262	-0.12996	-0.11850
4	0.62112	-0.07365	0.47623	1.03751	0.25843	0.21780
5	0.54645	0.27384	0.60432	0.87204	0.77389	-0.16957
6	0.40323	0.28893	0.90826	0.86548	0.79628	0.11198
7	0.30960	0.38049	1.17248	0.82676	0.93210	0.24038
8	0.37453	0.57324	0.98208	0.75080	1.21802	-0.23594

COMBINED LANGMUIR REGRESSION on TEMPERATURE=45

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25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.62004	2.62004	75.53	0.0003
Error	5	0.17345	0.03469		
Corrected Total	6	2.79349			

Root MSE 0.18625 R-Square 0.9379
 Dependent Mean 0.81584 Adj R-Sq 0.9255
 Coeff Var 22.82955

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-1.86826	0.31677	-5.90	0.0020
CMBLNGE	1	2.69058	0.30960	8.69	0.0003

COMBINED LANGMUIR REGRESSION on TEMPERATURE=45

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Obs	Temp	E	A	EbyA	EinV
1	45	0.001	0.001	.	.
2	45	0.477	0.46	1.03696	2.09644
3	45	0.715	0.78	0.91667	1.39860
4	45	0.929	1.61	0.57702	1.07643
5	45	1.315	1.83	0.71858	0.76046
6	45	1.335	2.48	0.53831	0.74906
7	45	1.463	3.23	0.45294	0.68353
8	45	1.774	2.67	0.66442	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	2.17391	-0.74024	-0.77653	1.44791	2.02746	0.14645
3	1.28205	-0.33547	-0.24846	1.18262	1.31370	-0.03164
4	0.62112	-0.07365	0.47623	1.03751	0.92325	-0.30213
5	0.54645	0.27384	0.60432	0.87204	0.47804	0.06840
6	0.40323	0.28893	0.90826	0.86548	0.46040	-0.05718
7	0.30960	0.38049	1.17248	0.82676	0.35620	-0.04661
8	0.37453	0.57324	0.98208	0.75080	0.15183	0.22270

A versus E REGRESSION on TEMPERATURE=55

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25, 2003

The REG Procedure
Model: MODEL1
Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	33.28316	33.28316	210.93	<.0001
Error	6	0.94677	0.15780		
Corrected Total	7	34.22993			

Root MSE	0.39724	R-Square	0.9723
Dependent Mean	3.57888	Adj R-Sq	0.9677
Coeff Var	11.09944		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.16281	0.29343	-0.55	0.5990
E	1	3.73748	0.25734	14.52	<.0001

A versus E REGRESSION on TEMPERATURE=55

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25, 2003

Obs	Temp	E	A	EbyA	Einv
1	55	0.001	0.001	.	.
2	55	0.477	1.03	0.46311	2.09644
3	55	0.715	2.39	0.29916	1.39860
4	55	0.929	3.93	0.23639	1.07643
5	55	1.315	4.63	0.28402	0.76046
6	55	1.335	5.07	0.26331	0.74906
7	55	1.463	5.41	0.27043	0.68353
8	55	1.774	6.17	0.28752	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.15908	0.16008
2	0.97087	-0.74024	0.02956	1.44791	1.61997	-0.58997
3	0.41841	-0.33547	0.87129	1.18262	2.50949	-0.11949
4	0.25445	-0.07365	1.36864	1.03751	3.30931	0.62069
5	0.21598	0.27384	1.53256	0.87204	4.75198	-0.12198
6	0.19724	0.28893	1.62334	0.86548	4.82673	0.24327
7	0.18484	0.38049	1.68825	0.82676	5.30513	0.10487
8	0.16207	0.57324	1.81970	0.75080	6.46748	-0.29748

EbyA versus E REGRESSION on TEMPERATURE=55

33

18:32 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.01197	0.01197	2.81	0.1547
Error	5	0.02131	0.00426		
Corrected Total	6	0.03328			

Root MSE	0.06529	R-Square	0.3596
Dependent Mean	0.30056	Adj R-Sq	0.2315
Coeff Var	21.72254		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.41297	0.07148	5.78	0.0022
E	1	-0.09826	0.05864	-1.68	0.1547

EbyA versus E REGRESSION on TEMPERATURE=55

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Obs	Temp	E	A	EbyA	Einv
1	55	0.001	0.001	.	.
2	55	0.477	1.03	0.46311	2.09644
3	55	0.715	2.39	0.29916	1.39860
4	55	0.929	3.93	0.23639	1.07643
5	55	1.315	4.63	0.28402	0.76046
6	55	1.335	5.07	0.26331	0.74906
7	55	1.463	5.41	0.27043	0.68353
8	55	1.774	6.17	0.28752	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	0.41287	.
2	0.97087	-0.74024	0.02956	1.44791	0.36610	0.097008
3	0.41841	-0.33547	0.87129	1.18262	0.34271	-0.043551
4	0.25445	-0.07365	1.36864	1.03751	0.32169	-0.085300
5	0.21598	0.27384	1.53256	0.87204	0.28376	0.000257
6	0.19724	0.28893	1.62334	0.86548	0.28179	-0.018481
7	0.18484	0.38049	1.68825	0.82676	0.26922	0.001207
8	0.16207	0.57324	1.81970	0.75080	0.23866	0.048860

LANGMUIR REGRESSION on TEMPERATURE=55

35

18:32 Saturday, October

25, 2003

The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.46738	0.46738	65.82	0.0005
Error	5	0.03550	0.00710		
Corrected Total	6	0.50288			

Root MSE	0.08427	R-Square	0.9294
Dependent Mean	0.34341	Adj R-Sq	0.9153
Coeff Var	24.53787		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.19571	0.07369	-2.66	0.0451
Einv	1	0.51497	0.06347	8.11	0.0005

LANGMUIR REGRESSION on TEMPERATURE=55

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Obs	Temp	E	A	EbyA	Einv
1	55	0.001	0.001	.	.
2	55	0.477	1.03	0.46311	2.09644
3	55	0.715	2.39	0.29916	1.39860
4	55	0.929	3.93	0.23639	1.07643
5	55	1.315	4.63	0.28402	0.76046
6	55	1.335	5.07	0.26331	0.74906
7	55	1.463	5.41	0.27043	0.68353
8	55	1.774	6.17	0.28752	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	0.97087	-0.74024	0.02956	1.44791	0.88390	0.08697
3	0.41841	-0.33547	0.87129	1.18262	0.52453	-0.10612
4	0.25445	-0.07365	1.36864	1.03751	0.35862	-0.10417
5	0.21598	0.27384	1.53256	0.87204	0.19591	0.02008
6	0.19724	0.28893	1.62334	0.86548	0.19004	0.00720
7	0.18484	0.38049	1.68825	0.82676	0.15629	0.02855
8	0.16207	0.57324	1.81970	0.75080	0.09458	0.06749

FREUNDLICH REGRESSION on TEMPERATURE=55

37

18:32 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.23573	2.23573	78.57	0.0003
Error	5	0.14228	0.02846		
Corrected Total	6	2.37801			

Root MSE	0.16869	R-Square	0.9402
Dependent Mean	1.27619	Adj R-Sq	0.9282
Coeff Var	13.21828		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.20683	0.06424	18.79	<.0001
LNE	1	1.32239	0.14919	8.86	0.0003

FREUNDLICH REGRESSION on TEMPERATURE=55

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Obs	Temp	E	A	EbyA	Einv
1	55	0.001	0.001	.	.
2	55	0.477	1.03	0.46311	2.09644
3	55	0.715	2.39	0.29916	1.39860
4	55	0.929	3.93	0.23639	1.07643
5	55	1.315	4.63	0.28402	0.76046
6	55	1.335	5.07	0.26331	0.74906
7	55	1.463	5.41	0.27043	0.68353
8	55	1.774	6.17	0.28752	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	0.97087	-0.74024	0.02956	1.44791	0.22795	-0.19839
3	0.41841	-0.33547	0.87129	1.18262	0.76321	0.10808
4	0.25445	-0.07365	1.36864	1.03751	1.10945	0.25919
5	0.21598	0.27384	1.53256	0.87204	1.56895	-0.03640
6	0.19724	0.28893	1.62334	0.86548	1.58891	0.03443
7	0.18484	0.38049	1.68825	0.82676	1.70999	-0.02174
8	0.16207	0.57324	1.81970	0.75080	1.96488	-0.14518

COMBINED LANGMUIR REGRESSION on TEMPERATURE=55

39

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25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Sum of	Mean
--------	------

Source	DF	Squares	Square	F Value	Pr > F
Model	1	0.44216	0.44216	36.41	0.0018
Error	5	0.06072	0.01214		
Corrected Total	6	0.50288			

Root MSE	0.11020	R-Square	0.8793
Dependent Mean	0.34341	Adj R-Sq	0.8551
Coeff Var	32.08999		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.75923	0.18742	-4.05	0.0098
CMBLNGE	1	1.10530	0.18318	6.03	0.0018

COMBINED LANGMUIR REGRESSION on TEMPERATURE=55

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25, 2003

Obs	Temp	E	A	EbyA	Einv
1	55	0.001	0.001	.	.
2	55	0.477	1.03	0.46311	2.09644
3	55	0.715	2.39	0.29916	1.39860
4	55	0.929	3.93	0.23639	1.07643
5	55	1.315	4.63	0.28402	0.76046
6	55	1.335	5.07	0.26331	0.74906
7	55	1.463	5.41	0.27043	0.68353
8	55	1.774	6.17	0.28752	0.56370

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	0.97087	-0.74024	0.02956	1.44791	0.84115	0.12972
3	0.41841	-0.33547	0.87129	1.18262	0.54793	-0.12952
4	0.25445	-0.07365	1.36864	1.03751	0.38754	-0.13308
5	0.21598	0.27384	1.53256	0.87204	0.20464	0.01134
6	0.19724	0.28893	1.62334	0.86548	0.19740	-0.00016
7	0.18484	0.38049	1.68825	0.82676	0.15459	0.03025
8	0.16207	0.57324	1.81970	0.75080	0.07063	0.09144

Appendix C

Statistical Analysis for Experimental Adsorption Data of EG
II to fit Adsorption Models

SAS CODE

```
%let path=D:\Dole Files\;
libname bus "&path.";
/* THIS SAS FILE IS FOR REGRESSION FOR DATA ON EGIIa FILE */

/* This step makes new variables for calculating different models */

data temp;
set bus.egIIa;
if E>0.001 then do;
m=2;
EbyA=E/A;
Einv=1/E;
Ainv=1/A;
LNE=log(E);
LNA=log(A);
CMBLNGE=1/(E**(1/m));
drop m;
end;
run;

/*****
/* regression models ..... FOR TEMPERATURE=25 */
*****/

/* Selecting Cases */

data temp25;
set temp;
if TEMP=25 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
  MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
  MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
  MODEL Ainv=Einv;
```

```

OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=25';
PROC REG data=temp25;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

/*****
/* regression models ..... FOR TEMPERATURE=35 */
*****/

/* Selecting Cases */

data temp35;
set temp;
if TEMP=35 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;

```

```

MODEL Ainv=Einv;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=35';
PROC REG data=temp35;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

/*****
/* regression models ..... FOR TEMPERATURE=45 */
*****/

/* Selecting Cases */

data temp45;
set temp;
if TEMP=45 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;

```

```

MODEL Ainv=Einv;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=45';
PROC REG data=temp45;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

/*****
/* regression models ..... FOR TEMPERATURE=55 */
*****/

/* Selecting Cases */

data temp55;
set temp;
if TEMP=55 then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */

```

```

title 'LANGMUIR REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
  MODEL Ainv=Einv;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on TEMPERATURE=55';
PROC REG data=temp55;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

```

SAS OUTPUT

A versus E REGRESSION on TEMPERATURE=25

1

18:38 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	9.55896	9.55896	140.83	<.0001
Error	6	0.40724	0.06787		
Corrected Total	7	9.96620			

Root MSE	0.26053	R-Square	0.9591
Dependent Mean	1.68526	Adj R-Sq	0.9523
Coeff Var	15.45907		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.06860	0.17414	-0.39	0.7072
E	1	1.61805	0.13634	11.87	<.0001

A versus E REGRESSION on TEMPERATURE=25

2

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25, 2003

Obs	Temp	E	A	EbyA	EinV
1	25	0.001	0.001	.	.
2	25	0.3619	0.21	1.72333	2.76319
3	25	0.6896	1.03	0.66951	1.45012
4	25	0.997	1.93	0.51658	1.00301
5	25	1.14	1.62	0.70370	0.87719
6	25	1.556	2.71	0.57417	0.64267
7	25	1.857	2.96	0.62736	0.53850
8	25	2.069	3.021111111	0.68485	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.06699	0.06799
2	4.76190	-1.01639	-1.56065	1.66229	0.51697	-0.30697
3	0.97087	-0.37164	0.02956	1.20421	1.04721	-0.01721
4	0.51813	-0.00300	0.65752	1.00150	1.54459	0.38541
5	0.61728	0.13103	0.48243	0.93659	1.77598	-0.15598
6	0.36900	0.44212	0.99695	0.80167	2.44909	0.26091
7	0.33784	0.61896	1.08519	0.73383	2.93612	0.02388
8	0.33100	0.72707	1.10562	0.69522	3.27915	-0.25804

EbyA versus E REGRESSION on TEMPERATURE=25

3

18:38 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.37746	0.37746	2.80	0.1552
Error	5	0.67433	0.13487		
Corrected Total	6	1.05179			

Root MSE	0.36724	R-Square	0.3589
Dependent Mean	0.78564	Adj R-Sq	0.2306
Coeff Var	46.74403		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.28625	0.32986	3.90	0.0114
E	1	-0.40416	0.24158	-1.67	0.1552

EbyA versus E REGRESSION on TEMPERATURE=25

4

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25, 2003

Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.3619	0.21	1.72333	2.76319
3	25	0.6896	1.03	0.66951	1.45012
4	25	0.997	1.93	0.51658	1.00301
5	25	1.14	1.62	0.70370	0.87719
6	25	1.556	2.71	0.57417	0.64267
7	25	1.857	2.96	0.62736	0.53850
8	25	2.069	3.021111111	0.68485	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	1.28585	.
2	4.76190	-1.01639	-1.56065	1.66229	1.13999	0.58335
3	0.97087	-0.37164	0.02956	1.20421	1.00754	-0.33803
4	0.51813	-0.00300	0.65752	1.00150	0.88331	-0.36673
5	0.61728	0.13103	0.48243	0.93659	0.82551	-0.12181
6	0.36900	0.44212	0.99695	0.80167	0.65738	-0.08321
7	0.33784	0.61896	1.08519	0.73383	0.53573	0.09163
8	0.33100	0.72707	1.10562	0.69522	0.45005	0.23480

LANGMUIR REGRESSION on TEMPERATURE=25

5

18:38 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	14.38528	14.38528	54.78	0.0007
Error	5	1.31306	0.26261		
Corrected Total	6	15.69834			

Root MSE	0.51246	R-Square	0.9164
Dependent Mean	1.12943	Adj R-Sq	0.8996

Coeff Var 45.37291

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-1.01229	0.34822	-2.91	0.0335
Einv	1	1.93246	0.26110	7.40	0.0007

LANGMUIR REGRESSION on TEMPERATURE=25

6

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25, 2003

Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.3619	0.21	1.72333	2.76319
3	25	0.6896	1.03	0.66951	1.45012
4	25	0.997	1.93	0.51658	1.00301
5	25	1.14	1.62	0.70370	0.87719
6	25	1.556	2.71	0.57417	0.64267
7	25	1.857	2.96	0.62736	0.53850
8	25	2.069	3.021111111	0.68485	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	4.76190	-1.01639	-1.56065	1.66229	4.32748	0.43442
3	0.97087	-0.37164	0.02956	1.20421	1.79001	-0.81913
4	0.51813	-0.00300	0.65752	1.00150	0.92599	-0.40785
5	0.61728	0.13103	0.48243	0.93659	0.68285	-0.06557
6	0.36900	0.44212	0.99695	0.80167	0.22965	0.13935
7	0.33784	0.61896	1.08519	0.73383	0.02835	0.30949
8	0.33100	0.72707	1.10562	0.69522	-0.07828	0.40929

FREUNDLICH REGRESSION on TEMPERATURE=25

7

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25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	4.95083	4.95083	57.92	0.0006
Error	5	0.42738	0.08548		
Corrected Total	6	5.37821			

Root MSE 0.29236 R-Square 0.9205
 Dependent Mean 0.39952 Adj R-Sq 0.9046
 Coeff Var 73.17906

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.28774	0.11147	2.58	0.0494
LNE	1	1.48149	0.19466	7.61	0.0006

FREUNDLICH REGRESSION on TEMPERATURE=25

8

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Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.3619	0.21	1.72333	2.76319
3	25	0.6896	1.03	0.66951	1.45012
4	25	0.997	1.93	0.51658	1.00301
5	25	1.14	1.62	0.70370	0.87719
6	25	1.556	2.71	0.57417	0.64267
7	25	1.857	2.96	0.62736	0.53850
8	25	2.069	3.021111111	0.68485	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	4.76190	-1.01639	-1.56065	1.66229	-1.21803	-0.34262
3	0.97087	-0.37164	0.02956	1.20421	-0.26284	0.29240
4	0.51813	-0.00300	0.65752	1.00150	0.28329	0.37423
5	0.61728	0.13103	0.48243	0.93659	0.48186	0.00057
6	0.36900	0.44212	0.99695	0.80167	0.94273	0.05421
7	0.33784	0.61896	1.08519	0.73383	1.20473	-0.11954
8	0.33100	0.72707	1.10562	0.69522	1.36488	-0.25926

COMBINED LANGMUIR REGRESSION on TEMPERATURE=25

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25, 2003

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The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	13.18129	13.18129	26.18	0.0037
Error	5	2.51705	0.50341		
Corrected Total	6	15.69834			

Root MSE	0.70951	R-Square	0.8397
Dependent Mean	1.12943	Adj R-Sq	0.8076
Coeff Var	62.82030		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-3.27213	0.90101	-3.63	0.0150
CMBLNGE	1	4.37949	0.85587	5.12	0.0037

COMBINED LANGMUIR REGRESSION on TEMPERATURE=25

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Obs	Temp	E	A	EbyA	Einv
1	25	0.001	0.001	.	.
2	25	0.3619	0.21	1.72333	2.76319
3	25	0.6896	1.03	0.66951	1.45012
4	25	0.997	1.93	0.51658	1.00301
5	25	1.14	1.62	0.70370	0.87719
6	25	1.556	2.71	0.57417	0.64267

7	25	1.857	2.96	0.62736	0.53850	
8	25	2.069	3.021111111	0.68485	0.48333	
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	4.76190	-1.01639	-1.56065	1.66229	4.00782	0.75408
3	0.97087	-0.37164	0.02956	1.20421	2.00168	-1.03080
4	0.51813	-0.00300	0.65752	1.00150	1.11394	-0.59580
5	0.61728	0.13103	0.48243	0.93659	0.82963	-0.21235
6	0.36900	0.44212	0.99695	0.80167	0.23877	0.13024
7	0.33784	0.61896	1.08519	0.73383	-0.05835	0.39618
8	0.33100	0.72707	1.10562	0.69522	-0.22744	0.55845

A versus E REGRESSION on TEMPERATURE=35

11

18:38 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	10.70670	10.70670	83.44	<.0001
Error	6	0.76992	0.12832		
Corrected Total	7	11.47662			

Root MSE	0.35822	R-Square	0.9329
Dependent Mean	1.76776	Adj R-Sq	0.9217
Coeff Var	20.26387		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.08841	0.23944	-0.37	0.7246
E	1	1.71244	0.18747	9.13	<.0001

A versus E REGRESSION on TEMPERATURE=35

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Obs	Temp	E	A	EbyA	Einv	
1	35	0.001	0.001	.	.	
2	35	0.3619	0.12	3.01583	2.76319	
3	35	0.6896	1.11	0.62126	1.45012	
4	35	0.997	2.01	0.49602	1.00301	
5	35	1.14	1.73	0.65896	0.87719	
6	35	1.556	2.85	0.54596	0.64267	
7	35	1.857	3.38	0.54941	0.53850	
8	35	2.069	2.941111111	0.70348	0.48333	
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.08670	0.08770
2	8.33333	-1.01639	-2.12026	1.66229	0.53132	-0.41132
3	0.90090	-0.37164	0.10436	1.20421	1.09248	0.01752
4	0.49751	-0.00300	0.69813	1.00150	1.61889	0.39111
5	0.57803	0.13103	0.54812	0.93659	1.86377	-0.13377
6	0.35088	0.44212	1.04732	0.80167	2.57614	0.27386
7	0.29586	0.61896	1.21788	0.73383	3.09159	0.28841

8 0.34001 0.72707 1.07879 0.69522 3.45462 -0.51351

EbyA versus E REGRESSION on TEMPERATURE=35

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The REG Procedure
Model: MODEL1
Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	1.85954	1.85954	2.91	0.1485
Error	5	3.19099	0.63820		
Corrected Total	6	5.05053			

Root MSE	0.79887	R-Square	0.3682
Dependent Mean	0.94156	Adj R-Sq	0.2418
Coeff Var	84.84563		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	2.05269	0.71756	2.86	0.0354
E	1	-0.89706	0.52553	-1.71	0.1485

EbyA versus E REGRESSION on TEMPERATURE=35

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Obs	Temp	E	A	EbyA	Einvs
1	35	0.001	0.001	.	.
2	35	0.3619	0.12	3.01583	2.76319
3	35	0.6896	1.11	0.62126	1.45012
4	35	0.997	2.01	0.49602	1.00301
5	35	1.14	1.73	0.65896	0.87719
6	35	1.556	2.85	0.54596	0.64267
7	35	1.857	3.38	0.54941	0.53850
8	35	2.069	2.941111111	0.70348	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	2.05180	.
2	8.33333	-1.01639	-2.12026	1.66229	1.72805	1.28778
3	0.90090	-0.37164	0.10436	1.20421	1.43408	-0.81282
4	0.49751	-0.00300	0.69813	1.00150	1.15833	-0.66231
5	0.57803	0.13103	0.54812	0.93659	1.03005	-0.37109
6	0.35088	0.44212	1.04732	0.80167	0.65687	-0.11091
7	0.29586	0.61896	1.21788	0.73383	0.38686	0.16255
8	0.34001	0.72707	1.07879	0.69522	0.19668	0.50679

LANGMUIR REGRESSION on TEMPERATURE=35

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The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	46.39448	46.39448	35.47	0.0019
Error	5	6.53927	1.30785		
Corrected Total	6	52.93375			

Root MSE	1.14361	R-Square	0.8765
Dependent Mean	1.61379	Adj R-Sq	0.8518
Coeff Var	70.86518		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-2.23247	0.77709	-2.87	0.0349
Ein v	1	3.47045	0.58268	5.96	0.0019

LANGMUIR REGRESSION on TEMPERATURE=35

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Obs	Temp	E	A	EbyA	Ein v
1	35	0.001	0.001	.	.
2	35	0.3619	0.12	3.01583	2.76319
3	35	0.6896	1.11	0.62126	1.45012
4	35	0.997	2.01	0.49602	1.00301
5	35	1.14	1.73	0.65896	0.87719
6	35	1.556	2.85	0.54596	0.64267
7	35	1.857	3.38	0.54941	0.53850
8	35	2.069	2.941111111	0.70348	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	8.33333	-1.01639	-2.12026	1.66229	7.35706	0.97628
3	0.90090	-0.37164	0.10436	1.20421	2.80009	-1.89919
4	0.49751	-0.00300	0.69813	1.00150	1.24842	-0.75091
5	0.57803	0.13103	0.54812	0.93659	0.81179	-0.23375
6	0.35088	0.44212	1.04732	0.80167	-0.00210	0.35298
7	0.29586	0.61896	1.21788	0.73383	-0.36362	0.65948
8	0.34001	0.72707	1.07879	0.69522	-0.55511	0.89512

FREUNDLICH REGRESSION on TEMPERATURE=35

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25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	7.06807	7.06807	34.54	0.0020
Error	5	1.02331	0.20466		
Corrected Total	6	8.09138			

Root MSE	0.45239	R-Square	0.8735
Dependent Mean	0.36776	Adj R-Sq	0.8482
Coeff Var	123.01293		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.23421	0.17249	1.36	0.2326
LNE	1	1.77015	0.30122	5.88	0.0020

FREUNDLICH REGRESSION on TEMPERATURE=35

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Obs	Temp	E	A	EbyA	Einvs
1	35	0.001	0.001	.	.
2	35	0.3619	0.12	3.01583	2.76319
3	35	0.6896	1.11	0.62126	1.45012
4	35	0.997	2.01	0.49602	1.00301
5	35	1.14	1.73	0.65896	0.87719
6	35	1.556	2.85	0.54596	0.64267
7	35	1.857	3.38	0.54941	0.53850
8	35	2.069	2.941111111	0.70348	0.48333

Obs	Ainvs	LNE	LNA	CMBLNGE	PRED	RESID
1
2	8.33333	-1.01639	-2.12026	1.66229	-1.56495	-0.55531
3	0.90090	-0.37164	0.10436	1.20421	-0.42366	0.52802
4	0.49751	-0.00300	0.69813	1.00150	0.22889	0.46925
5	0.57803	0.13103	0.54812	0.93659	0.46615	0.08197
6	0.35088	0.44212	1.04732	0.80167	1.01682	0.03050
7	0.29586	0.61896	1.21788	0.73383	1.32986	-0.11199
8	0.34001	0.72707	1.07879	0.69522	1.52122	-0.44243

COMBINED LANGMUIR REGRESSION on TEMPERATURE=35

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The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainvs

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	41.77570	41.77570	18.72	0.0075
Error	5	11.15806	2.23161		
Corrected Total	6	52.93375			

Root MSE	1.49386	R-Square	0.7892
Dependent Mean	1.61379	Adj R-Sq	0.7470
Coeff Var	92.56834		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-6.22214	1.89705	-3.28	0.0220
CMBLNGE	1	7.79661	1.80199	4.33	0.0075

COMBINED LANGMUIR REGRESSION on TEMPERATURE=35

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25, 2003

Obs	Temp	E	A	EbyA	Einv
1	35	0.001	0.001	.	.
2	35	0.3619	0.12	3.01583	2.76319
3	35	0.6896	1.11	0.62126	1.45012
4	35	0.997	2.01	0.49602	1.00301
5	35	1.14	1.73	0.65896	0.87719
6	35	1.556	2.85	0.54596	0.64267
7	35	1.857	3.38	0.54941	0.53850
8	35	2.069	2.941111111	0.70348	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	8.33333	-1.01639	-2.12026	1.66229	6.73806	1.59527
3	0.90090	-0.37164	0.10436	1.20421	3.16660	-2.26570
4	0.49751	-0.00300	0.69813	1.00150	1.58620	-1.08869
5	0.57803	0.13103	0.54812	0.93659	1.08006	-0.50203
6	0.35088	0.44212	1.04732	0.80167	0.02817	0.32271
7	0.29586	0.61896	1.21788	0.73383	-0.50077	0.79662
8	0.34001	0.72707	1.07879	0.69522	-0.80181	1.14181

A versus E REGRESSION on TEMPERATURE=45

21

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	17.61561	17.61561	210.08	<.0001
Error	6	0.50311	0.08385		
Corrected Total	7	18.11872			

Root MSE	0.28957	R-Square	0.9722
Dependent Mean	2.07249	Adj R-Sq	0.9676
Coeff Var	13.97223		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.30841	0.19356	-1.59	0.1622
E	1	2.19653	0.15155	14.49	<.0001

A versus E REGRESSION on TEMPERATURE=45

22

25, 2003

Obs	Temp	E	A	EbyA	Einv
1	45	0.001	0.001	.	.
2	45	0.3619	0.2	1.80950	2.76319
3	45	0.6896	1.17	0.58940	1.45012
4	45	0.997	1.92	0.51927	1.00301
5	45	1.14	1.89	0.60317	0.87719
6	45	1.556	3.23	0.48173	0.64267
7	45	1.857	4.17	0.44532	0.53850

8		45	2.069	3.998888889	0.51739	0.48333
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.30621	0.30721
2	5.00000	-1.01639	-1.60944	1.66229	0.48651	-0.28651
3	0.85470	-0.37164	0.15700	1.20421	1.20631	-0.03631
4	0.52083	-0.00300	0.65233	1.00150	1.88153	0.03847
5	0.52910	0.13103	0.63658	0.93659	2.19563	-0.30563
6	0.30960	0.44212	1.17248	0.80167	3.10938	0.12062
7	0.23981	0.61896	1.42792	0.73383	3.77054	0.39946
8	0.25007	0.72707	1.38602	0.69522	4.23620	-0.23731

EbyA versus E REGRESSION on TEMPERATURE=45

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The REG Procedure
Model: MODEL1
Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.66237	0.66237	4.31	0.0925
Error	5	0.76812	0.15362		
Corrected Total	6	1.43049			

Root MSE	0.39195	R-Square	0.4630
Dependent Mean	0.70940	Adj R-Sq	0.3556
Coeff Var	55.25085		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.37255	0.35206	3.90	0.0114
E	1	-0.53538	0.25784	-2.08	0.0925

EbyA versus E REGRESSION on TEMPERATURE=45

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Obs	Temp	E	A	EbyA	Einv	
1	45	0.001	0.001	.	.	
2	45	0.3619	0.2	1.80950	2.76319	
3	45	0.6896	1.17	0.58940	1.45012	
4	45	0.997	1.92	0.51927	1.00301	
5	45	1.14	1.89	0.60317	0.87719	
6	45	1.556	3.23	0.48173	0.64267	
7	45	1.857	4.17	0.44532	0.53850	
8	45	2.069	3.998888889	0.51739	0.48333	
Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	1.37201	.
2	5.00000	-1.01639	-1.60944	1.66229	1.17879	0.63071
3	0.85470	-0.37164	0.15700	1.20421	1.00335	-0.41395
4	0.52083	-0.00300	0.65233	1.00150	0.83877	-0.31950
5	0.52910	0.13103	0.63658	0.93659	0.76221	-0.15904
6	0.30960	0.44212	1.17248	0.80167	0.53949	-0.05776
7	0.23981	0.61896	1.42792	0.73383	0.37834	0.06698
8	0.25007	0.72707	1.38602	0.69522	0.26484	0.25255

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LANGMUIR REGRESSION on TEMPERATURE=45

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The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	16.41774	16.41774	51.28	0.0008
Error	5	1.60084	0.32017		
Corrected Total	6	18.01858			

Root MSE	0.56583	R-Square	0.9112
Dependent Mean	1.10059	Adj R-Sq	0.8934
Coeff Var	51.41192		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-1.18744	0.38449	-3.09	0.0272
Einv	1	2.06447	0.28830	7.16	0.0008

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LANGMUIR REGRESSION on TEMPERATURE=45

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Obs	Temp	E	A	EbyA	Einv
1	45	0.001	0.001	.	.
2	45	0.3619	0.2	1.80950	2.76319
3	45	0.6896	1.17	0.58940	1.45012
4	45	0.997	1.92	0.51927	1.00301
5	45	1.14	1.89	0.60317	0.87719
6	45	1.556	3.23	0.48173	0.64267
7	45	1.857	4.17	0.44532	0.53850
8	45	2.069	3.998888889	0.51739	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	5.00000	-1.01639	-1.60944	1.66229	4.51710	0.48290
3	0.85470	-0.37164	0.15700	1.20421	1.80628	-0.95158
4	0.52083	-0.00300	0.65233	1.00150	0.88324	-0.36241
5	0.52910	0.13103	0.63658	0.93659	0.62350	-0.09440
6	0.30960	0.44212	1.17248	0.80167	0.13934	0.17026
7	0.23981	0.61896	1.42792	0.73383	-0.07572	0.31553
8	0.25007	0.72707	1.38602	0.69522	-0.18963	0.43970

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FREUNDLICH REGRESSION on TEMPERATURE=45

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The REG Procedure
Model: MODEL1
Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	6.33572	6.33572	88.76	0.0002
Error	5	0.35690	0.07138		
Corrected Total	6	6.69262			

Root MSE 0.26717 R-Square 0.9467
 Dependent Mean 0.54613 Adj R-Sq 0.9360
 Coeff Var 48.92089

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.41968	0.10187	4.12	0.0092
LNE	1	1.67594	0.17789	9.42	0.0002

FREUNDLICH REGRESSION on TEMPERATURE=45

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25, 2003

Obs	Temp	E	A	EbyA	EinV
1	45	0.001	0.001	.	.
2	45	0.3619	0.2	1.80950	2.76319
3	45	0.6896	1.17	0.58940	1.45012
4	45	0.997	1.92	0.51927	1.00301
5	45	1.14	1.89	0.60317	0.87719
6	45	1.556	3.23	0.48173	0.64267
7	45	1.857	4.17	0.44532	0.53850
8	45	2.069	3.998888889	0.51739	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	5.00000	-1.01639	-1.60944	1.66229	-1.28372	-0.32572
3	0.85470	-0.37164	0.15700	1.20421	-0.20317	0.36018
4	0.52083	-0.00300	0.65233	1.00150	0.41464	0.23768
5	0.52910	0.13103	0.63658	0.93659	0.63927	-0.00270
6	0.30960	0.44212	1.17248	0.80167	1.16064	0.01184
7	0.23981	0.61896	1.42792	0.73383	1.45702	-0.02910
8	0.25007	0.72707	1.38602	0.69522	1.63819	-0.25218

COMBINED LANGMUIR REGRESSION on TEMPERATURE=45

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25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	15.02045	15.02045	25.05	0.0041
Error	5	2.99813	0.59963		
Corrected Total	6	18.01858			

Root MSE 0.77436 R-Square 0.8336
 Dependent Mean 1.10059 Adj R-Sq 0.8003
 Coeff Var 70.35839

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-3.59803	0.98336	-3.66	0.0146
CMBLNGE	1	4.67504	0.93408	5.00	0.0041

COMBINED LANGMUIR REGRESSION on TEMPERATURE=45

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25, 2003

Obs	Temp	E	A	EbyA	Einvs
1	45	0.001	0.001	.	.
2	45	0.3619	0.2	1.80950	2.76319
3	45	0.6896	1.17	0.58940	1.45012
4	45	0.997	1.92	0.51927	1.00301
5	45	1.14	1.89	0.60317	0.87719
6	45	1.556	3.23	0.48173	0.64267
7	45	1.857	4.17	0.44532	0.53850
8	45	2.069	3.998888889	0.51739	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	5.00000	-1.01639	-1.60944	1.66229	4.17323	0.82677
3	0.85470	-0.37164	0.15700	1.20421	2.03169	-1.17699
4	0.52083	-0.00300	0.65233	1.00150	1.08404	-0.56321
5	0.52910	0.13103	0.63658	0.93659	0.78055	-0.25145
6	0.30960	0.44212	1.17248	0.80167	0.14981	0.15979
7	0.23981	0.61896	1.42792	0.73383	-0.16735	0.40716
8	0.25007	0.72707	1.38602	0.69522	-0.34786	0.59793

A versus E REGRESSION on TEMPERATURE=55

31

18:38 Saturday, October

25, 2003

The REG Procedure
Model: MODEL1
Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	23.69622	23.69622	192.06	<.0001
Error	6	0.74028	0.12338		
Corrected Total	7	24.43650			

Root MSE	0.35125	R-Square	0.9697
Dependent Mean	2.24749	Adj R-Sq	0.9647
Coeff Var	15.62878		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.51393	0.23479	-2.19	0.0712
E	1	2.54757	0.18383	13.86	<.0001

A versus E REGRESSION on TEMPERATURE=55

32

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25, 2003

Obs	Temp	E	A	EbyA	EinV
1	55	0.001	0.001	.	.
2	55	0.3619	0.18	2.01056	2.76319
3	55	0.6896	0.97	0.71093	1.45012
4	55	0.997	1.85	0.53892	1.00301
5	55	1.14	2.04	0.55882	0.87719
6	55	1.556	3.75	0.41493	0.64267
7	55	1.857	4.53	0.40993	0.53850
8	55	2.069	4.658888889	0.44410	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.51138	0.51238
2	5.55556	-1.01639	-1.71480	1.66229	0.40804	-0.22804
3	1.03093	-0.37164	-0.03046	1.20421	1.24288	-0.27288
4	0.54054	-0.00300	0.61519	1.00150	2.02601	-0.17601
5	0.49020	0.13103	0.71295	0.93659	2.39031	-0.35031
6	0.26667	0.44212	1.32176	0.80167	3.45010	0.29990
7	0.22075	0.61896	1.51072	0.73383	4.21692	0.31308
8	0.21464	0.72707	1.53878	0.69522	4.75701	-0.09812

EbyA versus E REGRESSION on TEMPERATURE=55

33

18:38 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	1.08646	1.08646	6.02	0.0577
Error	5	0.90292	0.18058		
Corrected Total	6	1.98938			

Root MSE	0.42495	R-Square	0.5461
Dependent Mean	0.72688	Adj R-Sq	0.4554
Coeff Var	58.46210		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.57620	0.38170	4.13	0.0091
E	1	-0.68568	0.27955	-2.45	0.0577

EbyA versus E REGRESSION on TEMPERATURE=55

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Obs	Temp	E	A	EbyA	EinV
1	55	0.001	0.001	.	.
2	55	0.3619	0.18	2.01056	2.76319
3	55	0.6896	0.97	0.71093	1.45012
4	55	0.997	1.85	0.53892	1.00301
5	55	1.14	2.04	0.55882	0.87719
6	55	1.556	3.75	0.41493	0.64267
7	55	1.857	4.53	0.40993	0.53850
8	55	2.069	4.658888889	0.44410	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1	1.57552	.
2	5.55556	-1.01639	-1.71480	1.66229	1.32805	0.68250
3	1.03093	-0.37164	-0.03046	1.20421	1.10335	-0.39243
4	0.54054	-0.00300	0.61519	1.00150	0.89257	-0.35366
5	0.49020	0.13103	0.71295	0.93659	0.79452	-0.23570
6	0.26667	0.44212	1.32176	0.80167	0.50928	-0.09434
7	0.22075	0.61896	1.51072	0.73383	0.30289	0.10705
8	0.21464	0.72707	1.53878	0.69522	0.15752	0.28658

LANGMUIR REGRESSION on TEMPERATURE=55

35

18:38 Saturday, October

25, 2003

The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	20.99046	20.99046	60.05	0.0006
Error	5	1.74773	0.34955		
Corrected Total	6	22.73820			

Root MSE	0.59122	R-Square	0.9231
Dependent Mean	1.18847	Adj R-Sq	0.9078
Coeff Var	49.74679		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-1.39865	0.40174	-3.48	0.0176
Einv	1	2.33434	0.30123	7.75	0.0006

LANGMUIR REGRESSION on TEMPERATURE=55

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Obs	Temp	E	A	EbyA	Einv
1	55	0.001	0.001	.	.
2	55	0.3619	0.18	2.01056	2.76319
3	55	0.6896	0.97	0.71093	1.45012
4	55	0.997	1.85	0.53892	1.00301
5	55	1.14	2.04	0.55882	0.87719
6	55	1.556	3.75	0.41493	0.64267
7	55	1.857	4.53	0.40993	0.53850
8	55	2.069	4.658888889	0.44410	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	5.55556	-1.01639	-1.71480	1.66229	5.05158	0.50398
3	1.03093	-0.37164	-0.03046	1.20421	1.98641	-0.95548
4	0.54054	-0.00300	0.61519	1.00150	0.94271	-0.40217
5	0.49020	0.13103	0.71295	0.93659	0.64902	-0.15882
6	0.26667	0.44212	1.32176	0.80167	0.10157	0.16510
7	0.22075	0.61896	1.51072	0.73383	-0.14160	0.36235
8	0.21464	0.72707	1.53878	0.69522	-0.27040	0.48505

FREUNDLICH REGRESSION on TEMPERATURE=55

37

18:38 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	7.76702	7.76702	172.79	<.0001
Error	5	0.22475	0.04495		
Corrected Total	6	7.99177			

Root MSE	0.21201	R-Square	0.9719
Dependent Mean	0.56488	Adj R-Sq	0.9663
Coeff Var	37.53259		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.42487	0.08084	5.26	0.0033
LNE	1	1.85561	0.14116	13.15	<.0001

FREUNDLICH REGRESSION on TEMPERATURE=55

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25, 2003

Obs	Temp	E	A	EbyA	Einv
1	55	0.001	0.001	.	.
2	55	0.3619	0.18	2.01056	2.76319
3	55	0.6896	0.97	0.71093	1.45012
4	55	0.997	1.85	0.53892	1.00301
5	55	1.14	2.04	0.55882	0.87719
6	55	1.556	3.75	0.41493	0.64267
7	55	1.857	4.53	0.40993	0.53850
8	55	2.069	4.658888889	0.44410	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	5.55556	-1.01639	-1.71480	1.66229	-1.46115	-0.25365
3	1.03093	-0.37164	-0.03046	1.20421	-0.26475	0.23429
4	0.54054	-0.00300	0.61519	1.00150	0.41930	0.19589
5	0.49020	0.13103	0.71295	0.93659	0.66801	0.04494
6	0.26667	0.44212	1.32176	0.80167	1.24527	0.07648
7	0.22075	0.61896	1.51072	0.73383	1.57343	-0.06270
8	0.21464	0.72707	1.53878	0.69522	1.77402	-0.23525

COMBINED LANGMUIR REGRESSION on TEMPERATURE=55

39

18:38 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Sum of	Mean
--------	------

Source	DF	Squares	Square	F Value	Pr > F
Model	1	19.28568	19.28568	27.93	0.0032
Error	5	3.45251	0.69050		
Corrected Total	6	22.73820			

Root MSE	0.83096	R-Square	0.8482
Dependent Mean	1.18847	Adj R-Sq	0.8178
Coeff Var	69.91895		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-4.13563	1.05524	-3.92	0.0112
CMBLNGE	1	5.29739	1.00237	5.28	0.0032

COMBINED LANGMUIR REGRESSION on TEMPERATURE=55

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Obs	Temp	E	A	EbyA	EinV
1	55	0.001	0.001	.	.
2	55	0.3619	0.18	2.01056	2.76319
3	55	0.6896	0.97	0.71093	1.45012
4	55	0.997	1.85	0.53892	1.00301
5	55	1.14	2.04	0.55882	0.87719
6	55	1.556	3.75	0.41493	0.64267
7	55	1.857	4.53	0.40993	0.53850
8	55	2.069	4.658888889	0.44410	0.48333

Obs	Ainv	LNE	LNA	CMBLNGE	PRED	RESID
1
2	5.55556	-1.01639	-1.71480	1.66229	4.67014	0.88541
3	1.03093	-0.37164	-0.03046	1.20421	2.24352	-1.21260
4	0.54054	-0.00300	0.61519	1.00150	1.16972	-0.62918
5	0.49020	0.13103	0.71295	0.93659	0.82583	-0.33563
6	0.26667	0.44212	1.32176	0.80167	0.11112	0.15554
7	0.22075	0.61896	1.51072	0.73383	-0.24826	0.46901
8	0.21464	0.72707	1.53878	0.69522	-0.45280	0.66744

Appendix D

Statistical Analysis for Average values of Experimental
Adsorption Data of CBH I, CBH II and EG II to fit
Adsorption Models

SAS CODE

```
%let path=d:\Dole Files\;
libname bus "&path.";
/* THIS SAS FILE IS FOR REGRESSION FOR DATA ON AVERAGE FILE */
/* THIS DATASET CONTAINS AVERAGE OF ALL THREE FILES CBHIa CBHIIa and
EGIIa */

/* This step makes new variables for calculating different models */

data temp;
set bus.average;
m=2;
EbyA=E/A;
Einv=1/E;
Ainv=1/A;
LNE=log(E);
LNA=log(A);
CMBLNGE=1/(E**(1/m));
drop m;
run;

/*****
/* regression model...For AVERAGE OF FILE CBHI */
*****/

/* Selecting Cases */

data tempCBHI;
set temp;
if label='CBHI' then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on CBHI AVERAGES';
PROC REG data=tempCBHI;
  MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on CBHI AVERAGES';
PROC REG data=tempCBHI;
  MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on CBHI AVERAGES';
PROC REG data=tempCBHI;
```

```

MODEL Ainv=Einv;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on CBHI AVERAGES';
PROC REG data=tempCBHI;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on CBHI AVERAGES';
PROC REG data=tempCBHI;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

/*****
/* regression model....FOR AVERAGES OF FILE CBHII */
*****/

/* Selecting Cases */

data tempCBHII;
set temp;
if label='CBHII' then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on AVERAGES OF FILE CBHII';
PROC REG data=tempCBHII;
MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on AVERAGES OF FILE CBHII';
PROC REG data=tempCBHII;
MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;
PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

```

```

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on AVERAGES OF FILE CBHII';
PROC REG data=tempCBHII;
  MODEL Ainv=Einv;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on AVERAGES OF FILE CBHII';
PROC REG data=tempCBHII;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on AVERAGES OF FILE CBHII';
PROC REG data=tempCBHII;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

/*****
/* regression model....FOR AVERAGES OF FILE EGII */
*****/

/* Selecting Cases */

data tempEGII;
set temp;
if label='EGII' then output;
run;

/* A versus E Regression */
title 'A versus E REGRESSION on AVERAGES OF FILE EGII';
PROC REG data=tempEGII;
  MODEL A=E;
OUTPUT OUT=AE P=PRED R=RESID;
PLOT A*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=AE;
run;

/* EbyA versus E Regression */
title 'EbyA versus E REGRESSION on AVERAGES OF FILE EGII';
PROC REG data=tempEGII;
  MODEL EbyA=E;
OUTPUT OUT=EbyA P=PRED R=RESID;

```

```

PLOT EbyA*E='*' PREDICTED.*E='o' RESIDUAL.*E='x';
PROC PRINT DATA=EbyA;
run;

```

```

/* Langmuir Regression */
title 'LANGMUIR REGRESSION on AVERAGES OF FILE EGII';
PROC REG data=tempEGII;
MODEL Ainv=Einv;
OUTPUT OUT=LANGMUIR P=PRED R=RESID;
PLOT Ainv*Einv='*' PREDICTED.*Einv='o' RESIDUAL.*Einv='x';
PROC PRINT DATA=LANGMUIR;
run;

```

```

/* Freundlich Regression */
title 'FREUNDLICH REGRESSION on AVERAGES OF FILE EGII';
PROC REG data=tempEGII;
MODEL LNA=LNE;
OUTPUT OUT=FREUNDLICH P=PRED R=RESID;
PLOT LNA*LNE='*' PREDICTED.*LNE='o' RESIDUAL.*LNE='x';
PROC PRINT DATA=FREUNDLICH;
run;

```

```

/* Combined Langmuir Regression */
title 'COMBINED LANGMUIR REGRESSION on AVERAGES OF FILE EGII';
PROC REG data=tempEGII;
MODEL Ainv=CMBLNGE;
OUTPUT OUT=COMBLANGMUIR P=PRED R=RESID;
PLOT Ainv*CMBLNGE='*' PREDICTED.*CMBLNGE='o' RESIDUAL.*CMBLNGE='x';
PROC PRINT DATA=COMBLANGMUIR;
run;

```

SAS OUTPUT

A versus E REGRESSION on CBHI AVERAGES

1

18:14 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	5.69690	5.69690	330.67	<.0001
Error	5	0.08614	0.01723		
Corrected Total	6	5.78304			

Root MSE	0.13126	R-Square	0.9851
Dependent Mean	1.65143	Adj R-Sq	0.9821
Coeff Var	7.94806		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.21241	0.11387	-1.87	0.1211
E	1	1.43752	0.07905	18.18	<.0001

A versus E REGRESSION on CBHI AVERAGES

2

18:14 Saturday, October

25, 2003

Obs	Label	E	A	EbyA	Einvs	Ainvs
1	CBHI	0.493	0.515	0.95728	2.02840	1.94175
2	CBHI	0.737	0.703	1.04836	1.35685	1.42248
3	CBHI	0.976	1.168	0.83562	1.02459	0.85616
4	CBHI	1.159	1.45	0.79931	0.86281	0.68966
5	CBHI	1.412	2.057	0.68644	0.70822	0.48614
6	CBHI	1.827	2.41	0.75809	0.54735	0.41494
7	CBHI	2.472	3.257	0.75898	0.40453	0.30703

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.70725	-0.66359	1.42422	0.49628	0.01872
2	-0.30517	-0.35240	1.16484	0.84704	-0.14404
3	-0.02429	0.15529	1.01222	1.19060	-0.02260
4	0.14756	0.37156	0.92888	1.45367	-0.00367
5	0.34501	0.72125	0.84156	1.81736	0.23964
6	0.60268	0.87963	0.73983	2.41393	-0.00393
7	0.90503	1.18081	0.63603	3.34113	-0.08413

EbyA versus E REGRESSION on CBHI AVERAGES

3

18:14 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.04707	0.04707	4.86	0.0787
Error	5	0.04844	0.00969		
Corrected Total	6	0.09552			

Root MSE	0.09843	R-Square	0.4928
Dependent Mean	0.83487	Adj R-Sq	0.3914
Coeff Var	11.78996		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.00429	0.08539	11.76	<.0001
E	1	-0.13067	0.05928	-2.20	0.0787

EbyA versus E REGRESSION on CBHI AVERAGES

4

18:14 Saturday, October

25, 2003

Obs	Label	E	A	EbyA	Einvs	Ainvs
1	CBHI	0.493	0.515	0.95728	2.02840	1.94175
2	CBHI	0.737	0.703	1.04836	1.35685	1.42248
3	CBHI	0.976	1.168	0.83562	1.02459	0.85616
4	CBHI	1.159	1.45	0.79931	0.86281	0.68966
5	CBHI	1.412	2.057	0.68644	0.70822	0.48614
6	CBHI	1.827	2.41	0.75809	0.54735	0.41494
7	CBHI	2.472	3.257	0.75898	0.40453	0.30703

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.70725	-0.66359	1.42422	0.93987	0.01741
2	-0.30517	-0.35240	1.16484	0.90799	0.14038
3	-0.02429	0.15529	1.01222	0.87676	-0.04114
4	0.14756	0.37156	0.92888	0.85285	-0.05353
5	0.34501	0.72125	0.84156	0.81979	-0.13335
6	0.60268	0.87963	0.73983	0.76556	-0.00747
7	0.90503	1.18081	0.63603	0.68127	0.07771

LANGMUIR REGRESSION on CBHI AVERAGES

5

18:14 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainvs

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.11510	2.11510	247.45	<.0001
Error	5	0.04274	0.00855		
Corrected Total	6	2.15783			

Root MSE	0.09245	R-Square	0.9802
Dependent Mean	0.87402	Adj R-Sq	0.9762
Coeff Var	10.57794		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.18543	0.07588	-2.44	0.0584
Einvs	1	1.06973	0.06800	15.73	<.0001

LANGMUIR REGRESSION on CBHI AVERAGES

6

18:14 Saturday, October

25, 2003

Obs	Label	E	A	EbyA	Einvs	Ainvs
1	CBHI	0.493	0.515	0.95728	2.02840	1.94175
2	CBHI	0.737	0.703	1.04836	1.35685	1.42248
3	CBHI	0.976	1.168	0.83562	1.02459	0.85616
4	CBHI	1.159	1.45	0.79931	0.86281	0.68966
5	CBHI	1.412	2.057	0.68644	0.70822	0.48614
6	CBHI	1.827	2.41	0.75809	0.54735	0.41494
7	CBHI	2.472	3.257	0.75898	0.40453	0.30703

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.70725	-0.66359	1.42422	1.98441	-0.04266
2	-0.30517	-0.35240	1.16484	1.26604	0.15644
3	-0.02429	0.15529	1.01222	0.91061	-0.05444
4	0.14756	0.37156	0.92888	0.73755	-0.04789
5	0.34501	0.72125	0.84156	0.57217	-0.08602
6	0.60268	0.87963	0.73983	0.40008	0.01486
7	0.90503	1.18081	0.63603	0.24731	0.05972

FREUNDLICH REGRESSION on CBHI AVERAGES

7

18:14 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.61462	2.61462	264.08	<.0001
Error	5	0.04950	0.00990		
Corrected Total	6	2.66413			

Root MSE	0.09950	R-Square	0.9814
Dependent Mean	0.32751	Adj R-Sq	0.9777
Coeff Var	30.38189		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.16088	0.03898	4.13	0.0091
LNE	1	1.21049	0.07449	16.25	<.0001

FREUNDLICH REGRESSION on CBHI AVERAGES

8

18:14 Saturday, October

25, 2003

Obs	Label	E	A	EbyA	Einv	Ainv
1	CBHI	0.493	0.515	0.95728	2.02840	1.94175
2	CBHI	0.737	0.703	1.04836	1.35685	1.42248
3	CBHI	0.976	1.168	0.83562	1.02459	0.85616
4	CBHI	1.159	1.45	0.79931	0.86281	0.68966
5	CBHI	1.412	2.057	0.68644	0.70822	0.48614
6	CBHI	1.827	2.41	0.75809	0.54735	0.41494
7	CBHI	2.472	3.257	0.75898	0.40453	0.30703

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.70725	-0.66359	1.42422	-0.69523	0.03165
2	-0.30517	-0.35240	1.16484	-0.20852	-0.14388
3	-0.02429	0.15529	1.01222	0.13148	0.02382
4	0.14756	0.37156	0.92888	0.33950	0.03207
5	0.34501	0.72125	0.84156	0.57851	0.14274
6	0.60268	0.87963	0.73983	0.89041	-0.01079
7	0.90503	1.18081	0.63603	1.25641	-0.07560

COMBINED LANGMUIR REGRESSION on CBHI AVERAGES

9

18:14 Saturday, October

25, 2003

The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	2.07750	2.07750	129.30	<.0001
Error	5	0.08034	0.01607		
Corrected Total	6	2.15783			

Root MSE	0.12676	R-Square	0.9628
Dependent Mean	0.87402	Adj R-Sq	0.9553
Coeff Var	14.50268		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-1.24844	0.19271	-6.48	0.0013
CMBLNGE	1	2.20187	0.19364	11.37	<.0001

COMBINED LANGMUIR REGRESSION on CBHI AVERAGES

10

18:14 Saturday, October

25, 2003

Obs	Label	E	A	EbyA	Einv	Ainv
1	CBHI	0.493	0.515	0.95728	2.02840	1.94175
2	CBHI	0.737	0.703	1.04836	1.35685	1.42248
3	CBHI	0.976	1.168	0.83562	1.02459	0.85616
4	CBHI	1.159	1.45	0.79931	0.86281	0.68966
5	CBHI	1.412	2.057	0.68644	0.70822	0.48614
6	CBHI	1.827	2.41	0.75809	0.54735	0.41494
7	CBHI	2.472	3.257	0.75898	0.40453	0.30703

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.70725	-0.66359	1.42422	1.88750	0.05425

2	-0.30517	-0.35240	1.16484	1.31638	0.10609
3	-0.02429	0.15529	1.01222	0.98033	-0.12417
4	0.14756	0.37156	0.92888	0.79682	-0.10717
5	0.34501	0.72125	0.84156	0.60455	-0.11841
6	0.60268	0.87963	0.73983	0.38056	0.03438
7	0.90503	1.18081	0.63603	0.15201	0.15502

A versus E REGRESSION on AVERAGES OF FILE CBHII

11

18:14 Saturday, October

25, 2003

The REG Procedure
Model: MODEL1
Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	11.71216	11.71216	126.85	<.0001
Error	5	0.46165	0.09233		
Corrected Total	6	12.17380			

Root MSE	0.30386	R-Square	0.9621
Dependent Mean	2.62386	Adj R-Sq	0.9545
Coeff Var	11.58057		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.89267	0.33268	-2.68	0.0436
E	1	3.07389	0.27292	11.26	<.0001

A versus E REGRESSION on AVERAGES OF FILE CBHII

12

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25, 2003

Obs	Label	E	A	EbyA	EinV	AinV
1	CBHII	0.477	0.575	0.82957	2.09644	1.73913
2	CBHII	0.715	1.073	0.66636	1.39860	0.93197
3	CBHII	0.929	2.135	0.43513	1.07643	0.46838
4	CBHII	1.315	2.828	0.46499	0.76046	0.35361
5	CBHII	1.335	3.646	0.36615	0.74906	0.27427
6	CBHII	1.463	3.782	0.38683	0.68353	0.26441
7	CBHII	1.774	4.328	0.40989	0.56370	0.23105

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.74024	-0.55339	1.44791	0.57357	0.00143
2	-0.33547	0.07046	1.18262	1.30516	-0.23216
3	-0.07365	0.75847	1.03751	1.96297	0.17203
4	0.27384	1.03957	0.87204	3.14949	-0.32149
5	0.28893	1.29363	0.86548	3.21097	0.43503
6	0.38049	1.33025	0.82676	3.60443	0.17757
7	0.57324	1.46511	0.75080	4.56041	-0.23241

EbyA versus E REGRESSION on AVERAGES OF FILE CBHII

13

18:14 Saturday, October

25, 2003

The REG Procedure
Model: MODEL1

Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.13015	0.13015	13.04	0.0154
Error	5	0.04991	0.00998		
Corrected Total	6	0.18007			

Root MSE	0.09991	R-Square	0.7228
Dependent Mean	0.50842	Adj R-Sq	0.6674
Coeff Var	19.65170		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.87912	0.10939	8.04	0.0005
E	1	-0.32404	0.08974	-3.61	0.0154

EbyA versus E REGRESSION on AVERAGES OF FILE CBHII

14

18:14 Saturday, October

25, 2003

Obs	Label	E	A	EbyA	Einv	Ainv
1	CBHII	0.477	0.575	0.82957	2.09644	1.73913
2	CBHII	0.715	1.073	0.66636	1.39860	0.93197
3	CBHII	0.929	2.135	0.43513	1.07643	0.46838
4	CBHII	1.315	2.828	0.46499	0.76046	0.35361
5	CBHII	1.335	3.646	0.36615	0.74906	0.27427
6	CBHII	1.463	3.782	0.38683	0.68353	0.26441
7	CBHII	1.774	4.328	0.40989	0.56370	0.23105

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.74024	-0.55339	1.44791	0.72455	0.10501
2	-0.33547	0.07046	1.18262	0.64743	0.01893
3	-0.07365	0.75847	1.03751	0.57809	-0.14296
4	0.27384	1.03957	0.87204	0.45301	0.01199
5	0.28893	1.29363	0.86548	0.44653	-0.08037
6	0.38049	1.33025	0.82676	0.40505	-0.01822
7	0.57324	1.46511	0.75080	0.30427	0.10562

LANGMUIR REGRESSION on AVERAGES OF FILE CBHII

15

18:14 Saturday, October

25, 2003

The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	1.79021	1.79021	179.32	<.0001
Error	5	0.04992	0.00998		
Corrected Total	6	1.84013			

Root MSE	0.09992	R-Square	0.9729
Dependent Mean	0.60897	Adj R-Sq	0.9674

Coeff Var 16.40741

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.44615	0.08738	-5.11	0.0038
Einvs	1	1.00787	0.07526	13.39	<.0001

LANGMUIR REGRESSION on AVERAGES OF FILE CBHII

16

18:14 Saturday, October

25, 2003

Obs	Label	E	A	EbyA	Einvs	Ainvs
1	CBHII	0.477	0.575	0.82957	2.09644	1.73913
2	CBHII	0.715	1.073	0.66636	1.39860	0.93197
3	CBHII	0.929	2.135	0.43513	1.07643	0.46838
4	CBHII	1.315	2.828	0.46499	0.76046	0.35361
5	CBHII	1.335	3.646	0.36615	0.74906	0.27427
6	CBHII	1.463	3.782	0.38683	0.68353	0.26441
7	CBHII	1.774	4.328	0.40989	0.56370	0.23105

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.74024	-0.55339	1.44791	1.66678	0.07235
2	-0.33547	0.07046	1.18262	0.96346	-0.03149
3	-0.07365	0.75847	1.03751	0.63875	-0.17036
4	0.27384	1.03957	0.87204	0.32029	0.03332
5	0.28893	1.29363	0.86548	0.30881	-0.03454
6	0.38049	1.33025	0.82676	0.24276	0.02165
7	0.57324	1.46511	0.75080	0.12198	0.10907

FREUNDLICH REGRESSION on AVERAGES OF FILE CBHII

17

18:14 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	3.29073	3.29073	175.06	<.0001
Error	5	0.09399	0.01880		
Corrected Total	6	3.38472			

Root MSE 0.13711 R-Square 0.9722
 Dependent Mean 0.77201 Adj R-Sq 0.9667
 Coeff Var 17.75953

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.68787	0.05221	13.18	<.0001
LNE	1	1.60434	0.12126	13.23	<.0001

FREUNDLICH REGRESSION on AVERAGES OF FILE CBHII

18

18:14 Saturday, October

25, 2003

Obs	Label	E	A	EbyA	Einv	Ainv
1	CBHII	0.477	0.575	0.82957	2.09644	1.73913
2	CBHII	0.715	1.073	0.66636	1.39860	0.93197
3	CBHII	0.929	2.135	0.43513	1.07643	0.46838
4	CBHII	1.315	2.828	0.46499	0.76046	0.35361
5	CBHII	1.335	3.646	0.36615	0.74906	0.27427
6	CBHII	1.463	3.782	0.38683	0.68353	0.26441
7	CBHII	1.774	4.328	0.40989	0.56370	0.23105

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-0.74024	-0.55339	1.44791	-0.49972	-0.05366
2	-0.33547	0.07046	1.18262	0.14966	-0.07920
3	-0.07365	0.75847	1.03751	0.56972	0.18875
4	0.27384	1.03957	0.87204	1.12720	-0.08763
5	0.28893	1.29363	0.86548	1.15141	0.14222
6	0.38049	1.33025	0.82676	1.29830	0.03195
7	0.57324	1.46511	0.75080	1.60753	-0.14243

COMBINED LANGMUIR REGRESSION on AVERAGES OF FILE CBHII

19

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25, 2003

The REG Procedure
Model: MODEL1
Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	1.73091	1.73091	79.24	0.0003
Error	5	0.10922	0.02184		
Corrected Total	6	1.84013			

Root MSE	0.14779	R-Square	0.9406
Dependent Mean	0.60897	Adj R-Sq	0.9288
Coeff Var	24.26932		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-1.57266	0.25136	-6.26	0.0015
CMBLNGE	1	2.18691	0.24567	8.90	0.0003

COMBINED LANGMUIR REGRESSION on AVERAGES OF FILE CBHII

20

18:14 Saturday, October

25, 2003

Obs	Label	E	A	EbyA	Einv	Ainv
1	CBHII	0.477	0.575	0.82957	2.09644	1.73913
2	CBHII	0.715	1.073	0.66636	1.39860	0.93197
3	CBHII	0.929	2.135	0.43513	1.07643	0.46838
4	CBHII	1.315	2.828	0.46499	0.76046	0.35361
5	CBHII	1.335	3.646	0.36615	0.74906	0.27427
6	CBHII	1.463	3.782	0.38683	0.68353	0.26441
7	CBHII	1.774	4.328	0.40989	0.56370	0.23105

Obs	LNE	LNA	CMBLNGE	PRED	RESID
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1	-0.74024	-0.55339	1.44791	1.59378	0.14535
2	-0.33547	0.07046	1.18262	1.01363	-0.08166
3	-0.07365	0.75847	1.03751	0.69628	-0.22789
4	0.27384	1.03957	0.87204	0.33441	0.01919
5	0.28893	1.29363	0.86548	0.32008	-0.04580
6	0.38049	1.33025	0.82676	0.23538	0.02903
7	0.57324	1.46511	0.75080	0.06927	0.16179

A versus E REGRESSION on AVERAGES OF FILE EGII

21

18:14 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: A

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	10.67175	10.67175	159.84	<.0001
Error	5	0.33382	0.06676		
Corrected Total	6	11.00557			

Root MSE	0.25839	R-Square	0.9697
Dependent Mean	2.22086	Adj R-Sq	0.9636
Coeff Var	11.63465		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.44148	0.23213	-1.90	0.1156
E	1	2.14928	0.17000	12.64	<.0001

A versus E REGRESSION on AVERAGES OF FILE EGII

22

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25, 2003

Obs	Label	E	A	EbyA	EinV	Ainv
1	EGII	0.362	0.178	2.03371	2.76243	5.61798
2	EGII	0.69	1.07	0.64486	1.44928	0.93458
3	EGII	0.997	1.928	0.51712	1.00301	0.51867
4	EGII	1.14	1.82	0.62637	0.87719	0.54945
5	EGII	1.556	3.135	0.49633	0.64267	0.31898
6	EGII	1.857	3.76	0.49388	0.53850	0.26596
7	EGII	2.069	3.655	0.56607	0.48333	0.27360

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-1.01611	-1.72597	1.66206	0.33656	-0.15856
2	-0.37106	0.06766	1.20386	1.04152	0.02848
3	-0.00300	0.65648	1.00150	1.70135	0.22665
4	0.13103	0.59884	0.93659	2.00869	-0.18869
5	0.44212	1.14263	0.80167	2.90279	0.23221
6	0.61896	1.32442	0.73383	3.54972	0.21028
7	0.72707	1.29610	0.69522	4.00537	-0.35037

EbyA versus E REGRESSION on AVERAGES OF FILE EGII

23

18:14 Saturday, October

25, 2003

The REG Procedure

Model: MODEL1
 Dependent Variable: EbyA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	0.83730	0.83730	3.98	0.1027
Error	5	1.05259	0.21052		
Corrected Total	6	1.88990			

Root MSE	0.45882	R-Square	0.4430
Dependent Mean	0.76834	Adj R-Sq	0.3317
Coeff Var	59.71653		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	1.51408	0.41219	3.67	0.0144
E	1	-0.60203	0.30187	-1.99	0.1027

EbyA versus E REGRESSION on AVERAGES OF FILE EGII

24

18:14 Saturday, October

25, 2003

Obs	Label	E	A	EbyA	Einv	Ainv
1	EGII	0.362	0.178	2.03371	2.76243	5.61798
2	EGII	0.69	1.07	0.64486	1.44928	0.93458
3	EGII	0.997	1.928	0.51712	1.00301	0.51867
4	EGII	1.14	1.82	0.62637	0.87719	0.54945
5	EGII	1.556	3.135	0.49633	0.64267	0.31898
6	EGII	1.857	3.76	0.49388	0.53850	0.26596
7	EGII	2.069	3.655	0.56607	0.48333	0.27360

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-1.01611	-1.72597	1.66206	1.29614	0.73757
2	-0.37106	0.06766	1.20386	1.09868	-0.45382
3	-0.00300	0.65648	1.00150	0.91385	-0.39674
4	0.13103	0.59884	0.93659	0.82776	-0.20139
5	0.44212	1.14263	0.80167	0.57732	-0.08099
6	0.61896	1.32442	0.73383	0.39611	0.09777
7	0.72707	1.29610	0.69522	0.26848	0.29759

LANGMUIR REGRESSION on AVERAGES OF FILE EGII

25

18:14 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainv

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	20.86224	20.86224	49.20	0.0009
Error	5	2.12011	0.42402		
Corrected Total	6	22.98235			

Root MSE	0.65117	R-Square	0.9078
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Dependent Mean 1.21132 Adj R-Sq 0.8893
 Coeff Var 53.75724

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-1.36839	0.44253	-3.09	0.0271
Einvs	1	2.32813	0.33191	7.01	0.0009

LANGMUIR REGRESSION on AVERAGES OF FILE EGII

26

18:14 Saturday, October

25, 2003

Obs	Label	E	A	EbyA	Einvs	Ainvs
1	EGII	0.362	0.178	2.03371	2.76243	5.61798
2	EGII	0.69	1.07	0.64486	1.44928	0.93458
3	EGII	0.997	1.928	0.51712	1.00301	0.51867
4	EGII	1.14	1.82	0.62637	0.87719	0.54945
5	EGII	1.556	3.135	0.49633	0.64267	0.31898
6	EGII	1.857	3.76	0.49388	0.53850	0.26596
7	EGII	2.069	3.655	0.56607	0.48333	0.27360

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-1.01611	-1.72597	1.66206	5.06291	0.55506
2	-0.37106	0.06766	1.20386	2.00571	-1.07113
3	-0.00300	0.65648	1.00150	0.96675	-0.44807
4	0.13103	0.59884	0.93659	0.67383	-0.12438
5	0.44212	1.14263	0.80167	0.12784	0.19114
6	0.61896	1.32442	0.73383	-0.11468	0.38064
7	0.72707	1.29610	0.69522	-0.24315	0.51674

FREUNDLICH REGRESSION on AVERAGES OF FILE EGII

27

18:14 Saturday, October

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: LNA

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	6.46735	6.46735	74.79	0.0003
Error	5	0.43239	0.08648		
Corrected Total	6	6.89974			

Root MSE 0.29407 R-Square 0.9373
 Dependent Mean 0.48002 Adj R-Sq 0.9248
 Coeff Var 61.26222

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	0.35203	0.11213	3.14	0.0257
LNE	1	1.69368	0.19585	8.65	0.0003

FREUNDLICH REGRESSION on AVERAGES OF FILE EGII

28

25, 2003

Obs	Label	E	A	EbyA	Einvs	Ainvs
1	EGII	0.362	0.178	2.03371	2.76243	5.61798
2	EGII	0.69	1.07	0.64486	1.44928	0.93458
3	EGII	0.997	1.928	0.51712	1.00301	0.51867
4	EGII	1.14	1.82	0.62637	0.87719	0.54945
5	EGII	1.556	3.135	0.49633	0.64267	0.31898
6	EGII	1.857	3.76	0.49388	0.53850	0.26596
7	EGII	2.069	3.655	0.56607	0.48333	0.27360

Obs	LNE	LNA	CMBLNGE	PRED	RESID
1	-1.01611	-1.72597	1.66206	-1.36893	-0.35704
2	-0.37106	0.06766	1.20386	-0.27643	0.34409
3	-0.00300	0.65648	1.00150	0.34694	0.30954
4	0.13103	0.59884	0.93659	0.57395	0.02489
5	0.44212	1.14263	0.80167	1.10083	0.04179
6	0.61896	1.32442	0.73383	1.40035	-0.07593
7	0.72707	1.29610	0.69522	1.58344	-0.28735

COMBINED LANGMUIR REGRESSION on AVERAGES OF FILE EGII

29

25, 2003

The REG Procedure
 Model: MODEL1
 Dependent Variable: Ainvs

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	19.04212	19.04212	24.16	0.0044
Error	5	3.94024	0.78805		
Corrected Total	6	22.98235			

Root MSE	0.88772	R-Square	0.8286
Dependent Mean	1.21132	Adj R-Sq	0.7943
Coeff Var	73.28558		

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-4.08032	1.12756	-3.62	0.0152
CMBLNGE	1	5.26552	1.07117	4.92	0.0044

COMBINED LANGMUIR REGRESSION on AVERAGES OF FILE EGII

30

25, 2003

Obs	Label	E	A	EbyA	Einvs	Ainvs
1	EGII	0.362	0.178	2.03371	2.76243	5.61798
2	EGII	0.69	1.07	0.64486	1.44928	0.93458
3	EGII	0.997	1.928	0.51712	1.00301	0.51867
4	EGII	1.14	1.82	0.62637	0.87719	0.54945
5	EGII	1.556	3.135	0.49633	0.64267	0.31898
6	EGII	1.857	3.76	0.49388	0.53850	0.26596
7	EGII	2.069	3.655	0.56607	0.48333	0.27360

Obs	LNE	LNA	CMBLNGE	PRED	RESID
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1	-1.01611	-1.72597	1.66206	4.67127	0.94671
2	-0.37106	0.06766	1.20386	2.25862	-1.32404
3	-0.00300	0.65648	1.00150	1.19312	-0.67445
4	0.13103	0.59884	0.93659	0.85129	-0.30184
5	0.44212	1.14263	0.80167	0.14089	0.17809
6	0.61896	1.32442	0.73383	-0.21633	0.48229
7	0.72707	1.29610	0.69522	-0.41964	0.69324